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JC600UTILITY
PATENT APPLICATION
TRANSMITTAL

(Only for new nonprovisional applications under 37 C.F.R. § 1.53(b))

Attorney Docket No. MB 1-0010

First Inventor or Application Identifier Mecard

Title Disease-Induced Polymorphisms

Express Mail Label No. EK 483897023 US

APPLICATION ELEMENTS

See MPEP chapter 600 concerning utility patent application contents.

1. * Fee Transmittal Form (e.g., PTO/SB/17)
(Submit an original and a duplicate for fee processing)

2. Specification [Total Pages 29]
(preferred arrangement set forth below)

- Descriptive title of the Invention
- Cross References to Related Applications
- Statement Regarding Fed sponsored R & D
- Reference to Microfiche Appendix
- Background of the Invention
- Brief Summary of the Invention
- Brief Description of the Drawings (if filed)
- Detailed Description
- Claim(s)
- Abstract of the Disclosure

3. Drawing(s) (35 U.S.C. 113) [Total Sheets 4]

4. Oath or Declaration [Total Pages]

- a. Newly executed (original or copy)
- b. Copy from a prior application (37 C.F.R. § 1.63(d))
(for continuation/divisional with Box 16 completed)
 - i. DELETION OF INVENTOR(S)
Signed statement attached deleting
inventor(s) named in the prior application,
see 37 C.F.R. §§ 1.63(d)(2) and 1.33(b).

NOTE FOR ITEMS 1 & 13: IN ORDER TO BE ENTITLED TO PAY SMALL ENTITY FEES, A SMALL ENTITY STATEMENT IS REQUIRED (37 C.F.R. § 1.27), EXCEPT IF ONE FILED IN A PRIOR APPLICATION IS RELIED UPON (37 C.F.R. § 1.28).

ADDRESS TO: Assistant Commissioner for Patents
Box Patent Application
Washington, DC 20231

5. Microfiche Computer Program (Appendix)

6. Nucleotide and/or Amino Acid Sequence Submission
(if applicable, all necessary)

- a. Computer Readable Copy
- b. Paper Copy (identical to computer copy)
- c. Statement verifying identity of above copies

ACCOMPANYING APPLICATION PARTS

7. Assignment Papers (cover sheet & document(s))

8. 37 C.F.R. §3.73(b) Statement Power of
(when there is an assignee) Attorney

9. English Translation Document (if applicable)

10. Information Disclosure Statement (IDS)/PTO-1449 Copies of IDS
Statement (IDS)/PTO-1449 Citations

11. Preliminary Amendment

12. Return Receipt Postcard (MPEP 503)
(Should be specifically itemized)

13. * Small Entity Statement(s) Statement filed in prior application,
(PTO/SB/09-12) Status still proper and desired

14. Certified Copy of Priority Document(s)
(if foreign priority is claimed)

15. Other:

16. If a CONTINUING APPLICATION, check appropriate box, and supply the requisite information below and in a preliminary amendment:
 Continuation Divisional Continuation-in-part (CIP) of prior application No. J
 Prior application information: Examiner _____ Group / Art Unit: _____

For CONTINUATION or DIVISIONAL APPS only: The entire disclosure of the prior application, from which an oath or declaration is supplied under Box 4b, is considered a part of the disclosure of the accompanying continuation or divisional application and is hereby incorporated by reference. The incorporation can only be relied upon when a portion has been inadvertently omitted from the submitted application parts.

17. CORRESPONDENCE ADDRESS

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Signature	Karen Guevres	Date	3/22/00

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1675 U.S. PTO
03/22/00

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application

Inventor(s): Jacqueline Heard et al.

Application No.: Unassigned

Filed: Herewith

Title: Disease-induced polynucleotides

VERIFIED STATEMENT CLAIMING SMALL ENTITY STATUS
37 C.F.R. § 1.9(d) AND 1.27(c) - SMALL BUSINESS CONCERN

I hereby declare that I am an official of the small business concern empowered to act on behalf of the concern identified below.

Name: Mendel Biotechnology, Inc.

Address: 21375 Cabot Boulevard, Hayward, California 94545

I hereby declare that the above identified small business concern qualifies as a small business concern as defined in 13 C.F.R. § 121.12, and reproduced in 37 C.F.R. § 1.9(d), for purposes of paying reduced fees under Section 41(a) and (b) of Title 35 U.S.C. in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third-party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the invention.

entitled: Disease-induced polynucleotides

described in the Specification filed herewith

If the rights held by the above-identified small business concern are not exclusive, each individual, concern or organization having rights to the invention is listed below and no rights to the invention are held by any person, other than the inventor, who could not qualify as a small business concern under 37 C.F.R. § 1.9(d) or by any concern which would not qualify as a small business concern under 37 C.F.R. § 1.9(d) or a nonprofit organization under 37 C.F.R. § 1.9(e).

Individual Small Business Concern Nonprofit Organization

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small business entity is no longer appropriate. (37 C.F.R. § 1.28(b)).

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

Name of Person Signing: Guo-Liang Yu
Title of Person Signing: Senior Vice-President, Research and Development
Address of Person Signing: 21375 Cabot Boulevard, Hayward, California 94545

Signature: 
Date: 03/20/2006

DISEASE-INDUCED POLYNUCLEOTIDES

The present invention claims priority in part from US Provisional Application Serial No. 60/125,814 filed March 23, 1999.

5

FIELD OF THE INVENTION

This invention is in the field of plant molecular biology and relates to compositions and methods for modifying a plant's traits, in particular plant disease tolerance or resistance.

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BACKGROUND OF THE INVENTION

Gene expression levels are controlled in part at the level of transcription, and transcription is affected by transcription factors. Transcription factors regulate gene expression throughout the life cycle of an organism and so are responsible for differential levels of gene expression at various developmental stages, in different tissue and cell types, and in response to different stimuli. Transcription factors may interact with other proteins or with specific sites on a target gene sequence to activate, suppress or otherwise regulate transcription. In addition, the transcription of the transcription factors themselves may be regulated.

Because transcription factors are key controlling elements for biological pathways, altering the expression levels of one or more transcription factors may change entire biological pathways in an organism. For example, manipulation of the levels of selected transcription factors may result in increased expression of economically useful proteins or metabolic chemicals in plants or to improve other agriculturally relevant characteristics. Conversely, blocked or reduced expression of a transcription factor may reduce biosynthesis of unwanted compounds or remove an undesirable trait. Therefore, manipulating transcription factor levels in a plant offers tremendous potential in agricultural biotechnology for modifying a plant's traits.

The present invention provides transcription factors for use in modifying a plant's disease tolerance or resistance.

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SUMMARY OF THE INVENTION

In one aspect, the present invention relates to a transgenic plant comprising a recombinant polynucleotide. The recombinant polynucleotide comprises a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of protein SEQ ID Nos. 2N, where N=1-56. And the presence of the recombinant polynucleotide alters the disease tolerance or resistance

of the transgenic plant when compared with the same trait of another plant lacking the recombinant polynucleotide.

In one embodiment, the nucleotide sequence encodes a polypeptide comprising a conserved domain which may be 1) a localization domain, 2) an activation domain, 3) a repression domain, 4) an oligomerization domain or 5) a DNA binding domain. In a further embodiment, the nucleotide sequence further comprises a promoter operably linked to the nucleotide sequence. The promoter may be a constitutive or inducible or tissue-active.

In a second aspect, the present invention relates to a method for altering a plant's disease tolerance or resistance. The method comprises (a) transforming a plant with a recombinant polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of protein SEQ ID Nos. 2N, where N=1-56; (b) selecting transformed plants; and (c) identifying a transformed plant with roots having an altered trait.

In one embodiment, the nucleotide sequence encodes a polypeptide comprising a conserved domain which may be 1) a localization domain, 2) an activation domain, 3) a repression domain, 4) an oligomerization domain or 5) a DNA binding domain. In a further embodiment, the nucleotide sequence further comprises a promoter operably linked to the nucleotide sequence. The promoter may be a constitutive or inducible or tissue-active.

In a third aspect, the present invention relates to a method for altering the expression levels of at least one gene in a plant. The method comprises (a) transforming the plant with a recombinant polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of protein SEQ ID Nos. 2N, where N=1-56; and (b) selecting said transformed plant.

In one embodiment, the nucleotide sequence encodes a polypeptide comprising a conserved domain which may be 1) a localization domain, 2) an activation domain, 3) a repression domain, 4) an oligomerization domain or 5) a DNA binding domain. In a further embodiment, the nucleotide sequence further comprises a promoter operably linked to the nucleotide sequence. The promoter may be a constitutive or inducible or tissue-active.

In a fourth aspect, the present invention relates to another method for altering the disease tolerance of a plant. The method comprises (a) transforming the plant with a recombinant polynucleotide comprising a nucleotide sequence comprising at least 18 consecutive nucleotides of a sequence selected from the group consisting of SEQ ID Nos. 2N-1, where N= 1-56, and SEQ ID Nos. 113-121; and (b) selecting said transformed plant.

In yet another aspect, the present invention is yet another method for altering a plant's trait. The method comprises (a) providing a database sequence; (b) comparing the database sequence with a polypeptide selected from SEQ ID Nos. 2N, where N= 1-56; (c) selecting a database sequence that meets selected sequence criteria; and (d) transforming

said database sequence in the plant. Alternatively, the database sequence can be compared with a polynucleotide selected from SEQ ID Nos. 2N-1, where N= 1-56 or SEQ ID Nos. 113-121.

5 In a further aspect, the present invention is method for altering a plant's trait, and the method entails (a) providing a test polynucleotide; (b) hybridizing the test polynucleotide with a polynucleotide selected from SEQ ID Nos. 2N-1, where N= 1-56 or SEQ ID Nos. 113-121 at low stringency; and (c) transforming the hybridizing test polynucleotide in a plant to alter a trait of the plant.

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BRIEF DESCRIPTION OF THE FIGURES

Figures 1a-1e provide a table of exemplary polynucleotide and polypeptide sequences of the invention. The table includes from left to right for each sequence: the SEQ ID No., the internal code reference number, the transcription factor family of the sequence, particular DNA or protein fragments for each sequence, whether the sequence is a polynucleotide or polypeptide sequence, identification of the coding sequence for each full length and identification of any conserved domains for the polypeptide sequences.

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DETAILED DESCRIPTION OF THE INVENTION

DEFINITIONS

A "recombinant polynucleotide" is a nucleotide sequence comprising a gene coding sequence or a fragment thereof (comprising at least 18 consecutive nucleotides, preferably at least 30 consecutive nucleotides, and more preferably at least 50 consecutive nucleotides). Additionally, the polynucleotide may comprise a promoter, an intron, an enhancer region, a polyadenylation site, a translation initiation site, 5' or 3' untranslated regions, a reporter gene, a selectable marker or the like. The polynucleotide may comprise single stranded or double stranded DNA or RNA. The polynucleotide may comprise modified bases or a modified backbone. The polynucleotide may be genomic, a transcript (such as an mRNA) or a processed nucleotide sequence (such as a cDNA). The polynucleotide may comprise a sequence in either sense or antisense orientations.

A "recombinant polynucleotide" is a polynucleotide that is not in its native state, e.g., the polynucleotide is comprised of a nucleotide sequence not found in nature or the polynucleotide is separated from nucleotide sequences with which it typically is in proximity or is next to nucleotide sequences with which it typically is not in proximity.

An "recombinant polypeptide" is a polypeptide derived from the translation of a recombinant polynucleotide or is more enriched in a cell than the polypeptide in its natural

state in a wild type cell, e.g. more than 5% enriched, more than 10% enriched or more than 20% enriched and is not the result of a natural response of a wild type plant or is separated from other components with which it is typically associated with in a cell.

5 A “transgenic plant” may refer to a plant that contains genetic material not normally found in a wild type plant of the same species, or in a naturally occurring variety or in a cultivar, and which has been introduced into the plant by human manipulation. A transgenic plant is a plant that may contain an expression vector or cassette. The expression cassette comprises a gene coding sequence and allows for the expression of the gene coding sequence. The expression cassette may be introduced into a plant by transformation or by 10 breeding after transformation of a parent plant.

A transgenic plant refers to a whole plant as well as to a plant part, such as seed, fruit, leaf, or root, plant tissue, plant cells or any other plant material, and progeny thereof.

15 The phrase “altered expression” in reference to polynucleotide or polypeptide expression refers to an expression pattern in the transgenic plant that is different from the expression pattern in the wild type plant or a reference; for example, by expression in a cell type other than a cell type in which the sequence is expressed in the wild type plant, or by expression at a time other than at the time the sequence is expressed in the wild type plant, or by a response to different inducible agents, such as hormones or environmental signals, or at different expression levels (either higher or lower) compared with those found in a wild type plant. The term also refers to lowering the levels of expression to below the detection level or 20 completely abolishing expression. The resulting expression pattern may be transient or stable, constitutive or inducible.

25 A “transcription factor” (TF) refers to a polynucleotide or polypeptide that controls the expression of a gene or genes either directly by binding to one or more nucleotide sequences associated with a gene coding sequence or indirectly by affecting the level or activity of other polypeptides that do bind directly or indirectly to one or more nucleotide sequences associated with a gene coding sequence. A TF, in this definition, includes any polypeptide that can activate or repress transcription of a single gene or a number of genes. This polypeptide group includes, but is not limited to, DNA binding proteins, protein kinases, protein 30 phosphatases, GTP-binding proteins and receptors.

35 The transcription factor sequence may comprise a whole coding sequence or a fragment or domain of a coding sequence. A “fragment or domain”, as referred to polypeptides, may be a portion of a polypeptide which performs at least one biological function of the intact polypeptide in substantially the same manner or to a similar extent as does the intact polypeptide. A fragment may comprise, for example, a DNA binding domain that binds to a specific DNA promoter region, an activation domain or a domain for protein-protein interactions. Fragments may vary in size from as few as 6 amino acids to the length of the

intact polypeptide, but are preferably at least 30 amino acids in length and more preferably at least 60 amino acids in length. In reference to a nucleotide sequence “a fragment” refers to any sequence of at least consecutive 18 nucleotides, preferably at least 30 nucleotides, more preferably at least 50, of any of the sequences provided herein. Exemplary polynucleotides or polypeptides comprise a sequence provided in the Sequence Listing as SEQ ID No.1 (G1043), SEQ ID No.2 (G1043 protein), SEQ ID No.3 (G759), SEQ ID No.4 (G759 protein), SEQ ID No.5 (G185), SEQ ID No.6 (G185 protein), SEQ ID No.7 (G629), SEQ ID No.8 (G629 protein), SEQ ID No.9 (G435), SEQ ID No.10 (G435 protein), SEQ ID No.11 (G4), SEQ ID No.12 (G4 protein), SEQ ID No.13 (G1035), SEQ ID No.14 (G1035 protein), SEQ ID No.15 (G179), SEQ ID No.16 (G179 protein), SEQ ID No.17 (G28), SEQ ID No.18 (G28 protein), SEQ ID No.19 (G1241), SEQ ID No.20 (G1241 protein), SEQ ID No.21 (G19), SEQ ID No.22 (G19 protein), SEQ ID No.23 (G503), SEQ ID No.24 (G503 protein), SEQ ID No.25 (G263), SEQ ID No.26 (G263 protein), SEQ ID No.27 (G921), SEQ ID No.28 (G921 protein), SEQ ID No.29 (G1275), SEQ ID No.30 (G1275 protein), SEQ ID No.31 (G242), SEQ ID No.32 (G242 protein), SEQ ID No.33 (G1006), SEQ ID No.34 (G1006 protein), SEQ ID No.35 (G1049), SEQ ID No.36 (G1049 protein), SEQ ID No.37 (G502), SEQ ID No.38 (G502 protein), SEQ ID No.39 (G239), SEQ ID No.40 (G239 protein), SEQ ID No.41 (G555), SEQ ID No.42 (G555 protein), SEQ ID No.43 (G352), SEQ ID No.44 (G352 protein), SEQ ID No.45 (G1352), SEQ ID No.46 (G1352 protein), SEQ ID No.47 (G1089), SEQ ID No.48 (G1089 protein), SEQ ID No.49 (G553), SEQ ID No.50 (G553 protein), SEQ ID No.51 (G1221), SEQ ID No.52 (G1221 protein), SEQ ID No.53 (G580), SEQ ID No.54 (G580 protein), SEQ ID No.55 (G270), SEQ ID No.56 (G270 protein), SEQ ID No.57 (G201), SEQ ID No.58 (G201 protein), SEQ ID No.59 (G1417), SEQ ID No.60 (G1417 protein), SEQ ID No.61 (G233), SEQ ID No.62 (G233 protein), SEQ ID No.63 (G920), SEQ ID No.64 (G920 protein), SEQ ID No.65 (G867), SEQ ID No.66 (G867 protein), SEQ ID No.67 (G659), SEQ ID No.68 (G659 protein), SEQ ID No.69 (G620), SEQ ID No.70 (G620 protein), SEQ ID No.71 (G596), SEQ ID No.72 (G596 protein), SEQ ID No.73 (G511), SEQ ID No.74 (G511 protein), SEQ ID No.75 (G471), SEQ ID No.76 (G471 protein), SEQ ID No.77 (G385), SEQ ID No.78 (G385 protein), SEQ ID No.79 (G261), SEQ ID No.80 (G261 protein), SEQ ID No.81 (G25), SEQ ID No.82 (G25 protein), SEQ ID No.83 (G610), SEQ ID No.84 (G610 protein), SEQ ID No.85 (G229), SEQ ID No.86 (G229 protein), SEQ ID No.87 (G221), SEQ ID No.88 (G221 protein), SEQ ID No.89 (G186), SEQ ID No.90 (G186 protein), SEQ ID No.91 (G562), SEQ ID No.92 (G562 protein), SEQ ID No.93 (G255), SEQ ID No.94 (G255 protein), SEQ ID No.95 (G3), SEQ ID No.96 (G3 protein), SEQ ID No.97 (G713), SEQ ID No.98 (G713 protein), SEQ ID No.99 (G515), SEQ ID No.100 (G515 protein), SEQ ID No.101 (G390), SEQ ID No.102 (G390 protein), SEQ ID No.103 (G1034), SEQ ID No.104 (G1034 protein), SEQ ID No.105 (G1149), SEQ ID No.106 (G1149 protein), SEQ ID No.107 (G1334), SEQ ID No.108 (G1334 protein), SEQ ID No.109 (G1650), SEQ ID

No.110 (G1650 protein), SEQ ID No.111 (G241), SEQ ID No.112 (G241 protein), SEQ ID No.113 (G348), SEQ ID No.114 (G171), SEQ ID No.115 (G521), SEQ ID No.116 (G1274), SEQ ID No.117 (G182), SEQ ID No.118 (G1290), SEQ ID No.119 (G374), SEQ ID No.120 (G682) and SEQ ID No.121 (G501).

5 A "conserved domain" refers to a polynucleotide or polypeptide fragment that is more conserved at a sequence level than other fragments when the polynucleotide or polypeptide is compared with homologous genes or proteins from other plants. The conserved domain may be 1) a localization domain, 2) an activation domain, 3) a repression domain, 4) an oligomerization domain or 5) a DNA binding domain.

10 A nucleotide sequence is "operably linked" when it is placed into a functional relationship with another nucleotide sequence. For example, a promoter or enhancer is operably linked to a gene coding sequence if the presence of the promoter or enhancer increases the level of expression of the gene coding sequence.

15 "Trait" refers to a physiological, morphological, biochemical or physical characteristic of a plant or particular plant material or cell. This characteristic may be visible to the human eye, such as seed or plant size, or be measured by biochemical techniques, such as the protein, starch or oil content of seed or leaves or by the observation of the expression level of genes by employing Northerns, RT PCR, microarray gene expression assays or reporter gene expression systems or be measured by agricultural observations such as stress tolerance, yield or disease resistance.

20 "Trait modification" refers to a detectable difference in a characteristic in a transgenic plant expressing a polynucleotide or polypeptide of the present invention relative to a plant not doing so, such as a wild type plant. The trait modification may entail at least a 5% increase or decrease in an observed trait (difference), at least a 10% difference, at least a 20% difference, at least a 30%, at least a 50%, at least a 70%, at least a 100% or a greater difference. It is known that there may be a natural variation in the modified trait. Therefore, the trait modification observed entails a change in the normal distribution of the trait in transgenic plants compared with the distribution observed in wild type plant.

25 Trait modifications of particular interest include those to seed (embryo), fruit, root, flower, leaf, stem, shoot, seedling or the like, including: enhanced tolerance to environmental conditions including freezing, chilling, heat, drought, water saturation, radiation and ozone; enhanced resistance to microbial, fungal or viral diseases; resistance to nematodes, decreased herbicide sensitivity, enhanced tolerance of heavy metals (or enhanced ability to take up heavy metals), enhanced growth under poor photoconditions (e.g., low light and/or short day length), or changes in expression levels of genes of interest. Other phenotypes that may be modified relate to the production of plant metabolites, such as variations in the production of taxol, tocopherol, tocotrienol, sterols, phytosterols, vitamins, wax monomers,

anti-oxidants, amino acids, lignins, cellulose, tannins, prenyllipids (such as chlorophylls and carotenoids), glucosinolates, and terpenoids, enhanced or compositionally altered protein or oil production (especially in seeds), or modified sugar (insoluble or soluble) and/or starch composition. Physical plant characteristics that may be modified include cell development (such as the number of trichomes), fruit and seed size and number, yields of plant parts such as stems, leaves and roots, the stability of the seeds during storage, characteristics of the seed pod (e.g., susceptibility to shattering), root hair length and quantity, internode distances, or the quality of seed coat. Plant growth characteristics that may be modified include growth rate, germination rate of seeds, vigor of plants and seedlings, leaf and flower senescence, male sterility, apomixis, flowering time, flower abscission, rate of nitrogen uptake, biomass or transpiration characteristics, as well as plant architecture characteristics such as apical dominance, branching patterns, number of organs, organ identity, organ shape or size.

Of particular interest are traits relating to increased disease resistance or tolerance of a plant, such as alterations in cell wall composition, trichome number or structure, callose induction, phytoalexin induction, alterations in the cell death response or the like. These transgenic plants may be more resistant to biotrophic or necrotrophic pathogens such as a fungus, bacterium, mollicute, virus, nematode, a parasitic higher plant or the like and associated diseases. Another desirable phenotype is a change in the overall gene expression pattern of the plant in response to disease.

1. The Sequences

We have discovered particular plant transcription factors (TFs) that are induced when plants are exposed to either biotrophic or necrotrophic pathogens.. These transgenic plants may be more resistant to biotrophic or necrotrophic pathogens such as a fungus, bacterium, mollicute, virus, nematode, a parasitic higher plant or the like and associated diseases, in particular, pathogens such as *Fusarium oxysporum*, *Erysyphe orontii* and other powdery mildews, *Sclerotinia* spp., soil-borne oomycetes, foliar oomycetes, *Botrytis* spp., *Rhizoctonia* spp., *Verticillium dahliae/albo-atrum*, *Alternaria* spp., rusts, *Mycosphaerella* spp., *Fusarium solani*, or the like. The diseases include fungal diseases such as rusts, smuts, wilts, yellows, root rot, leaf drop, ergot, leaf blight of potato, brown spot of rice, leaf blight, late blight, powdery mildew, downy mildew, and the like; viral diseases such as sugarcane mosaic, cassava mosaic, sugar beet yellows, plum pox, barley yellow dwarf, tomato yellow leaf curl, tomato spotted wilt virus, and the like; bacterial diseases such as citrus canker, bacterial leaf blight, bacterial wilt, soft rot of vegetables, and the like; nematode diseases such as root knot, sugar beet cyst nematode or the like.

These transcription factors can be used to modulate a plant's response to disease. The plant transcription factors may belong to one of the following transcription factor families:

the AP2 (APETALA2) domain transcription factor family (Riechmann and Meyerowitz (1998) *J. Biol. Chem.* 379:633-646); the MYB transcription factor family (Martin and Paz-Ares, (1997) *Trends Genet.* 13:67-73); the MADS domain transcription factor family (Riechmann and Meyerowitz (1997) *J. Biol. Chem.* 378:1079-1101); the WRKY protein family (Ishiguro and Nakamura (1994) *Mol. Gen. Genet.* 244:563-571); the ankyrin-repeat protein family (Zhang et al. (1992) *Plant Cell* 4:1575-1588); the zinc finger protein (Z) family (Klug and Schwabe (1995) *FASEB J.* 9: 597-604); the homeobox (HB) protein family (Duboule (1994) *Guidebook to the Homeobox Genes*, Oxford University Press); the CAAT-element binding proteins (Forsburg and Guarente (1989) *Genes Dev.* 3:1166-1178); the squamosa promoter binding proteins

5 (SPB) (Klein et al. (1996) *Mol. Gen. Genet.* 1996 250:7-16); the NAM protein family (Souer et al. (1996) *Cell* 85:159-170); the IAA/AUX proteins (Rouse et al. (1998) *Science* 279:1371–1373); the HLH/MYC protein family (Littlewood et al. (1994) *Prot. Profile* 1:639-709); the DNA-binding protein (DBP) family (Tucker et al. (1994) *EMBO J.* 13:2994-3002); the bZIP family of transcription factors (Foster et al. (1994) *FASEB J.* 8:192-200); the Box P-binding protein (the

10 BPF-1) family (da Costa e Silva et al. (1993) *Plant J.* 4:125-135); the high mobility group (HMG) family (Bustin and Reeves (1996) *Prog. Nucl. Acids Res. Mol. Biol.* 54:35-100); the scarecrow (SCR) family (Di Laurenzio et al. (1996) *Cell* 86:423-433); the GF14 family (Wu et al. (1997) *Plant Physiol.* 114:1421-1431); the polycomb (PCOMB) family (Kennison (1995) *Annu. Rev. Genet.* 29:289-303); the teosinte branched (TEO) family (Luo et al. (1996) *Nature* 383:794-799; the ABI3 family (Giraudat et al. (1992) *Plant Cell* 4:1251-1261); the triple helix

15 (TH) family (Dehesh et al. (1990) *Science* 250:1397-1399); the EIL family (Chao et al. (1997) *Cell* 89:1133-44); the AT-HOOK family (Reeves and Nissen (1990) *Journal of Biological Chemistry* 265:8573-8582); the S1FA family (Zhou et al. (1995) *Nucleic Acids Res.* 23:1165-1169); the bZIP2 family (Lu and Ferl (1995) *Plant Physiol.* 109:723); the YABBY family

20 (Bowman et al. (1999) *Development* 126:2387-96); the PAZ family (Bohmert et al. (1998) *EMBO J.* 17:170-80); a family of miscellaneous (MISC) transcription factors including the DPBF family (Kim et al. (1997) *Plant J.* 11:1237-1251) and the SPF1 family (Ishiguro and Nakamura (1994) *Mol. Gen. Genet.* 244:563-571); the golden (GLD) family (Hall et al. (1998) *Plant Cell* 10:925-936).

25 Producing transgenic plants with modified expression levels of one or more of these transcription factors compared with those levels found in a wild type or reference plant may be used to modify a plant's traits. The effect of modifying the expression levels of a particular transcription factor on the traits of a transgenic plant is described further in the Examples.

30 The polynucleotides and polypeptides are provided in the Sequence Listing and are tabulated in Figure 1. Figure 1 identifies a SEQ ID No., its corresponding GID number, the transcription factor family to which the sequence belongs, fragments derived from the sequences, whether the sequence is a polynucleotide or a polypeptide sequence, the full

length coding sequences and conserved domains. We have also identified domains or fragments derived from the sequences. The numbers indicating the fragment location for the DNA sequences may be from either 5' or 3' end of the DNA. For the protein sequences the fragment location is determined from the N-terminus of the protein and may include adjacent amino acid sequences, such as for example for SEQ ID No. 2 an additional 10, 20, 40, 60 or 100 amino acids in either N-terminal or C-terminal direction of the described fragments.

The identified polypeptide fragments may be linked to fragments or sequences derived from other transcription factors so as to generate additional novel sequences, such as by employing the methods described in Short, PCT publication WO9827230, entitled "Methods and Compositions for Polypeptide Engineering" or in Patten et al., PCT publication WO9923236, entitled "Method of DNA Shuffling". Alternatively, the identified fragment may be linked to a transcription activation domain. A transcription activation domain assists in initiating transcription from a DNA binding site. A common feature of some activation domains is that they are designed to form amphiphilic alpha helices with excess positive or negative charge (Giniger and Ptashne (1987) *Nature* 330:670-672, Gill and Ptashne (1987) *Cell* 51:121-126, Estruch et al (1994) *Nucl. Acids Res.* 22:3983-3989). Examples include the transcription activation region of VP16 or GAL4 (Moore et al. (1998) *Proc. Natl. Acad. Sci. USA* 95: 376-381; and Aoyama et al. (1995) *Plant Cell* 7:1773-1785), peptides derived from bacterial sequences (Ma and Ptashne (1987) *Cell* 51: 113-119) and synthetic peptides (Giniger and Ptashne, *supra*).

The isolated polynucleotides and polypeptides may be used to modify plant development, physiology or biochemistry such that the modified plants have a trait advantage over wild type plants. The identified polynucleotide fragments are also useful as nucleic acid probes and primers. A nucleic acid probe is useful in hybridization protocols, including protocols for microarray experiments. Primers may be annealed to a complementary target DNA strand by nucleic acid hybridization to form a hybrid between the primer and the target DNA strand, and then extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR) or other nucleic-acid amplification methods. See Sambrook et al., *Molecular Cloning. A Laboratory Manual*, Ed. 2, Cold Spring Harbor Laboratory Press, New York (1989) and Ausubel et al. (eds) *Current Protocols in Molecular Biology*, John Wiley & Sons (1998).

2. Identification of Homologous Sequences (Homologs)

Homologous sequences to those provided in the Sequence Listing derived from *Arabidopsis thaliana* or from other plants may be used to modify a plant trait. Homologous sequences may be derived from any plant including monocots and dicots and in particular

agriculturally important plant species, including but not limited to, crops such as soybean, wheat, corn, potato, cotton, rice, oilseed rape (including canola), sunflower, alfalfa, sugarcane and turf; or fruits and vegetables, such as banana, blackberry, blueberry, strawberry, and raspberry, cantaloupe, carrot, cauliflower, coffee, cucumber, eggplant, grapes, honeydew, lettuce, mango, melon, onion, papaya, peas, peppers, pineapple, spinach, squash, sweet corn, tobacco, tomato, watermelon, rosaceous fruits (such as apple, peach, pear, cherry and plum) and vegetable brassicas (such as broccoli, cabbage, cauliflower, brussel sprouts and kohlrabi). Other crops, fruits and vegetables whose phenotype may be changed include barley, currant, avocado, citrus fruits such as oranges, lemons, grapefruit and tangerines, artichoke, cherries, nuts such as the walnut and peanut, endive, leek, roots, such as arrowroot, beet, cassava, turnip, radish, yam, sweet potato and beans. The homologs may also be derived from woody species, such pine, poplar and eucalyptus.

Substitutions, deletions and insertions introduced into the sequences provided in the Sequence Listing are also envisioned by the invention. Such sequence modifications can be engineered into a sequence by site-directed mutagenesis (Wu (ed.) *Meth. Enzymol.* (1993) vol. 217, Academic Press). Amino acid substitutions are typically of single residues; insertions usually will be on the order of about from 1 to 10 amino acid residues; and deletions will range about from 1 to 30 residues. In preferred embodiments, deletions or insertions are made in adjacent pairs, e.g., a deletion of two residues or insertion of two residues.

Substitutions, deletions, insertions or any combination thereof may be combined to arrive at a sequence. The mutations that are made in the polynucleotide encoding the transcription factor should not place the sequence out of reading frame and should not create complementary regions that could produce secondary mRNA structure.

Substitutions are those in which at least one residue in the amino acid sequence has been removed and a different residue inserted in its place. Such substitutions may be conservative with little effect on the function of the gene, for example by substituting alanines for serines, arginines for lysines, glutamate for aspartate and the like. The substitutions which are not conservative are expected to produce the greatest changes in protein properties will be those in which (a) a hydrophilic residue, e.g., seryl or threonyl, is substituted for (or by) a hydrophobic residue, e.g., leucyl, isoleucyl, phenylalanyl, valyl or alanyl; (b) a cysteine or proline is substituted for (or by) any other residue; (c) a residue having an electropositive side chain, e.g., lysyl, arginyl, or histidyl, is substituted for (or by) an electronegative residue, e.g., glutamyl or aspartyl; or (d) a residue having a bulky side chain, e.g., phenylalanine, is substituted for (or by) one not having a side chain, e.g., glycine.

Additionally, the term "homologous sequence" may encompass a polypeptide sequence that is modified by chemical or enzymatic means. The homologous sequence may be a sequence modified by lipids, sugars, peptides, organic or inorganic compounds, by the

use of modified amino acids or the like. Protein modification techniques are illustrated in Ausubel et al. (eds) *Current Protocols in Molecular Biology*, John Wiley & Sons (1998).

Homologous sequences also may mean two sequences having a substantial percentage of sequence identity after alignment as determined by using sequence analysis programs for database searching and sequence alignment and comparison available, for example, from the Wisconsin Package Version 10.0, such as BLAST, FASTA, PILEUP, FINDPATTERNS or the like (GCG, Madison, WI). Public sequence databases such as GenBank, EMBL, Swiss-Prot and PIR or private sequence databases such as PhytoSeq (Incyte Pharmaceuticals, Palo Alto, CA) may be searched. Alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman (1981) *Adv. Appl. Math.* 2:482, by the homology alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity method of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci. U.S.A.* 85: 2444, by computerized implementations of these algorithms. After alignment, sequence comparisons between two (or more) polynucleotides or polypeptides are typically performed by comparing sequences of the two sequences over a comparison window to identify and compare local regions of sequence similarity. The comparison window may be a segment of at least about 20 contiguous positions, usually about 50 to about 200, more usually about 100 to about 150 contiguous positions. A description of the method is provided in Ausubel et al. (eds) (1999) *Current Protocols in Molecular Biology*, John Wiley & Sons.

Transcription factors that are homologs of the disclosed sequences will typically share at least 40% amino acid sequence identity. More closely related TFs may share at least 50%, 60%, 65%, 70%, 75% or 80% sequence identity with the disclosed sequences. Factors that are most closely related to the disclosed sequences share at least 85%, 90% or 95% sequence identity. At the nucleotide level, the sequences will typically share at least 40% nucleotide sequence identity, preferably at least 50%, 60%, 70% or 80% sequence identity, and more preferably 85%, 90%, 95% or 97% sequence identity. The degeneracy of the genetic code enables major variations in the nucleotide sequence of a polynucleotide while maintaining the amino acid sequence of the encoded protein.

One way to identify whether two nucleic acid molecules are closely related is that the two molecules hybridize to each other under stringent conditions. Generally, stringent conditions are selected to be about 5°C to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Conditions for nucleic acid hybridization and calculation of stringencies can be found in Sambrook et al. (1989) *Molecular Cloning. A Laboratory Manual*, Ed. 2, Cold Spring Harbor Laboratory Press, New York and Tijssen (1993) *Laboratory Techniques in Biochemistry and*

Molecular Biology--Hybridization with Nucleic Acid Probes Part I, Elsevier, New York . Nucleic acid molecules that hybridize under stringent conditions will typically hybridize to a probe based on either the entire cDNA or selected portions of the cDNA under wash conditions of 0.2x SSC to 2.0 x SSC, 0.1% SDS at 50-65° C, for example 0.2 x SSC, 0.1% SDS at 65° C. For detecting less closely related homologs washes may be performed at 50° C.

For conventional hybridization the hybridization probe is conjugated with a detectable label such as a radioactive label, and the probe is preferably of at least 20 nucleotides in length. As is well known in the art, increasing the length of hybridization probes tends to give enhanced specificity. The labeled probe derived from the *Arabidopsis* nucleotide sequence may be hybridized to a plant cDNA or genomic library and the hybridization signal detected using means known in the art. The hybridizing colony or plaque (depending on the type of library used) is then purified and the cloned sequence contained in that colony or plaque isolated and characterized. Homologs may also be identified by PCR-based techniques, such as inverse PCR or RACE, using degenerate primers. See Ausubel et al. (eds) (1998) *Current Protocols in Molecular Biology*, John Wiley & Sons.

TF homologs may alternatively be obtained by immunoscreening an expression library. With the provision herein of the disclosed TF nucleic acid sequences, the polypeptide may be expressed and purified in a heterologous expression system (e.g., *E. coli*) and used to raise antibodies (monoclonal or polyclonal) specific for the TF. Antibodies may also be raised against synthetic peptides derived from TF amino acid sequences. Methods of raising antibodies are well known in the art and are described in Harlow and Lane (1988) *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, New York. Such antibodies can then be used to screen an expression library produced from the plant from which it is desired to clone the TF homolog, using the methods described above. The selected cDNAs may be confirmed by sequencing and enzymatic activity.

3. Altered Expression of Transcription Factors

Any of the identified sequences may be incorporated into a cassette or vector for expression in plants. A number of expression vectors suitable for stable transformation of plant cells or for the establishment of transgenic plants have been described including those described in Weissbach and Weissbach, (1989) *Methods for Plant Molecular Biology*, Academic Press, and Gelvin et al., (1990) *Plant Molecular Biology Manual*, Kluwer Academic Publishers. Specific examples include those derived from a Ti plasmid of *Agrobacterium tumefaciens*, as well as those disclosed by Herrera-Estrella, L., et al., (1983) *Nature* 303: 209, Bevan, M., *Nucl. Acids Res.* (1984) 12: 8711-8721, Klee, H. J., (1985) *Bio/Technology* 3: 637-642, for dicotyledonous plants. Ti-derived plasmids can be transferred into both

monocotinous and dicotyledonous species using *Agrobacterium*-mediated transformation (Ishida et al (1996) *Nat. Biotechnol.* 14:745-50; Barton et al. (1983) *Cell* 32:1033-1043).

Alternatively, non-Ti vectors can be used to transfer the DNA into plants and cells by using free DNA delivery techniques. Such methods may involve, for example, the use of 5 liposomes, electroporation, microprojectile bombardment, silicon carbide whiskers, and viruses. By using these methods transgenic plants such as wheat, rice (Christou, P., (1991) *Bio/Technology* 9: 957-962) and corn (Gordon-Kamm, W., (1990) *Plant Cell* 2: 603-618) can be produced. An immature embryo can also be a good target tissue for monocots for direct 10 DNA delivery techniques by using the particle gun (Weeks, T. et al., (1993) *Plant Physiol.* 102: 1077-1084; Vasil, V., (1993) *Bio/Technology* 10: 667-674; Wan, Y. and Lemeaux, P., (1994) *Plant Physiol.* 104: 37-48, and for *Agrobacterium*-mediated DNA transfer (Ishida et al., (1996) *Nature Biotech.* 14: 745-750).

Typically, plant transformation vectors include one or more cloned plant coding 15 sequences (genomic or cDNA) under the transcriptional control of 5' and 3' regulatory sequences and a dominant selectable marker. Such plant transformation vectors typically also contain a promoter (e.g., a regulatory region controlling inducible or constitutive, environmentally-or developmentally-regulated, or cell- or tissue-specific expression), a transcription initiation start site, an RNA processing signal (such as intron splice sites), a transcription termination site, and/or a polyadenylation signal.

20 Examples of constitutive plant promoters which may be useful for expressing the TF sequence include: the cauliflower mosaic virus (CaMV) 35S promoter, which confers constitutive, high-level expression in most plant tissues (see, e.g., Odel et al., (1985) *Nature* 313:810); the nopaline synthase promoter (An et al., (1988) *Plant Physiol.* 88:547); and the octopine synthase promoter (Fromm et al., (1989) *Plant Cell* 1: 977).

25 A variety of plant gene promoters that regulate gene expression in response to environmental, hormonal, chemical, developmental signals, and in a tissue-active manner can be used for expression of the TFs in plants, as illustrated by seed-specific promoters (such as the napin, phaseolin or DC3 promoter described in US Pat. No. 5,773,697), root-specific promoters, such as those disclosed in US Patent Nos. 5,618,988, 5,837,848 and 5,905,186; 30 fruit-specific promoters that are active during fruit ripening (such as the dru 1 promoter (US Pat. No. 5,783,393), or the 2A11 promoter (US Pat. No. 4,943,674) and the tomato polygalacturonase promoter (Bird et al. (1988) *Plant Mol. Biol.* 11:651), root-specific promoters, such as those disclosed in US Patent Nos. 5,618,988, 5,837,848 and 5,905,186, pollen-active promoters such as PTA29, PTA26 and PTA13 (US Pat. No. 5,792,929), 35 promoters active in vascular tissue (Ringli and Keller (1998) *Plant Mol. Biol.* 37:977-988), flower-specific (Kaiser et al, (1995) *Plant Mol. Biol.* 28:231-243), pollen (Baerson et al. (1994) *Plant Mol. Biol.* 26:1947-1959), carpels (Ohl et al. (1990) *Plant Cell* 2:837-848), pollen and

ovules (Baerson et al. (1993) *Plant Mol. Biol.* 22:255-267) auxin-inducible promoters (such as that described in van der Kop et al (1999) *Plant Mol. Biol.* 39:979-990 or Baumann et al. (1999) *Plant Cell* 11:323-334), cytokinin-inducible promoter (Guevara-Garcia (1998) *Plant Mol. Biol.* 38:743-753), promoters responsive to gibberellin (Shi et al. (1998) *Plant Mol. Biol.* 38:1053-1060, Willmott et al. (1998) 38:817-825) and the like. Additional promoters are those that elicit expression in response to heat (Ainley, et al. (1993) *Plant Mol. Biol.* 22: 13-23), light (e.g., the pea rbcS-3A promoter, Kuhlemeier et al., (1989) *Plant Cell* 1:471, and the maize rbcS promoter, Schaffner and Sheen, (1991) *Plant Cell* 3: 997); wounding (e.g., *wun1*, Siebertz et al., (1989) *Plant Cell* 1: 961); pathogen resistance, and chemicals such as methyl 5 jasmonate or salicylic acid (Gatz et al., (1997) *Plant Mol. Biol.* 48: 89-108). In addition, the 10 timing of the expression can be controlled by using promoters such as those acting at late seed development (Odell et al. (1994) *Plant Physiol.* 106:447-458).

Plant expression vectors may also include RNA processing signals that may be positioned within, upstream or downstream of the coding sequence. In addition, the 15 expression vectors may include additional regulatory sequences from the 3'-untranslated region of plant genes, e.g., a 3' terminator region to increase mRNA stability of the mRNA, such as the PI-II terminator region of potato or the octopine or nopaline synthase 3' terminator regions.

Finally, as noted above, plant expression vectors may also include dominant 20 selectable marker genes to allow for the ready selection of transformants. Such genes include those encoding antibiotic resistance genes (e.g., resistance to hygromycin, kanamycin, bleomycin, G418, streptomycin or spectinomycin) and herbicide resistance genes (e.g., phosphinothricin acetyltransferase).

A reduction of TF expression in a transgenic plant to modify a plant trait may be 25 obtained by introducing into plants antisense constructs based on the TF cDNA. For antisense suppression, the TF cDNA is arranged in reverse orientation relative to the promoter sequence in the expression vector. The introduced sequence need not be the full length TF cDNA or gene, and need not be identical to the TF cDNA or a gene found in the plant type to be transformed. Generally, however, where the introduced sequence is of shorter length, a 30 higher degree of homology to the native TF sequence will be needed for effective antisense suppression. Preferably, the introduced antisense sequence in the vector will be at least 30 nucleotides in length, and improved antisense suppression will typically be observed as the length of the antisense sequence increases. Preferably, the length of the antisense sequence in the vector will be greater than 100 nucleotides. Transcription of an antisense construct as 35 described results in the production of RNA molecules that are the reverse complement of mRNA molecules transcribed from the endogenous TF gene in the plant cell. Suppression of endogenous TF gene expression can also be achieved using a ribozyme. Ribozymes are

synthetic RNA molecules that possess highly specific endoribonuclease activity. The production and use of ribozymes are disclosed in U.S. Patent No. 4,987,071 to Cech and U.S. Patent No. 5,543,508 to Haselhoff. The inclusion of ribozyme sequences within antisense RNAs may be used to confer RNA cleaving activity on the antisense RNA, such that 5 endogenous mRNA molecules that bind to the antisense RNA are cleaved, which in turn leads to an enhanced antisense inhibition of endogenous gene expression.

Vectors in which RNA encoded by the TF cDNA (or variants thereof) is over-expressed may also be used to obtain co-suppression of the endogenous TF gene in the manner described in U.S. Patent No. 5,231,020 to Jorgensen. Such co-suppression (also 10 termed sense suppression) does not require that the entire TF cDNA be introduced into the plant cells, nor does it require that the introduced sequence be exactly identical to the endogenous TF gene. However, as with antisense suppression, the suppressive efficiency will be enhanced as (1) the introduced sequence is lengthened and (2) the sequence similarity between the introduced sequence and the endogenous TF gene is increased.

15 Vectors expressing an untranslatable form of the TF mRNA may also be used to suppress the expression of endogenous TF activity to modify a trait. Methods for producing such constructs are described in U.S. Patent No. 5,583,021 to Dougherty et al. Preferably, such constructs are made by introducing a premature stop codon into the TF gene. Alternatively, a plant trait may be modified by gene silencing using double-strand RNA (Sharp (1999) *Genes and 20 Development* 13: 139-141).

25 Another method for abolishing the expression of a gene is by insertion mutagenesis using the T-DNA of *Agrobacterium tumefaciens*. After generating the insertion mutants, the mutants can be screened to identify those containing the insertion in a TF gene. Mutants containing a single mutation event at the desired gene may be crossed to generate homozygous plants for the mutation (Koncz et al. (1992) *Methods in Arabidopsis Research*. World Scientific).

30 A plant trait may also be modified by using the cre-lox system (for example, as described in US Pat. No. 5,658,772). A plant genome may be modified to include first and second lox sites that are then contacted with a Cre recombinase. If the lox sites are in the same orientation, the intervening DNA sequence between the two sites is excised. If the lox sites are in the opposite orientation, the intervening sequence is inverted.

35 The polynucleotides and polypeptides of this invention may also be expressed in a plant in the absence of an expression cassette by manipulating the activity or expression level of the endogenous gene by other means. For example, by ectopically expressing a gene by T-DNA activation tagging (Ichikawa et al., (1997) *Nature* 390 698-701, Kakimoto et al., (1996) *Science* 274: 982-985). This method entails transforming a plant with a gene tag containing multiple transcriptional enhancers and once the tag has inserted into the genome, expression of a flanking gene coding sequence becomes deregulated. In another example, the

transcriptional machinery in a plant may be modified so as to increase transcription levels of a polynucleotide of the invention (See PCT Publications WO9606166 and WO 9853057 which describe the modification of the DNA binding specificity of zinc finger proteins by changing particular amino acids in the DNA binding motif).

5 The transgenic plant may also comprise the machinery necessary for expressing or altering the activity of a polypeptide encoded by an endogenous gene, for example by altering the phosphorylation state of the polypeptide to maintain it in an activated state.

4. Transgenic Plants with Modified TF Expression

10 Once an expression cassette comprising a polynucleotide encoding a TF gene of this invention has been constructed, standard techniques may be used to introduce the polynucleotide into a plant in order to modify a trait of the plant. The plant may be any higher plant, including gymnosperms, monocotyledonous and dicotyledonous plants. Suitable protocols are available for *Leguminosae* (alfalfa, soybean, clover, etc.), *Umbelliferae* (carrot, 15 celery, parsnip), *Cruciferae* (cabbage, radish, rapeseed, broccoli, etc.), *Curcurbitaceae* (melons and cucumber), *Gramineae* (wheat, corn, rice, barley, millet, etc.), *Solanaceae* (potato, tomato, tobacco, peppers, etc.), and various other crops. See protocols described in Ammirato et al. (1984) *Handbook of Plant Cell Culture –Crop Species*. Macmillan Publ. Co. Shimamoto et al. (1989) *Nature* 338:274-276; Fromm et al. (1990) *Bio/Technology* 8:833-20 839; and Vasil et al. (1990) *Bio/Technology* 8:429-434.

25 Transformation and regeneration of both monocotyledonous and dicotyledonous plant cells is now routine, and the selection of the most appropriate transformation technique will be determined by the practitioner. The choice of method will vary with the type of plant to be transformed; those skilled in the art will recognize the suitability of particular methods for given plant types. Suitable methods may include, but are not limited to: electroporation of plant protoplasts; liposome-mediated transformation; polyethylene glycol (PEG) mediated transformation; transformation using viruses; micro-injection of plant cells; micro-projectile bombardment of plant cells; vacuum infiltration; and *Agrobacterium tumefaciens* mediated transformation. Transformation means introducing a nucleotide sequence in a plant in a 30 manner to cause stable or transient expression of the sequence.

Successful examples of the modification of plant characteristics by transformation with cloned sequences which serve to illustrate the current knowledge in this field of technology, and which are herein incorporated by reference, include: U.S. Patent Nos. 5,571,706; 5,677,175; 5,510,471; 5,750,386; 5,597,945; 5,589,615; 5,750,871; 5,268,526; 5,780,708; 35 5,538,880; 5,773,269; 5,736,369 and 5,610,042.

Following transformation, plants are preferably selected using a dominant selectable marker incorporated into the transformation vector. Typically, such a marker will confer

antibiotic or herbicide resistance on the transformed plants, and selection of transformants can be accomplished by exposing the plants to appropriate concentrations of the antibiotic or herbicide.

After transformed plants are selected and grown to maturity, those plants showing a modified trait are identified. The modified trait may be any of those traits described above. Additionally, to confirm that the modified trait is due to changes in expression levels or activity of the polypeptide or polynucleotide of the invention may be determined by analyzing mRNA expression using Northern blots, RT-PCR or microarrays, or protein expression using immunoblots or Western blots or gel shift assays.

The plants may have commercial utility for increasing tolerance or resistance to pathogens and pests. These transgenic plants may be more resistant to biotrophic or necrotrophic pathogens or belonging to the following groups such as a fungus, bacterium, mollicute, virus, nematode, a parasitic higher plant or the like and associated diseases. In particular, pathogens such as *Fusarium oxysporum*, *Erysyphe orontii* and other powdery mildews, *Sclerotinia* spp., soil-borne oomycetes, foliar oomycetes, *Botrytis* spp., *Rhizoctonia* spp., *Verticillium dahliae/albo-atrum*, *Alternaria* spp., rusts, *Mycosphaerella* spp., *Fusarium solani*, or the like. The diseases include fungal diseases such as rusts, smuts, wilts, yellows, root rot, leaf drop, ergot, leaf blight of potato, brown spot of rice, leaf blight, late blight, powdery mildew, downy mildew, and the like; viral diseases such as sugarcane mosaic, cassava mosaic, sugar beet yellows, plum pox, barley yellow dwarf, tomato yellow leaf curl, tomato spotted wilt virus, and the like; bacterial diseases such as citrus canker, bacterial leaf blight, bacterial wilt, soft rot of vegetables, and the like; nematode diseases caused by parasitic nematodes such as root-knot nematodes, cyst nematodes or the like.

5. Other Utility of the Polypeptide and Polynucleotide Sequences

A transcription factor provided by the present invention may also be used to identify exogenous or endogenous molecules that may affect expression of the transcription factors and may affect any of the traits described herein. These molecules may include organic or inorganic compounds.

For example, the method may entail first placing the molecule in contact with a plant or plant cell. The molecule may be introduced by topical administration, such as spraying or soaking of a plant, and then the molecule's effect on the expression or activity of the TF polypeptide or the expression of the polynucleotide monitored. Changes in the expression of the TF polypeptide may be monitored by use of polyclonal or monoclonal antibodies, gel electrophoresis or the like. Changes in the expression of the corresponding polynucleotide sequence may be detected by use of microarrays, Northerns or any other technique for monitoring changes in mRNA expression. These techniques are exemplified in Ausubel et al.

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(eds) *Current Protocols in Molecular Biology*, John Wiley & Sons (1998). Such changes in the expression levels may be correlated with modified plant traits and thus identified molecules may be useful for soaking or spraying on fruit, vegetable and grain crops to modify traits in plants.

5 The transcription factors may also be employed to identify promoter sequences with which they may interact. After identifying a promoter sequence, interactions between the transcription factor and the promoter sequence may be modified by changing specific nucleotides in the promoter sequence or specific amino acids in the transcription factor that interact with the promoter sequence to alter a plant trait. Typically, transcription factor DNA binding sites are identified by gel shift assays. After identifying the promoter regions, the 10 promoter region sequences may be employed in double-stranded DNA arrays to identify molecules that affect the interactions of the TFs with their promoters (Bulyk et al. (1999) *Nature Biotechnology* 17:573-577).

15 The identified transcription factors are also useful to identify proteins that modify the activity of the transcription factor. Such modification may occur by covalent modification, such as by phosphorylation, or by protein-protein (homo or-heteropolymer) interactions. Any method suitable for detecting protein-protein interactions may be employed. Among the methods that may be employed are co-immunoprecipitation, cross-linking and co-purification through gradients or chromatographic columns, and the two-hybrid yeast system.

20 The two-hybrid system detects protein interactions *in vivo* and is described in Chien, et al., (1991), *Proc. Natl. Acad. Sci. USA*, 88, 9578-9582 and is commercially available from Clontech (Palo Alto, Calif.). In such a system, plasmids are constructed that encode two hybrid proteins: one consists of the DNA-binding domain of a transcription activator protein fused to the TF polypeptide and the other consists of the transcription activator protein's 25 activation domain fused to an unknown protein that is encoded by a cDNA that has been recombined into the plasmid as part of a cDNA library. The DNA-binding domain fusion plasmid and the cDNA library are transformed into a strain of the yeast *Saccharomyces cerevisiae* that contains a reporter gene (e.g., lacZ) whose regulatory region contains the transcription activator's binding site. Either hybrid protein alone cannot activate transcription of 30 the reporter gene. Interaction of the two hybrid proteins reconstitutes the functional activator protein and results in expression of the reporter gene, which is detected by an assay for the reporter gene product. Then, the library plasmids responsible for reporter gene expression are isolated and sequenced to identify the proteins encoded by the library plasmids. After identifying proteins that interact with the transcription factors, assays for compounds that 35 interfere with the TF protein-protein interactions may be performed.

The following examples are intended to illustrate but not limit the present invention.

Example I. Full Length Gene Identification and Cloning

Putative transcription factor sequences (genomic or ESTs) related to known transcription factors were identified in the *Arabidopsis thaliana* GenBank database using the tblastn sequence analysis program using default parameters and a P-value cutoff threshold of –4 or –5 or lower, depending on the length of the query sequence. Putative transcription factor sequence hits were then screened to identify those containing particular sequence strings. If the sequence hits contained such sequence strings, the sequences were confirmed as transcription factors.

Alternatively, *Arabidopsis thaliana* cDNA libraries derived from different tissues or treatments, or genomic libraries were screened to identify novel members of a transcription family using a low stringency hybridization approach. Probes were synthesized using gene specific primers in a standard PCR reaction (annealing temperature 60° C) and labeled with ^{32}P dCTP using the High Prime DNA Labeling Kit (Boehringer Mannheim). Purified radiolabelled probes were added to filters immersed in Church hybridization medium (0.5 M NaPO₄ pH 7.0, 7% SDS, 1 % w/v bovine serum albumin) and hybridized overnight at 60 °C with shaking. Filters were washed two times for 45 to 60 minutes with 1xSCC, 1% SDS at 60° C.

To identify additional sequence 5' or 3' of a partial cDNA sequence in a cDNA library, 5' and 3' rapid amplification of cDNA ends (RACE) was performed using the Marathon™ cDNA amplification kit (Clontech, Palo Alto, CA). Generally, the method entailed first isolating poly(A) mRNA, performing first and second strand cDNA synthesis to generate double stranded cDNA, blunting cDNA ends, followed by ligation of the Marathon™ Adaptor to the cDNA to form a library of adaptor-ligated ds cDNA. Gene-specific primers were designed to be used along with adaptor specific primers for both 5' and 3' RACE reactions. Nested primers, rather than single primers, were used to increase PCR specificity. Using 5' and 3' RACE reactions, 5' and 3' RACE fragments were obtained, sequenced and cloned. The process may be repeated until 5' and 3' ends of the full-length gene were identified. Then the full-length cDNA was generated by PCR using primers specific to 5' and 3' ends of the gene by end-to-end PCR.

Example II Pathogen Resistance Genes

RT-PCR and microarray experiments were performed to identify those genes induced after exposure to biotrophic fungal pathogens, such as *Erysiphe orontii*, necrotrophic fungal pathogens, such as *Fusarium oxysporum*, and disease associated growth-regulators such as salicylic acid, methyl jasmonate and ethylene (ACC). The gene expression patterns from soil grown as well as tissue culture grown plant tissue were investigated.

Fusarium oxysporum isolates cause vascular wilts and damping off of various annual vegetables, perennials and weeds (Mauch-Mani and Slusarenko (1994) Molecular Plant-Microbe Interactions 7: 378-383). For *Fusarium oxysporum* experiments, plants grown on petri dishes were sprayed with a fresh spore suspension of *F. oxysporum*. The spore suspension was prepared as follows: A plug of fungal hyphae from a plate culture was placed on a fresh potato dextrose agar plate and allowed to spread for one week. 5 ml sterile water was then added to the plate, swirled, and pipetted into 50 ml Armstrong Fusarium medium. Spores were grown overnight in Fusarium medium and then sprayed onto plants using a Preval paint sprayer. Plant tissue was harvested and frozen in liquid nitrogen 48 hours post infection

Erysiphe orontii is a causal agent of powdery mildew. For *Erysiphe orontii* experiments, plants were grown approximately 4 weeks in a greenhouse under 12 hour light (20 C, ~30% relative humidity (rh)). Individual leaves were infected with *E. orontii* spores from infected plants using a camel's hair brush, and the plants were transferred to a Percival growth chamber (20 C, 80% rh.). Plant tissue was harvested and frozen in liquid nitrogen 7 days post infection.

For salicylic acid experiments, 15 day old seedlings grown on petri dishes were transferred to plates containing 0.5 mM salicylic acid (SA). After 72 hours, leaves were harvested and frozen in liquid nitrogen.

Reverse transcriptase PCR was done using gene specific primers within the coding region for each sequence identified. The primers were designed near the 3' region of each coding sequence initially identified.

Total RNA from these tissues were isolated using the CTAB extraction protocol. Once extracted total RNA was normalized in concentration across all the tissue types to ensure that the PCR reaction for each tissue received the same amount of cDNA template using the 28S band as reference. Poly A+ was purified using a modified protocol from the Qiagen Oligotex kit batch protocol. cDNA was synthesized using standard protocols. After the first strand cDNA synthesis, primers for Actin 2 were used to normalize the concentration of cDNA across the tissue types. Actin 2 is found to be constitutively expressed in fairly equal levels across the tissue types we are investigating.

For RT PCR, cDNA template was mixed with corresponding primers and Taq polymerase. Each reaction consisted of 0.2 ul cDNA template, 2ul 10X Tricine buffer, 2 ul 10X Tricine buffer and 16.8 ul water, 0.05ul Primer 1, 0.05 ul, Primer 2, 0.3 ul Taq polymerase and 8.6 ul water.

The 96 well plate was covered with microfilm and set in the Thermocycler to start the following reaction cycle. Step1 93° C for 3 mins, Step 2 93° C for 30 sec, Step 3 65° C for 1 min, Step 4 72° C for 2 mins,. Steps 2, 3 and 4 were repeated for 28 cycles, Step 5 72° C

for 5 mins and Step 6 4° C. The PCR plate was placed back in the thermocycler to amplify more products at 8 more cycles to identify genes that have very low expression. The reaction cycle was as follows: Step 2 93° C for 30 sec, Step 3 65° C for 1 min, and Step 4 72° C for 2 ins, repeated for 8 cycles, and Step 4 4° C.

5 8ul of PCR product and 1.5 ul of loading dye were loaded on a 1.2% agarose gel for analysis after 28 cycles and 36 cycles. Expression levels of specific transcripts were considered low if they were only detectable after 36 cycles of PCR. Expression levels were considered medium or high depending on the levels of transcript compared with observed transcript levels for actin2.

10 In some instances, expression patterns of the transcription factors was monitored by microarray experiments. cDNAs were generated by PCR and resuspended at a final concentration of ~ 100 ng/ul in 3X SSC or 150mM Na-phosphate (Eisen and Brown (1999) *Meth. in Enzymol.* 303:179-205). The cDNAs were spotted on microscope glass slides coated with polylysine. The prepared cDNAs were aliquoted into 384 well plates and spotted on the slides using an x-y-z gantry (OmniGrid) purchased from GeneMachines (Menlo Park, CA) outfitted with quill type pins purchased from Telechem International (Sunnyvale, CA). After spotting, the arrays were cured for a minimum of one week at room temperature, rehydrated and blocked following the protocol recommended by Eisen and Brown (1999).

15 Sample total RNA (10 ug) samples were labeled using fluorescent Cy3 and Cy5 dyes. Labeled samples were resuspended in 4X SSC/0.03% SDS/4 ug salmon sperm DNA/2 ug tRNA/ 50mM Na-pyrophosphate, heated for 95°C for 2.5 minutes, spun down and placed on the array. The array was then covered with a glass coverslip and placed in a sealed chamber. The chamber was then kept in a water bath at 62°C overnight. The arrays were washed as described in Eisen and Brown (1999) and scanned on a General Scanning 3000 laser scanner. The resulting files are subsequently quantified using Imagene a software purchased from BioDiscovery (Los Angeles, CA).

20 The transcript levels were observed to be upregulated between 1.5 and 100 fold when compared with control plants not exposed to the pathogens.

30 Example III. Construction of Expression Vectors

35 The sequence was amplified from a genomic or cDNA library using primers specific to sequences upstream and downstream of the coding region. The expression vector was pMEN20, which is derived from pMON316 (Sanders et al, (1987) *Nucleic Acids Research* 15:1543-58). To clone the sequence into the vector, both pMEN20 and the amplified DNA fragment were digested separately with Sall and NotI restriction enzymes at 37° C for 2 hours. The digestion products were subject to electrophoresis in a 0.8% agarose gel and visualized

by ethidium bromide staining. The DNA fragments containing the sequence and the linearized plasmid were excised and purified by using a Qiaquick gel extraction kit (Qiagen, CA). The fragments of interest were ligated at a ratio of 3:1 (vector to insert). Ligation reactions using T4 DNA ligase (New England Biolabs, MA) were carried out at 16° C for 16 hours. The ligated DNAs were transformed into competent cells of the *E. coli* strain DH5alpha by using the heat shock method. The transformations were plated on LB plates containing 50 mg/l spectinomycin (Sigma).

Individual colonies were grown overnight in five milliliters of LB broth containing 50 mg/l spectinomycin at 37° C. Plasmid DNA was purified by using Qiaquick Mini Prep kits (Qiagen, CA).

Example IV. Transformation of *Agrobacterium* with the Expression Vector

After the plasmid vector containing the gene was constructed, the vector was used to transform *Agrobacterium tumefaciens* cells expressing the gene products. The stock of *Agrobacterium tumefaciens* cells for transformation were made as described by Nagel et al. *FEMS Microbiol Letts* 67: 325-328 (1990). *Agrobacterium* strain GV3101 was grown in 250 ml LB medium (Sigma) overnight at 28° C with shaking until an absorbance (A_{600}) of 0.5 – 1.0 was reached. Cells were harvested by centrifugation at 4,000 x g for 15 min at 4° C. Cells were then resuspended in 250 μ l chilled buffer (1 mM HEPES, pH adjusted to 7.0 with KOH). Cells were centrifuged again as described above and resuspended in 125 μ l chilled buffer. Cells were then centrifuged and resuspended two more times in the same HEPES buffer as described above at a volume of 100 μ l and 750 μ l, respectively. Resuspended cells were then distributed into 40 μ l aliquots, quickly frozen in liquid nitrogen, and stored at -80° C.

Agrobacterium cells were transformed with plasmids prepared as described above following the protocol described by Nagel et al. *FEMS Microbiol Letts* 67: 325-328 (1990). For each DNA construct to be transformed, 50 – 100 ng DNA (generally resuspended in 10 mM Tris-HCl, 1 mM EDTA, pH 8.0) was mixed with 40 μ l of *Agrobacterium* cells. The DNA/cell mixture was then transferred to a chilled cuvette with a 2mm electrode gap and subject to a 2.5 kV charge dissipated at 25 μ F and 200 μ F using a Gene Pulser II apparatus (Bio-Rad). After electroporation, cells were immediately resuspended in 1.0 ml LB and allowed to recover without antibiotic selection for 2 – 4 hours at 28° C in a shaking incubator. After recovery, cells were plated onto selective medium of LB broth containing 100 μ g/ml spectinomycin (Sigma) and incubated for 24-48 hours at 28° C. Single colonies were then picked and inoculated in fresh medium. The presence of the plasmid construct was verified by PCR amplification and sequence analysis.

Example V. Transformation of *Arabidopsis* Plants with *Agrobacterium tumefaciens* with Expression Vector

After transformation of *Agrobacterium tumefaciens* with plasmid vectors containing the gene, single *Agrobacterium* colonies were identified, propagated, and used to transform 5 *Arabidopsis* plants. Briefly, 500 ml cultures of LB medium containing 50 mg/l spectinomycin were inoculated with the colonies and grown at 28° C with shaking for 2 days until an absorbance (A_{600}) of > 2.0 is reached. Cells were then harvested by centrifugation at 4,000 x g for 10 min, and resuspended in infiltration medium (1/2 X Murashige and Skoog salts 10 (Sigma), 1 X Gamborg's B-5 vitamins (Sigma), 5.0% (w/v) sucrose (Sigma), 0.044 μ M benzylamino purine (Sigma), 200 μ l/L Silwet L-77 (Lehle Seeds) until an absorbance (A_{600}) of 0.8 was reached.

Prior to transformation, *Arabidopsis thaliana* seeds (ecotype Columbia) were sown at a density of ~10 plants per 4" pot onto Pro-Mix BX potting medium (Humert International) 15 covered with fiberglass mesh (18 mm X 16 mm). Plants were grown under continuous illumination (50-75 μ E/m²/sec) at 22-23° C with 65-70% relative humidity. After about 4 weeks, primary inflorescence stems (bolts) are cut off to encourage growth of multiple 20 secondary bolts. After flowering of the mature secondary bolts, plants were prepared for transformation by removal of all siliques and opened flowers.

The pots were then immersed upside down in the mixture of *Agrobacterium* infiltration 25 medium as described above for 30 sec, and placed on their sides to allow draining into a 1' x 2' flat surface covered with plastic wrap. After 24 h, the plastic wrap was removed and pots are turned upright. The immersion procedure was repeated one week later, for a total of two immersions per pot. Seeds were then collected from each transformation pot and analyzed following the protocol described below.

Example VI. Identification of *Arabidopsis* Primary Transformants

Seeds collected from the transformation pots were sterilized essentially as follows. Seeds were dispersed into in a solution containing 0.1% (v/v) Triton X-100 (Sigma) and sterile 30 H_2O and washed by shaking the suspension for 20 min. The wash solution was then drained and replaced with fresh wash solution to wash the seeds for 20 min with shaking. After removal of the second wash solution, a solution containing 0.1% (v/v) Triton X-100 and 70% ethanol (Equistar) was added to the seeds and the suspension was shaken for 5 min. After 35 removal of the ethanol/detergent solution, a solution containing 0.1% (v/v) Triton X-100 and 30% (v/v) bleach (Clorox) was added to the seeds, and the suspension was shaken for 10 min. After removal of the bleach/detergent solution, seeds were then washed five times in sterile distilled H_2O . The seeds were stored in the last wash water at 4° C for 2 days in the dark before being plated onto antibiotic selection medium (1 X Murashige and Skoog salts (pH

adjusted to 5.7 with 1M KOH), 1 X Gamborg's B-5 vitamins, 0.9% phytagar (Life Technologies), and 50 mg/l kanamycin). Seeds were germinated under continuous illumination (50-75 μ E/m²/sec) at 22-23° C. After 7-10 days of growth under these conditions, kanamycin resistant primary transformants (T₁ generation) were visible and obtained. These

5 seedlings were transferred first to fresh selection plates where the seedlings continued to grow for 3-5 more days, and then to soil (Pro-Mix BX potting medium).

Primary transformants are self-crossed and progeny seeds (T₂) collected.

10 **Example VII. Analysis of *Arabidopsis* T₂ progeny plants for Pathogen Resistance or Pathogen Tolerance**

T₂ or knockout mutant seed were surface sterilized and sown on MS media containing sucrose. Ten days post-planting, seedlings were transferred to MS media without sucrose. At two weeks of age *Arabidopsis* seedlings were inoculated with *Fusarium* by spraying with a spore suspension (2×10^6 conidia per milliliter) and incubated under high humidity. Plants were then scored macroscopically for disease symptoms or microscopically for fungal growth or using microarrays for the induction of resistance associated genes (such as the defensin genes) to detect resistance or tolerance of the plant tissue. A wild type plant shows the first signs of damage (gradual yellowing of leaves, damping off of seedlings or growth of fungal mycelium) after two to four days after inoculation. Transgenic plants that are pathogen resistant or tolerant showed a delay in disease or symptom development compared to wild-type control plants.

15 Alternatively, *Erysiphe* inoculations were done by tapping conidia from 1 to 2 heavily infected leaves onto the mesh cover of a settling tower, brushing the mesh with a camel's hair paint brush to break up the conidial chains, and letting the conidia settle for 10 minutes. Plants were 4 to 4.5 weeks old at the time of inoculation. Spores were obtained from 20 10 to 14 day old *Erysiphe* cultures. Typically, within the first twenty-four hours, the spores differentiated into several fungal structures including the haustorium that invaginates a host's epidermal plasma membrane. Formation of aerial mycelium and sporulation represent late differentiation events between 4 and 7 days post inoculation (Freilaldenhoven et al. (1994) 25 *Plant Cell* 6: 983-994). Plant resistance was scored based on the relative number and size of mycelial patches bearing conidia compared to wild-type control plants. Events associated with disease resistance to the pathogens and pests include: the induction of pathogen resistance related genes (R genes), the activation of cell death in the attacked epidermal cells 30 (hypersensitive response), the induction of anti-microbial compounds, such as phytoalexins, and the lignification that occurs at attempted penetration sites. Assays are performed to 35 observe these events. Transgenic plants identified that induce R genes, activate cell death, induce anti-microbial compounds or increase lignification sooner or to a greater extent than

wild-type plants when exposed to pathogen are potentially more resistant to infection by *Erysiphe* as well as a number of other pathogens and pests.

We have observed that when the expression levels of the genes are altered, that the disease phenotype can be varied. For example, G19 was significantly induced upon infection by the fungal pathogen *Erysiphe orontii* as well as the disease associated growth regulator, ethylene. Our data show that G19 overexpressing plants were more tolerant to infection with a moderate dose of *Erysiphe orontii* and in a nematode screen. The transgenic plants overexpressing G19 under the control of the 35S promoter were morphologically similar to control plants.

Additionally, G511 was another example of a gene that when overexpressed showed an increased tolerance to the fungal pathogen *Erysiphe orontii*. In both cases increased tolerance includes a significant reduction in pathogen growth and symptom development compared to wild type plants that were treated with pathogen in an identical manner.

Example VIII. Transformation of Cereal Plants with the Expression Vector

A cereal plant, such as corn, wheat, rice, sorghum or barley, can also be transformed with the plasmid vectors containing the sequence and constitutive or inducible promoters to modify a trait. In these cases, a cloning vector, pMEN020, is modified to replace the NptII coding region with the BAR gene of *Streptomyces hygroscopicus* that confers resistance to phosphinothricin. The KpnI and BgIII sites of the Bar gene are removed by site-directed mutagenesis with silent codon changes.

Plasmids according to the present invention may be transformed into corn embryogenic cells derived from immature scutellar tissue by using microprojectile bombardment, with the A188XB73 genotype as the preferred genotype (Fromm et al., *Bio/Technology* 8: 833-839 (1990); Gordon-Kamm et al., *Plant Cell* 2: 603-618 (1990)). After microprojectile bombardment the tissues are selected on phosphinothricin to identify the transgenic embryogenic cells (Gordon-Kamm et al., *Plant Cell* 2: 603-618 (1990)). Transgenic plants are regenerated by standard corn regeneration techniques (Fromm, et al., *Bio/Technology* 8: 833-839 (1990); Gordon-Kamm et al., *Plant Cell* 2: 603-618 (1990)).

Example IX. Identification of Homologous Sequences

Homologs from the same plant, different plant species or other organisms were identified using database sequence search tools, such as the Basic Local Alignment Search Tool (BLAST) (Altschul et al. (1990) *J. Mol. Biol.* 215:403-410; and Altschul et al. (1997) *Nucl. Acid Res.* 25: 3389-3402). The tblastn or blastn sequence analysis programs were employed using the BLOSUM-62 scoring matrix (Henikoff, S. and Henikoff, J. G. (1992) *Proc. Natl. Acad. Sci. USA* 89: 10915-10919). The output of a BLAST report provides a score that takes

into account the alignment of similar or identical residues and any gaps needed in order to align the sequences. The scoring matrix assigns a score for aligning any possible pair of sequences. The P values reflect how many times one expects to see a score occur by chance. Higher scores are preferred and a low threshold P value threshold is preferred.

5 These are the sequence identity criteria. The tblastn sequence analysis program was used to query a polypeptide sequence against six-way translations of sequences in a nucleotide database. Hits with a P value less than -25, preferably less than -70, and more preferably less than -100, were identified as homologous sequences (exemplary selected sequence criteria).

10 The blastn sequence analysis program was used to query a nucleotide sequence against a nucleotide sequence database. In this case too, higher scores were preferred and a preferred threshold P value was less than -13, preferably less than -50, and more preferably less than -100.

Alternatively, a fragment of a sequence from Figure 1 is ^{32}P -radiolabeled by random priming (Sambrook et al., (1989) *Molecular Cloning. A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory Press, New York) and used to screen a plant genomic library (the exemplary test polynucleotides). As an example, total plant DNA from *Arabidopsis thaliana*, *Nicotiana tabacum*, *Lycopersicon pimpinellifolium*, *Prunus avium*, *Prunus cerasus*, *Cucumis sativus*, or *Oryza sativa* are isolated according to Stockinger al (Stockinger, E. J., et al., (1996), *J. Heredity*, 87:214-218). Approximately 2 to 10 μg of each DNA sample are restriction digested, transferred to nylon membrane (Micron Separations, Westboro, MA) and hybridized. Hybridization conditions are: 42° C in 50% formamide, 5X SSC, 20 mM phosphate buffer 1X Denhardt's, 10% dextran sulfate, and 100 $\mu\text{g}/\text{ml}$ herring sperm DNA. Four low stringency washes at RT in 2X SSC, 0.05% sodium sarcosyl and 0.02% sodium pyrophosphate are performed prior to high stringency washes at 55° C in 0.2X SSC, 0.05% sodium sarcosyl and 0.01% sodium pyrophosphate. High stringency washes are performed until no counts are detected in the washout according to Walling et al. (Walling, L. L., et al., (1988) *Nucl. Acids Res.* 16:10477-10492).

30 All references (publications and patents) are incorporated herein by reference in their entirety for all purposes.

Although the invention has been described with reference to the embodiments and examples above, it should be understood that various modifications can be made without departing from the spirit of the invention. Accordingly, the invention is limited only by the following claims.

We Claim:

5 1. A transgenic plant comprising a recombinant polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of SEQ ID Nos. 2N, where N=1-56, wherein the recombinant polynucleotide alters the plant's disease tolerance or resistance when compared with the same trait of another plant lacking the recombinant polynucleotide.

10 2. The transgenic plant of claim 1, wherein the nucleotide sequence encodes a polypeptide comprising a conserved domain selected from the group consisting of SEQ ID Nos. 2N, where N=1-56

15 3. The transgenic plant of claim 1, wherein the recombinant polynucleotide further comprises a promoter operably linked to said nucleotide sequence.

20 4. The transgenic plant of claim 3, wherein said promoter is constitutive or inducible or tissue-active.

25 5. A method for altering the disease tolerance or resistance of a plant, said method comprising (a) transforming a plant with a recombinant polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of SEQ ID Nos. 2N, where N=1-56, (b) selecting said transformed plants; and (c) identifying a transformed plant having an altered disease tolerance or resistance.

30 6. The method of claim 5, wherein the nucleotide sequence encodes a polypeptide comprising a conserved domain selected from the group consisting of SEQ ID Nos. 2N, where N=1-56.

35 8. The method of claim 5, wherein the recombinant polynucleotide further comprises a promoter operably linked to said nucleotide sequence.

9. The method of claim 8, wherein said promoter is constitutive or inducible or tissue-active.

10. A method for altering the expression levels of at least one gene in a plant, said method comprising (a) transforming the plant with a recombinant polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising at least 6 consecutive amino acids of a sequence selected from the group consisting of SEQ ID Nos. 2N, where N=1-56; and (b) selecting said transformed plant.

5

11. The method of claim 10, wherein said recombinant polynucleotide encodes a polypeptide comprising a conserved domain selected from the group consisting of SEQ ID Nos. 2N, where N=1-56.

10

12. The method of claim 10, wherein the nucleotide sequence further comprises a promoter operably linked to said nucleotide sequence.

15

13. The method of claim 10, wherein said promoter is constitutive or inducible or tissue-active.

14. A method for altering the disease tolerance or resistance in a plant, said method comprising (a) transforming the plant with a recombinant polynucleotide comprising at least 18 consecutive nucleotides of a sequence selected from the group consisting of SEQ ID Nos. 2N-1, where N= 1-56, and SEQ ID Nos. 113-121; and (b) selecting said transformed plant.

20

15. A method for altering a plant's trait, said method comprising (a) providing a database sequence; (b) comparing said database sequence with a polypeptide selected from SEQ ID Nos. 2N, where N= 1-56; (c) selecting a database sequence that meets selected sequence criteria; and (d) transforming said selected database sequence in the plant.

25

16. A method for altering a plant's trait, said method comprising (a) providing a database sequence; (b) comparing said database sequence with a polynucleotide selected from SEQ ID Nos. 2N-1, where N= 1-56 or SEQ ID Nos. 113-121; (c) selecting a database sequence that meets selected sequence criteria; and (d) transforming said selected database sequence in the plant.

30

17. A method for altering a plant's trait, said method comprising (a) providing a test polynucleotide; (b) hybridizing said test polynucleotide with a polynucleotide selected from SEQ ID Nos. 2N-1, where N= 1-56 or SEQ ID Nos. 113-121 at low stringency; and (c) transforming said hybridizing test polynucleotide in a plant to alter a trait of the plant.

35

ABSTRACT OF THE INVENTION

Recombinant polynucleotides and methods for altering the regulation of gene expression in plants are provided to modify a plant's traits, in particular disease tolerance.

Figure 1a

SEQ ID No	GID No.	Family	Fragments	DNA or protein	coding sequence	conserved domain
1	G1043	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	43-927	
2	G1043	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		120-179
3	G759	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	8-961	
4	G759	NAM	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		17-159
5	G185	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	77-988	
6	G185	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		113-172
7	G629	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	169-1275	
8	G629	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		92-152
9	G435	HB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	32-502	
10	G435	HB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		4-67
11	G4	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	90-1217	
12	G4	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		121-188
13	G1035	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	103-624	
14	G1035	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		39-91
15	G179	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	68-511	
16	G179	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		65-121
17	G28	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	63-869	
18	G28	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		145-213
19	G1241	MISC	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	108-605	
20	G1241	MISC	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		
21	G19	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	70-816	
22	G19	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		76-145
23	G503	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	80-886	
24	G503	NAM	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		12-158
25	G263	HS	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	48-902	
26	G263	HS	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		15-105
27	G921	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	116-1024	
28	G921	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		146-203
29	G1275	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	58-579	
30	G1275	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		113-169

Figure 1b

SEQ ID No.	GID No.	Family	Fragments		DNA or protein	coding sequence	conserved domain
31	G242	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	66-983	6-105
32	G242	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		
33	G1006	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	52-783	114-182
34	G1006	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		
35	G1049	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	29-550	
36	G1049	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		77-132
37	G502	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	224-1186	
38	G502	NAM	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		10-155
39	G239	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	1-822	
40	G239	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		21-125
41	G555	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	250-1242	
42	G555	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		38-110
43	G352	Z	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	80-817	
44	G352	Z	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		99-119, 166-186
45	G1352	Z	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	79-900	
46	G1352	Z	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		108-129, 167-188
47	G1089	bZIP ¹²	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	31-2427	
48	G1089	bZIP ¹²	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		425-500
49	G553	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	82-1236	
50	G553	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		94-160
51	G1221	MISC	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	287-2314	
52	G1221	MISC	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		490-515
53	G580	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	43-747	
54	G580	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		162-218
55	G270	AKR	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	43-1350	
56	G270	AKR	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		
57	G201	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	1-1011	
58	G201	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		14-114
59	G1417	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400		DNA	32-1501	
60	G1417	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200		protein		239-296

Figure 1c

SEQ ID No	GID No.	Family	Fragments	DNA or protein	coding sequence	conserved domain
61	G233	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	46-867	
62	G233	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		14-114
63	G920	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	114-1154	
64	G920	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		152-211
65	G867	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	64-1098	
66	G867	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		59-124
67	G659	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	1-984	
68	G659	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		16-116
69	G620	CAAT	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	40-666	
70	G620	CAAT	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		20-118
71	G596	AT-Hook	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	168-1121	
72	G596	AT-Hook	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		89-96
73	G511	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	31-738	
74	G511	NAM	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		8-159
75	G471	IAA/ARF	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	115-2112	
76	G471	IAA/ARF	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		22-354
77	G385	HB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	37-2202	
78	G385	HB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		60-123
79	G261	HS	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	458-1663	
80	G261	HS	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		16-104
81	G25	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	80-595	
82	G25	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		47-114
83	G610	BPF-1	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	137-2059	
84	G610	BPF-1	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		577-609
85	G229	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	41-1156	
86	G229	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		14-120
87	G221	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	115-795	
88	G221	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		21-125
89	G186	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	100-1761	
90	G186	WRKY	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		312-369

Figure 1d

SEQ ID No	GID No.	Family	Fragments	DNA or protein	coding sequence	conserved domain
91	G562	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	137-1285	
92	G562	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		253-315
93	G255	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	30-839	
94	G255	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		14-115
95	G3	AP2	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	16-477	
96	G3	AP2	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		11-95
97	G713	HB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	58-765	
98	G713	HB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		23-86
99	G515	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	154-1170	
100	G515	NAM	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		6-144
101	G390	HB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	1-2526	
102	G390	HB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		18-81
103	G1034	bZIP	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	214-1443	
104	G1034	bZIP	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		97-160
105	G1149	PAZ	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	1-2910	
106	G1149	PAZ	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		870-880
107	G1334	CAAT	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	76-885	
108	G1334	CAAT	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		18-190
109	G1650	HLH/MYC	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA	84-1199	
110	G1650	HLH/MYC	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		284-334
111	G241	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
112	G241	MYB	1-50, 50-75, 76-81, 82-100, 100-150, 150-200	protein		14-116
113	G348	GATA Zn	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
114	G171	MADS	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
115	G521	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
116	G1274	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
117	G182	WRKY	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
118	G1290	AKR	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
119	G374	Z	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		
120	G682	MYB	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		

Figure 1e

SEQ ID No	GID No.	Family	Fragments	DNA or protein	coding sequence	conserved domain
121	G501	NAM	1-100, 30-48, 75-125, 150-200, 200-300, 350-400	DNA		

DECLARATION FOR UTILITY PATENT APPLICATION

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name;

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

Disease-induced polynucleotides

the specification of which is attached hereto.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose all information which is material to patentability as defined in Title 37, Code of Federal Regulations, § 1.56(a) which states in relevant part: "Each individual associated with the filing and prosecution of a patent application has a duty of candor and good faith in dealing with the Office, which includes a duty to disclose to the Office all information known to that individual to be material to patentability as defined in this section. The duty to disclose all information known to be material to patentability is deemed to be satisfied if all information known to be material to patentability of any claim issued in a patent was cited by the Office or submitted to the Office in the manner prescribed by §§ 1.97(b)-(d) and 1.98.

I hereby claim foreign priority benefits under Title 35 United States Code, § 119(a)-(d) or 365(a)-(b) of any foreign applications for patent or inventor's certificate as indicated below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

I hereby claim the benefit of priority under Title 35 United States Code, § 119(e) of any United States provisional application(s) listed below:

Provisional Serial No.: Filing Date:
60/125,814 3/23/99

I hereby claim the benefit under Title 35 United States Code, § 120 of any United States applications listed below and, insofar as this is a continuation-in-part application filed under the conditions set forth in 35 United States Code, § 120, which discloses and claims subject matter in addition to the prior copending application(s) listed below, I acknowledge the duty to disclose to the United States Patent Office all information known to be material to patentability as defined in Title 37 Code of Federal Regulations, § 1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Title 18, United States Code, § 1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of first joint coinventor: Jacqueline Heard

Inventor's signature: *Jacqueline Heard*

Date: 3/20/00

Citizenship: U. S.

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Post Office Address: Same as above.

Full name of second joint coinventor: Pierre Broun

Inventor's signature: *P. Broun*

Date: 3-20-00

Citizenship: France

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Post Office Address: Same as above.

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Date: 3/20/2000

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Residence: 115 Moss Avenue, Apt. 308, Oakland, CA 94611

Post Office Address: Same as above.

Full name of fourth joint coinventor: James Keddie

Inventor's signature:

Date: 3-20-2000

Citizenship: U. K.

Residence: 54 McLellan Ave, San Mateo, CA 94403

Post Office Address: Same as above.

Full name of fifth joint coinventor: Omaira Pineda

Inventor's signature:

Date: 3/21/00

Citizenship: Colombia

Residence: 19563 Helen Place, Castro Valley, CA 94546

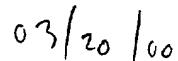
Post Office Address: Same as above.

Full name of sixth joint coinventor: Luc Adam

Inventor's signature:



Date:



Citizenship: Canada

Residence: 25800 Industrial Blvd. Apt. L403, Hayward, CA 94545

Post Office Address: Same as above.

Full name of seventh joint coinventor: Raymond Samaha

Inventor's signature:



Date: 3/20/00

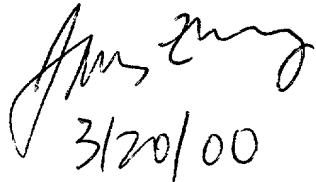
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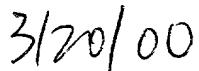
Post Office Address: Same as above.

Full name of eighth joint coinventor: James Zhang

Inventor's signature:



Date:



Citizenship: U. S.

Residence: 951 Amarillo Avenue, Palo Alto CA 94303

Post Office Address: Same as above.

Full name of ninth joint coinventor: Guo-Liang Yu

Inventor's signature:



Date: 03/20/2006

Citizenship: China

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Full name of tenth joint coinventor: Oliver Ratcliffe

Inventor's signature: 

Date: 3.20.00

Citizenship: U. K.

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Post Office Address: Same as above.

Full name of eleventh joint coinventor: Marsha Pilgrim

Inventor's signature: 

Date: 3/20/00

Citizenship: U.S.

Residence: 2200 Emerson Street, Palo Alto, CA 94301

Post Office Address: Same as above.

Full name of twelfth joint coinventor: Cai-Zhong Jiang

Inventor's signature: *Cai-Zhong Jiang*

Date: 3/21/00

Citizenship: China

Residence: 34495 Heathrow Terrace, Fremont, CA 94555

Post Office Address: Same as above.

Full name of thirteenth joint coinventor: Lynne Reuber

Inventor's signature: *Lynne Reuber*

Date: 3/21/00

Citizenship: U.S.

Residence: 1115 S. Grant, San Mateo, CA 94402

Post Office Address: Same as above.

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

IN RE PATENT APPLICATION OF

Jacqueline Heard et al.

Examiner: Unknown

Application No. Unassigned

Group Art Unit: Unknown

Filing Date: Herewith

Title: Disease-induced polynucleotides

POWER OF ATTORNEY BY ASSIGNEE
TO EXCLUSION OF INVENTOR UNDER 37 C.F.R. § 3.71

Commissioner of Patents
and Trademarks
Washington, D.C. 20231

Sir:

The undersigned ASSIGNEE having an interest in the above-identified application for letters patent hereby appoints Karen J. Guerrero, Reg. No. 37,071 to prosecute this application and transact all business in the United States Patent and Trademark Office in connection therewith and hereby revokes all prior powers of attorney; said appointment to be to the exclusion of the inventors and the inventors' attorneys in accordance with the provisions of 37 C.F.R. § 3.71.

The following evidentiary documents establish a chain of title from the original owner to the Assignee:

X a copy of an Assignment attached hereto, which Assignment has been (or is herewith) forwarded to the Patent and Trademark Office for recording; or

- the Assignment recorded on _____ at reel __, frames __ - __.

Pursuant to 37 C.F.R. § 3.73(b) the undersigned Assignee hereby states that evidentiary documents have been reviewed and hereby certifies that, to the best of ASSIGNEE's knowledge and belief, title is in the identified ASSIGNEE.

Direct all telephone calls to Karen J. Guerrero (510) 264-0280 ext. 125.

Address all correspondence to:

Karen J. Guerrero
MENDEL BIOTECHNOLOGY, INC.
21375 Cabot Boulevard
Hayward, California 94545

ASSIGNEE: Mendel Biotechnology, Inc.

Name: 

Name: Guo-Liang Yu

Title: Senior Vice-President, Research and Development

Date: 03/20/2000

SEQUENCE LISTING

<110> Heard, Jacqueline
Broun, Pierre
Riechmann, Jose-Luis
Keddie, James
Pineda, Omaira
Adam, Luc
Samaha, Raymond
Zhang, James
Yu, Guo-Liang
Ratcliffe, Oliver
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 gtggagttca aaagtcagaa ttgcccctgg agctgggtttt gatagaacgc tggacgatgg 420
 attcagttgg agaaaagtacg gccagaagga tattctcgga gccaaatttc caagaggata 480
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 ttagtggaaat cagatgtcc ttgagatcgtt accatgttctt gctctcaagc 600
 tgcaaattgtc ggtacaacaa tgccgatataaaacccctcgaa cccgaaaccaga cccaaagaaca 660
 cggaaatctt gacatggtaa aggaaagtgt agacaactac aatcaccacaa cacattgca 720
 tcacaacccctt cactatccat tgcatactac cccaaatcta gagaataaca atgccttat 780
 gcttcaaattgcgatcaaa acatcgaata ttttggatctt acgagcttctt ctagtgcatt 840
 aggaacttagt atcaactaca atttccacgatcttccgtcg gcttcttactt cagcatcaaa 900
 ctctccgtcc accgtccctt tggaaatcccc gtttggaaagc tatgatccaa atcatccata 960
 tggaggattt ggtgggttctt attcttagttt atctacttaa gggagggacg gaactttta 1020
 catgacccctt tgattaaaga gagagtttc ataatacgta atcaatttcc tattcaaata 1080
 tccgagttttt ttttctaaatc atgtttatca attgtttat tacagaaggc ttattttcag 1140
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 aaaaa 1205

<210> 6

<211> 303

<212> PRT

<213> Arabidopsis thaliana

<220>

<223> G185

<400> 6

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 20 25 30

Ser Ala Ser Ser Gln Glu Thr Arg Glu His Leu Ala Lys Lys Ile Leu
 35 40 45

Gln Ser Tyr His Lys Ser Leu Thr Ile Met Asn Tyr Ser Gly Glu Leu
 50 55 60

Asp Gln Val Ser Gln Gly Gly Ser Pro Lys Ser Asp Asp Ser Asp
 65 70 75 80

Gln Glu Pro Leu Val Ile Lys Ser Ser Lys Lys Ser Met Pro Arg Trp
 85 90 95

Ser Ser Lys Val Arg Ile Ala Pro Gly Ala Gly Val Asp Arg Thr Leu
 100 105 110

Asp Asp Gly Phe Ser Trp Arg Lys Tyr Gly Gln Lys Asp Ile Leu Gly
 115 120 125
 Ala Lys Phe Pro Arg Gly Tyr Tyr Arg Cys Thr Tyr Arg Lys Ser Gln
 130 135 140
 Gly Cys Glu Ala Thr Lys Gln Val Gln Arg Ser Asp Glu Asn Gln Met
 145 150 155 160
 Leu Leu Glu Ile Ser Tyr Arg Gly Ile His Ser Cys Ser Gln Ala Ala
 165 170 175
 Asn Val Gly Thr Thr Met Pro Ile Gln Asn Leu Glu Pro Asn Gln Thr
 180 185 190
 Gln Glu His Gly Asn Leu Asp Met Val Lys Glu Ser Val Asp Asn Tyr
 195 200 205
 Asn His Gln Ala His Leu His His Asn Leu His Tyr Pro Leu Ser Ser
 210 215 220
 Thr Pro Asn Leu Glu Asn Asn Ala Tyr Met Leu Gln Met Arg Asp
 225 230 235 240
 Gln Asn Ile Glu Tyr Phe Gly Ser Thr Ser Phe Ser Ser Asp Leu Gly
 245 250 255
 Thr Ser Ile Asn Tyr Asn Phe Pro Ala Ser Gly Ser Ala Ser His Ser
 260 265 270
 Ala Ser Asn Ser Pro Ser Thr Val Pro Leu Glu Ser Pro Phe Glu Ser
 275 280 285
 Tyr Asp Pro Asn His Pro Tyr Gly Gly Phe Gly Phe Tyr Ser
 290 295 300

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 <211> 1557
 <212> DNA
 <213> *Arabidopsis thaliana*

<220>
 <223> G629

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 ttaagggcc agcttggatc tctataactg aagggatat atatagat gatggatct 180
 tcttcctccaa cacaacttgc atctttttaaga gacatggaa tctatggatc atttcaacaa 240
 attgtcggtt gggaaatgt tttcaaatct gatatcaatg atcatatgtcc caataactgct 300
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 aattatgatt cttctcataa ccagatcgaa gcagaacaac cttcttagtaa tgataatcaa 420
 gatgtatgtc gcaggattca tgataagatg aaacggcggtt tagcgcagaa ccgagaagcg 480
 gctcgcaaaa gtcgttgag aaagaaggct tatgttcagc agtttagagga aagccgggtt 540
 aagttatcgc agtttagagca agaactcgaa aaggttaagc agcaggccca ttttaggatca 600
 tctggagta ttaacacagg gattgcatca tttgagatgg aatattcaca ctggctacaa 660

gaacaaagca gaagagttag cgaactacga acagcgcttc aatctcatat aagcgacata 720
 gaactcaaga tgcttaga gagttgcttg aaccattacg ctaatcttt ccgaatgaaa 780
 tccgatgcag caaaagccga tgtttctac ttgatatcggaatgtggcg aacttcaacc 840
 gaaagattct tccaatggat tggagggttt cgtccatccg aactttaaa cgttgtatg 900
 ccttatcttc aaccattaac ggatcaacaa atcttggaaag tgagaaacact ccaacaatca 960
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 gaaagcattg tgattgatgc ggttatcgag tccacgcatt atcccactca catggctgca 1080
 gctatagaga atcttcaaggc attagaagga tttgtgaatc aagcagatca tctgaggcaa 1140
 caaacttgc aacaatggc gaagatctt acgacaagac aatcggctcg aggttacta 1200
 gctttaggag agtacattca tagacttcgt gctcttagtt ctctttggc agctcgtcca 1260
 caagaaccaa cttaaaagag gaacttatta aaacttaaa aacaagaaac agcagaatca 1320
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 gtttaattaag tagagtgaga ttctctttagt tagaacttta tggttttgc tttatgaagt 1440
 atctctccag agaagattgt aaatttgggt tgaaactttg taatatatta agatccacca 1500
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<210> 8

<211> 368

<212> PRT

<213> Arabidopsis thaliana

<220>

<223> G629

<400> 8

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								20				30			

Lys	Ser	Asp	Ile	Asn	Asp	His	Ser	Pro	Asn	Thr	Ala	Thr	Ser	Ser	Ile
								35				45			

Ile	Gln	Val	Asp	Pro	Arg	Ile	Asp	Asp	His	Asn	Asn	Ile	Lys	Ile
								50				60		

Asn	Tyr	Asp	Ser	Ser	His	Asn	Gln	Ile	Glu	Ala	Glu	Gln	Pro	Ser	Ser
								65				75			80

Asn	Asp	Asn	Gln	Asp	Asp	Gly	Arg	Ile	His	Asp	Lys	Met	Lys	Arg
								85				90		95

Arg	Leu	Ala	Gln	Asn	Arg	Glu	Ala	Ala	Arg	Lys	Ser	Arg	Leu	Arg	Lys
								100				105		110	

Lys	Ala	Tyr	Val	Gln	Gln	Leu	Glu	Glu	Ser	Arg	Leu	Lys	Leu	Ser	Gln
								115				120		125	

Leu	Glu	Gln	Glu	Leu	Glu	Lys	Val	Lys	Gln	Gln	Gly	His	Leu	Gly	Pro
								130				135		140	

Ser	Gly	Ser	Ile	Asn	Thr	Gly	Ile	Ala	Ser	Phe	Glu	Met	Glu	Tyr	Ser
								145				155		160	

His	Trp	Leu	Gln	Glu	Gln	Ser	Arg	Arg	Val	Ser	Glu	Leu	Arg	Thr	Ala
								165				170		175	

Leu Gln Ser His Ile Ser Asp Ile Glu Leu Lys Met Leu Val Glu Ser
 180 185 190
 Cys Leu Asn His Tyr Ala Asn Leu Phe Arg Met Lys Ser Asp Ala Ala
 195 200 205
 Lys Ala Asp Val Phe Tyr Leu Ile Ser Gly Met Trp Arg Thr Ser Thr
 210 215 220
 Glu Arg Phe Phe Gln Trp Ile Gly Gly Phe Arg Pro Ser Glu Leu Leu
 225 230 235 240
 Asn Val Val Met Pro Tyr Leu Gln Pro Leu Thr Asp Gln Gln Ile Leu
 245 250 255
 Glu Val Arg Asn Leu Gln Gln Ser Ser Gln Gln Ala Glu Asp Ala Leu
 260 265 270
 Ser Gln Gly Ile Asp Lys Leu Gln Gln Ser Leu Ala Glu Ser Ile Val
 275 280 285
 Ile Asp Ala Val Ile Glu Ser Thr His Tyr Pro Thr His Met Ala Ala
 290 295 300
 Ala Ile Glu Asn Leu Gln Ala Leu Glu Gly Phe Val Asn Gln Ala Asp
 305 310 315 320
 His Leu Arg Gln Gln Thr Leu Gln Gln Met Ala Lys Ile Leu Thr Thr
 325 330 335
 Arg Gln Ser Ala Arg Gly Leu Leu Ala Leu Gly Glu Tyr Leu His Arg
 340 345 350
 Leu Arg Ala Leu Ser Ser Leu Trp Ala Ala Arg Pro Gln Glu Pro Thr
 355 360 365

<210> 9
 <211> 627
 <212> DNA
 <213> *Arabidopsis thaliana*

<220>
 <223> G435

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 caagaagctt gagccagatc tgaaacttca actgtcgaac cagcttgcac tacctcaaag 180
 acaagtcgtt gtctggttcc aaaacaagcg agccagggtc aagactcagt ctcttgagg 240
 ccaacactgc actcttcagt ccaagcacga agcagctctc tccgacaagg caaagttaga 300
 gcatcaagtg cagttctcc aagatgagct gaagagagca aggaatcagc ttgctctgtt 360
 cacaaatcaa gatttcctcg ttgataattc taatcttggt tcttgtatg aagatcatga 420
 tgatcaagtg gtggatttcg acgagctta cgcttgctt gtttagcaatg gacatggatc 480
 ttcatcaacc tcatgggtct gattctgtt cgacgcagac aagattccaa tatatatagt 540
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 taattaaagt cattcagaca ttcacta 627

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 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G435

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 20 25 30
 Leu Glu Pro Asp Leu Lys Leu Gln Leu Ser Asn Gln Leu Gly Leu Pro
 35 40 45
 Gln Arg Gln Val Ala Val Trp Phe Gln Asn Lys Arg Ala Arg Phe Lys
 50 55 60
 Thr Gln Ser Leu Glu Val Gln His Cys Thr Leu Gln Ser Lys His Glu
 65 70 75 80
 Ala Ala Leu Ser Asp Lys Ala Lys Leu Glu His Gln Val Gln Phe Leu
 85 90 95
 Gln Asp Glu Leu Lys Arg Ala Arg Asn Gln Leu Ala Leu Phe Thr Asn
 100 105 110
 Gln Asp Ser Pro Val Asp Asn Ser Asn Leu Gly Ser Cys Asp Glu Asp
 115 120 125
 His Asp Asp Gln Val Val Phe Asp Glu Leu Tyr Ala Cys Phe Val
 130 135 140
 Ser Asn Gly His Gly Ser Ser Ser Thr Ser Trp Val
 145 150 155

<210> 11
 <211> 1577
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G4

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 aaaaagagta gagcttcgt gaagccacca tgtgtggagg agctataatc tccgatttca 120
 tacctccggc gaggtccctc cgcgtcacta acgagttat ctggccgat ctgaaaaaca 180
 aagtgaaagc ttcaaagaag agatcgaata agcgatccga ttcttcgtat cttgacgatg 240
 atttcgaagc tgattccaa gggtttaagg atgactcgcc ttttgactgc gaagacgatg 300
 atgatgtttt cgtcaatgtt aagccttcg tcttcaccgc aactactaag cccgttagctt 360
 ccgcttcgtt ctccactgta ggttcagcat atgccaagaa aactgttagag tccgctgagc 420

aagctgagaa atcttctaag aggaagagga agaatcaata ccgagggatt aggca cgctc 480
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 gaacattcga cactgctgag gaagcagcaa gagcttatga tgctgcagca cgcagaatcc 600
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 aacgtcctag tgctaagact aataatctt agaaatcagt ggctaaacca aacaaaagcg 720
 taacttttgt tcagcagcca acacatctga gtcagcagta ctgcaacaac tccttgaca 780
 actctttgg tcatatgagt ttcatgaa agaaggctca gatgtacaac aatcagttt 840
 ggttaacaaa ctcgttcgt gctggaggta acaatggata ccagtattc agttccgatc 900
 agggcagtaa ctccttcgac tgttctgagt tcgggtggag tgatcacgac cctaaaacac 960
 ccgagatctc ttcaatgctt gtcaataaca acgaagcatc atttggtaa gaaaccaatg 1020
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 atttctttc tatttaaaag acagtttatt agtctctga gctctttt tgatctttgt 1500
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 ataataaaagt ctcttg 1577

<210> 12
 <211> 375
 <212> PRT
 <213> *Arabidopsis thaliana*

<220>
 <223> G4

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 20 25 30
 Lys Ala Ser Lys Lys Arg Ser Asn Lys Arg Ser Asp Phe Phe Asp Leu
 35 40 45
 Asp Asp Asp Phe Glu Ala Asp Phe Gln Gly Phe Lys Asp Asp Ser Ala
 50 55 60
 Phe Asp Cys Glu Asp Asp Asp Val Phe Val Asn Val Lys Pro Phe
 65 70 75 80
 Val Phe Thr Ala Thr Thr Lys Pro Val Ala Ser Ala Phe Val Ser Thr
 85 90 95
 Val Gly Ser Ala Tyr Ala Lys Lys Thr Val Glu Ser Ala Glu Gln Ala
 100 105 110
 Glu Lys Ser Ser Lys Arg Lys Asn Gln Tyr Arg Gly Ile Arg
 115 120 125
 Gln Arg Pro Trp Gly Lys Trp Ala Ala Glu Ile Arg Asp Pro Arg Lys
 130 135 140

Gly Ser Arg Glu Trp Leu Gly Thr Phe Asp Thr Ala Glu Glu Ala Ala
 145 150 155 160
 Arg Ala Tyr Asp Ala Ala Ala Arg Arg Ile Arg Gly Thr Lys Ala Lys
 165 170 175
 Val Asn Phe Pro Glu Glu Lys Asn Pro Ser Val Val Ser Gln Lys Arg
 180 185 190
 Pro Ser Ala Lys Thr Asn Asn Leu Gln Lys Ser Val Ala Lys Pro Asn
 195 200 205
 Lys Ser Val Thr Leu Val Gln Gln Pro Thr His Leu Ser Gln Gln Tyr
 210 215 220
 Cys Asn Asn Ser Phe Asp Asn Ser Phe Gly Asp Met Ser Phe Met Glu
 225 230 235 240
 Glu Lys Pro Gln Met Tyr Asn Asn Gln Phe Gly Leu Thr Asn Ser Phe
 245 250 255
 Asp Ala Gly Gly Asn Asn Gly Tyr Gln Tyr Phe Ser Ser Asp Gln Gly
 260 265 270
 Ser Asn Ser Phe Asp Cys Ser Glu Phe Gly Trp Ser Asp His Gly Pro
 275 280 285
 Lys Thr Pro Glu Ile Ser Ser Met Leu Val Asn Asn Asn Glu Ala Ser
 290 295 300
 Phe Val Glu Glu Thr Asn Ala Ala Lys Lys Leu Lys Pro Asn Ser Asp
 305 310 315 320
 Glu Ser Asp Asp Leu Met Ala Tyr Leu Asp Asn Ala Leu Trp Asp Thr
 325 330 335
 Pro Leu Glu Val Glu Ala Met Leu Gly Ala Asp Ala Gly Ala Val Thr
 340 345 350
 Gln Glu Glu Glu Asn Pro Val Glu Leu Trp Ser Leu Asp Glu Ile Asn
 355 360 365
 Phe Met Leu Glu Gly Asp Phe
 370 375

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 <212> DNA
 <213> *Arabidopsis thaliana*

<220>
 <223> G1035

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 atgggatctt ccacaagtgg aaattgctcg tcggttcaa ccactggttt agctaactcc 180

ggttcagaat ctgatctccg gcaacgtgat ctaatcgacg agcggaaagag aaagaggaaa 240
 cagtcgaaca gagaatctgc gaggaggtcg aggatgagga agcagaagca tttggatgat 300
 ctcactgctc aggtgactca tctacgtaaa gaaaacgctc agatcgctc cggaatcgcc 360
 gtcacgacgc agcactacgt cactatcgag gcggagaacg acattctcg agctcagggt 420
 cttgaactta accaccgtct ccaatctt aacgagatcg ttgatttcgt cgaatcttct 480
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 gtgatgaatc ctagaatct agggttttat aatcaaccaa tcatggctc tgcttctact 600
 gctggtgatg ttttcaactg ttagaaaact tcacatcatt atcatcgta gtgagactaa 660
 tcatcgacgc aggggtaaaaa ctgtaatttt tcttataaaat tatgtgatga tgcttgggt 720
 ctttatttta taagatggtt aattagtgtt taaaactgat tgtaatgata gacagtgtaa 780
 gaaatgtgtg atatcatgga gatggtgatg tgagtttggt acaaataattt taagatctt 840
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<210> 14
 <211> 173
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G1035

<400> 14
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 Arg Gln Arg Asp Leu Ile Asp Glu Arg Lys Arg Lys Arg Lys Gln Ser
 35 40 45
 Asn Arg Glu Ser Ala Arg Arg Ser Arg Met Arg Lys Gln Lys His Leu
 50 55 60
 Asp Asp Leu Thr Ala Gln Val Thr His Leu Arg Lys Glu Asn Ala Gln
 65 70 75 80
 Ile Val Ala Gly Ile Ala Val Thr Thr Gln His Tyr Val Thr Ile Glu
 85 90 95
 Ala Glu Asn Asp Ile Leu Arg Ala Gln Val Leu Glu Leu Asn His Arg
 100 105 110
 Leu Gln Ser Leu Asn Glu Ile Val Asp Phe Val Glu Ser Ser Ser
 115 120 125
 Gly Phe Gly Met Glu Thr Gly Gln Gly Leu Phe Asp Gly Gly Leu Phe
 130 135 140
 Asp Gly Val Met Asn Pro Met Asn Leu Gly Phe Tyr Asn Gln Pro Ile
 145 150 155 160
 Met Ala Ser Ala Ser Thr Ala Gly Asp Val Phe Asn Cys
 165 170

<210> 15
 <211> 724
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G179

<400> 15
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 tcgcttgaca gagtttcatg gggtcgacaa ctctgctcag ccgacaacat catccgaaga 180
 gaagccaagg agtaagaaga agaagaaaga gagagaagcg aggtacggt tccagacaag 240
 aagccaggtt gatatactgg atgatggata caggtggagg aagtacggcc aaaaagcagt 300
 caagaacaat ccattccca ggagctatta taagtgcaca gaagaaggat gcagagtgaa 360
 gaagcaagtg cagaggcaat ggggagacga aggagtggtg gtgacgacat accaagggtgt 420
 tcatacacat gccgttgata aaccctctga taatttccac cacatcttga cacaatgca 480
 catctccct ccctttgtt tgaaggaatg attagaggaa ttggatttta atatttactt 540
 tcccaaaaac gttgggctca caccatcaga ccttacttt taaacttagca gcaactcaca 600
 tatctcaaaa atactaatcc ttatctttgt ctttatgggaa ctttgaatc catctgctt 660
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 taaa 724

<210> 16
 <211> 147
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G179

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 Thr Thr Ser Ser Glu Glu Lys Pro Arg Ser Lys Lys Lys Lys Glu
 35 40 45
 Arg Glu Ala Arg Tyr Ala Phe Gln Thr Arg Ser Gln Val Asp Ile Leu
 50 55 60
 Asp Asp Gly Tyr Arg Trp Arg Lys Tyr Gly Gln Lys Ala Val Lys Asn
 65 70 75 80
 Asn Pro Phe Pro Arg Ser Tyr Tyr Lys Cys Thr Glu Glu Gly Cys Arg
 85 90 95
 Val Lys Lys Gln Val Gln Arg Gln Trp Gly Asp Glu Gly Val Val Val
 100 105 110
 Thr Thr Tyr Gln Gly Val His Thr His Ala Val Asp Lys Pro Ser Asp
 115 120 125

Asn Phe His His Ile Leu Thr Gln Met His Ile Phe Pro Pro Phe Cys
 130 135 140

Leu Lys Glu
 145

<210> 17
 <211> 964
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G28

<400> 17
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 acttactagg agaatcggag ccgatactca gtgagtcgac agcgagttcg gttactcaat 180
 cttgtgtAAC cggtagagc attaaaccgg tgcacggacg aaacccttagc tttagcaaac 240
 tgcataccttg ctccaccggag agctggggag atttgcgtt gaaagaaaaac gattctgagg 300
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 cgtctccga cgaagatcg agctcttcc cgagtgtaa gatcgagact ccggagagtt 420
 tcgcggcggt ggattctgtt ccggtaaga aggagaagac gagtcctgtt tcggcggcg 480
 tgcggcggtc gaaggaaag cattatagag gagtgagaca aaggccgtgg gggaaatttg 540
 cggcggagat tagagatccg gcgaagaacg gagctagggt ttggtagga acgtttgaga 600
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 ccaagagatc ttcttttctt tottctaaccg agaacggacg tccgaagaag aggagaacgg 780
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 cacgtggcga tcgttattt gtttataat tttgattttt ctttggtaa tgattatatg 900
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 964
 aaaa

<210> 18
 <211> 268
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G28

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Ile Arg Arg His Leu Leu Gly Glu Ser Glu Pro Ile Leu Ser Glu Ser
 20 25 30

Thr Ala Ser Ser Val Thr Gln Ser Cys Val Thr Gly Gln Ser Ile Lys
 35 40 45

Pro Val Tyr Gly Arg Asn Pro Ser Phe Ser Lys Leu Tyr Pro Cys Phe
 50 55 60

Thr Glu Ser Trp Gly Asp Leu Pro Leu Lys Glu Asn Asp Ser Glu Asp
 65 70 75 80

Met	Leu	Val	Tyr	Gly	Ile	Leu	Asn	Asp	Ala	Phe	His	Gly	Gly	Trp	Glu
					85				90					95	
Pro	Ser	Ser	Ser	Ser	Ser	Asp	Glu	Asp	Arg	Ser	Ser	Phe	Pro	Ser	Val
						100			105				110		
Lys	Ile	Glu	Thr	Pro	Glu	Ser	Phe	Ala	Ala	Val	Asp	Ser	Val	Pro	Val
						115			120				125		
Lys	Lys	Glu	Lys	Thr	Ser	Pro	Val	Ser	Ala	Ala	Val	Thr	Ala	Ala	Lys
						130			135				140		
Gly	Lys	His	Tyr	Arg	Gly	Val	Arg	Gln	Arg	Pro	Trp	Gly	Lys	Phe	Ala
						145			150				155		160
Ala	Glu	Ile	Arg	Asp	Pro	Ala	Lys	Asn	Gly	Ala	Arg	Val	Trp	Leu	Gly
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						180			185				190		
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 35 40 45
 Asn Ser Ile Pro Ala Ile Glu Glu Val Asn Ile Phe Lys Asp Asp Val
 50 55 60
 Val Ile Gln Phe Ile Asn Pro Lys Val Gln Ala Ser Ile Ala Ala Asn
 65 70 75 80
 Thr Trp Val Val Ser Gly Thr Pro Gln Thr Lys Lys Leu Gln Asp Ile
 85 90 95
 Leu Pro Gln Ile Ile Ser Gln Leu Gly Pro Asp Asn Leu Asp Asn Leu
 100 105 110
 Arg Lys Leu Ala Glu Gln Phe Gln Lys Gln Ala Pro Gly Ala Gly Asp
 115 120 125
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 35 40 45

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Gln Ala Thr Glu Pro Gly Lys Arg Arg Lys Arg Lys Asn Val Tyr Arg
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Gly Ile Arg Lys Arg Pro Trp Gly Lys Trp Ala Ala Glu Ile Arg Asp
85 90 95

Pro Arg Lys Gly Val Arg Val Trp Leu Gly Thr Phe Asn Thr Ala Glu
100 105 110

Glu Ala Ala Met Ala Tyr Asp Val Ala Ala Lys Gln Ile Arg Gly Asp
115 120 125

Lys Ala Lys Leu Asn Phe Pro Asp Leu His His Pro Pro Pro Pro Asn
 100 125 140

Tyr Thr Pro Pro Pro Ser Ser Pro Arg Ser Thr Asp Gln Pro Pro Ala
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Lys Lys Val Cys Val Val Ser Gln Ser Glu Ser Glu Leu Ser Gln Pro
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 Asn Leu Ser Tyr Gly Phe Glu Pro Asp Tyr Asp Leu Lys Gln Gln Ile
 195 200 205
 Ser Ser Leu Glu Ser Phe Leu Glu Leu Asp Gly Asn Thr Ala Glu Gln
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 35 40 45

Phe Asp Pro Trp Gln Leu Pro Glu Lys Thr Glu Phe Gly Glu Asn Glu
 50 55 60

Trp Tyr Phe Phe Ser Pro Arg Glu Arg Lys Tyr Pro Asn Gly Val Arg
 65 70 75 80

Pro Asn Arg Ala Ala Val Ser Gly Tyr Trp Lys Ala Thr Gly Thr Asp
 85 90 95

Lys Ala Ile His Ser Gly Ser Ser Asn Val Gly Val Lys Lys Ala Leu
 100 105 110

Val Phe Tyr Lys Gly Arg Pro Pro Lys Gly Ile Lys Thr Asp Trp Ile
 115 120 125

Met His Glu Tyr Arg Leu His Asp Ser Arg Lys Ala Ser Thr Lys Arg
 130 135 140

Ser Gly Ser Met Arg Leu Asp Glu Trp Val Leu Cys Arg Ile Tyr Lys
 145 150 155 160

Lys Arg Gly Ala Ser Lys Leu Leu Asn Glu Gln Glu Gly Phe Met Asp
 165 170 175

Glu Val Leu Met Glu Asp Glu Thr Lys Val Val Ile Asn Glu Ala Glu
 180 185 190

Arg Arg Asn Asp Glu Glu Ile Met Met Met Thr Ser Met Lys Leu Pro
 195 200 205

Arg Thr Cys Ser Leu Ala His Leu Leu Glu Met Asp Tyr Met Gly Pro
 210 215 220

Val Ser His Ile Asp Asn Phe Ser Gln Phe Asp His Leu His Gln Pro
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Asp Ser Glu Ser Ser Trp Phe Gly Asp Leu Gln Phe Asn Gln Asp Glu
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 35 40 45

Phe Ala Lys Asp Leu Leu Pro Gln Tyr Phe Lys His Asn Asn Phe Ser
 50 55 60

Ser Phe Ile Arg Gln Leu Asn Thr Tyr Gly Phe Arg Lys Thr Val Pro
 65 70 75 80

Asp Lys Trp Glu Phe Ala Asn Asp Tyr Phe Arg Arg Gly Glu Asp
 85 90 95

Leu Leu Thr Asp Ile Arg Arg Lys Ser Val Ile Ala Ser Thr Ala
 100 105 110

Gly Lys Cys Val Val Val Gly Ser Pro Ser Glu Ser Asn Ser Gly Gly
 115 120 125

Gly Asp Asp His Gly Ser Ser Thr Ser Ser Pro Gly Ser Ser Lys
 130 135 140

Asn Pro Gly Ser Val Glu Asn Met Val Ala Asp Leu Ser Gly Glu Asn
 145 150 155 160
 Glu Lys Leu Lys Arg Glu Asn Asn Asn Leu Ser Ser Glu Leu Ala Ala
 165 170 175
 Ala Lys Lys Gln Arg Asp Glu Leu Val Thr Phe Leu Thr Gly His Leu
 180 185 190
 Lys Val Arg Pro Glu Gln Ile Asp Lys Met Ile Lys Gly Gly Lys Phe
 195 200 205
 Lys Pro Val Glu Ser Asp Glu Glu Ser Glu Cys Glu Gly Cys Asp Gly
 210 215 220
 Gly Gly Gly Ala Glu Glu Gly Val Gly Glu Gly Leu Lys Leu Phe Gly
 225 230 235 240
 Val Trp Leu Lys Gly Glu Arg Lys Lys Arg Asp Arg Asp Glu Lys Asn
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 sequence may be A, T, C, G, other or unknown

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 35 40 45
 Glu Met Leu Thr Leu Met Cys Asp Asn Tyr Asn Val Leu Arg Lys Gln
 50 55 60
 Leu Met Glu Tyr Val Asn Lys Ser Asn Ile Thr Glu Arg Asp Gln Ile
 65 70 75 80
 Ser Pro Pro Lys Lys Arg Lys Ser Pro Ala Arg Glu Asp Ala Phe Ser
 85 90 95
 Cys Ala Val Ile Gly Gly Val Ser Glu Ser Ser Ser Thr Asp Gln Asp
 100 105 110
 Glu Tyr Leu Cys Lys Lys Gln Arg Glu Glu Thr Val Val Lys Glu Lys
 115 120 125
 Val Ser Arg Val Tyr Tyr Lys Thr Glu Ala Ser Asp Thr Thr Leu Val
 130 135 140
 Val Lys Asp Gly Tyr Gln Trp Arg Lys Tyr Gly Gln Lys Val Thr Arg
 145 150 155 160
 Asp Asn Pro Ser Pro Arg Ala Tyr Phe Lys Cys Ala Cys Ala Pro Ser
 165 170 175
 Cys Ser Val Lys Lys Lys Val Gln Arg Ser Val Glu Asp Gln Ser Val
 180 185 190
 Leu Val Ala Thr Tyr Glu Gly Glu His Asn His Pro Met Pro Ser Gln
 195 200 205
 Ile Asp Ser Asn Asn Gly Leu Asn Arg His Ile Ser His Gly Gly Ser
 210 215 220
 Ala Ser Thr Pro Val Ala Ala Asn Arg Arg Ser Ser Leu Thr Val Pro
 225 230 235 240

Val Thr Thr Val Asp Met Ile Glu Ser Lys Lys Val Thr Ser Pro Thr
 245 250 255

Ser Arg Ile Asp Phe Pro Gln Val Gln Lys Leu Leu Val Glu Gln Met
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 35 40 45

Tyr Gln Thr Ser Asp Val Ala Gly Ala Leu Phe Ser Gly Ser Ser Ser
 50 55 60

Cys Phe Ser His Pro Glu Ser Pro Ser Thr Lys Thr Tyr Val Ala Ala
 65 70 75 80
 Thr Ala Thr Ala Ser Ala Asp Asn Gln Asn Lys Lys Glu Lys Lys Lys
 85 90 95
 Ile Lys Gly Arg Val Ala Phe Lys Thr Arg Ser Glu Val Glu Val Leu
 100 105 110
 Asp Asp Gly Phe Lys Trp Arg Lys Tyr Gly Lys Lys Met Val Lys Asn
 115 120 125
 Ser Pro His Pro Arg Asn Tyr Tyr Lys Cys Ser Val Asp Gly Cys Pro
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245	250
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 Ile Thr Arg His Leu Leu Gly Gly Gly Glu Asn Glu Leu Arg Leu
 20 25 30
 Asn Glu Ser Thr Pro Ser Ser Cys Phe Thr Glu Ser Trp Gly Gly Leu
 35 40 45
 Pro Leu Lys Glu Asn Asp Ser Glu Asp Met Leu Val Tyr Gly Leu Leu
 50 55 60
 Lys Asp Ala Phe His Phe Asp Thr Ser Ser Asp Leu Ser Cys Leu
 65 70 75 80

Phe Asp Phe Pro Ala Val Lys Val Glu Pro Thr Glu Asn Phe Thr Ala
 85 90 95
 Met Glu Glu Lys Pro Lys Lys Ala Ile Pro Val Thr Glu Thr Ala Val
 100 105 110
 Lys Ala Lys His Tyr Arg Gly Val Arg Gln Arg Pro Trp Gly Lys Phe
 115 120 125
 Ala Ala Glu Ile Arg Asp Pro Ala Lys Asn Gly Ala Arg Val Trp Leu
 130 135 140
 Gly Thr Phe Glu Thr Ala Glu Asp Ala Ala Leu Ala Tyr Asp Ile Ala
 145 150 155 160
 Ala Phe Arg Met Arg Gly Ser Arg Ala Leu Leu Asn Phe Pro Leu Arg
 165 170 175
 Val Asn Ser Gly Glu Pro Asp Pro Val Arg Ile Thr Ser Lys Arg Ser
 180 185 190
 Ser Ser Ser Ser Ser Ser Ser Ser Thr Ser Ser Ser Glu Asn
 195 200 205
 Gly Lys Leu Lys Arg Arg Arg Lys Ala Glu Asn Leu Thr Ser Glu Val
 210 215 220
 Val Gln Val Lys Cys Glu Val Gly Asp Glu Thr Arg Val Asp Glu Leu
 225 230 235 240
 Leu Val Ser

<210> 35
 <211> 725
 <212> DNA
 <213> *Arabidopsis thaliana*

<220>
 <223> G1049

<400> 35
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 gccatttcca accaacggtc aaaaccgta cctcctctac ggattccaaa gcccataaaa 180
 caatccacaa tccatgagcc taagcagcaa caactcaaca tcagatgaag cagaagagca 240
 gcagacgaac aacaatataa tcaacgagcg gaagcagaga aggatgattt caaaccgaga 300
 atccgcaagg agatcgcgta tgaggaagca aagacacctt gacgagctt ggtcacaagt 360
 gatgtggta aggatcgaga atcatcagtt gcttgataag cttacaatc tctctgagtc 420
 tcacgacaag gttcttcaag agaatgctca gcttaaagaa gaaacatttgc agcttaagca 480
 agtcatcgc gatatgcaaa ttcaagccc tttcttgc ttttagagacg atataatccc 540
 cattgaataa agcatttttc cccgattcat atttatgaaa attttcttca agagtatgtt 600
 tctttgtatg tatatgtgga gatgtatttc agggtttga taatatgacc ctttacgacg 660
 acgttttag attgttagtaa atttataaac taaagaagat tagtgttaat gaagaacaaa 720
 tataa 725

<210> 36
 <211> 173
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G1049

<400> 36
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 Ser Ile Leu Gln Ser Pro Tyr Pro Ser Asn Phe Pro Ile Ser Thr Pro
 20 25 30
 Phe Pro Thr Asn Gly Gln Asn Pro Tyr Leu Leu Tyr Gly Phe Gln Ser
 35 40 45
 Pro Thr Asn Asn Pro Gln Ser Met Ser Leu Ser Ser Asn Asn Ser Thr
 50 55 60
 Ser Asp Glu Ala Glu Glu Gln Gln Thr Asn Asn Asn Ile Ile Asn Glu
 65 70 75 80
 Arg Lys Gln Arg Arg Met Ile Ser Asn Arg Glu Ser Ala Arg Arg Ser
 85 90 95
 Arg Met Arg Lys Gln Arg His Leu Asp Glu Leu Trp Ser Gln Val Met
 100 105 110
 Trp Leu Arg Ile Glu Asn His Gln Leu Leu Asp Lys Leu Asn Asn Leu
 115 120 125
 Ser Glu Ser His Asp Lys Val Leu Gln Glu Asn Ala Gln Leu Lys Glu
 130 135 140
 Glu Thr Phe Glu Leu Lys Gln Val Ile Ser Asp Met Gln Ile Gln Ser
 145 150 155 160
 Pro Phe Ser Cys Phe Arg Asp Asp Ile Ile Pro Ile Glu
 165 170

<210> 37
 <211> 1409
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G502

<400> 37
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 cgcgtccgaa ttgatttagga taggatcagg atcatcctca acaacctctt cctaattctt 180
 cctccattca tagtaacaat aatattaaga aagaggtaa actatgtcag aattattaca 240
 gttgcctcca ggttccgat ttcaccctac cgatgaagag cttgtcatgc actatctctg 300
 ccgcaaatgt gcctctcagt ccatcgccgt tccgatcatc gctgagatcg atctctacaa 360

atacgatcca tgggagcttc ctggtttagc cttgtatggt gagaaggaat ggtacttctt 420
 ctctcccagg gacagaaaat atcccaacgg ttgcgtcct aaccggcgg ctgggtctgg 480
 ttactggaaa gctaccggag ctgataaacc gatcggacta cctaaaccgg tcggaattaa 540
 gaaagcttt gttttctacg cccgcaaaagc tccaaaggga gagaaaaacca attggatcat 600
 gcacgagtac cgtctcgccg acgttgcaccg gtccgttcgc aagaagaaga atagtcctag 660
 gctggatgat tgggttctct gccggattta caacaaaaaa ggagctaccg agaggcgggg 720
 accaccgcct cccggtgttt acggcgcacga aatcatggag gagaagccga aggtgacgg 780
 gatggttatg cctccgcccgc cgcaacagac aagttagttc gcgtatttcg acacgtcgg 840
 ttccgtgccc aagctgcata ctacggattc gagttgctcg gaggcaggtagtgg tgtcgccgg 900
 gttcacgagc gaggttcaga gcgagcccaa gtggaaagat tggtcggccg taagtaatga 960
 caataacaat acccttgatt ttgggtttaa ttacattgtat gccaccgtgg ataacgcgtt 1020
 tggaggagga gggagtagta atcagatgtt tccgctacag gatatgttca tgtacatgca 1080
 gaagccttac tagaaggaa ttcccttcct gccggcggaaa cgcaacgc当地 aacgaccctc 1140
 gttttgcgt ttatggcaac acgagaccgt tttatatggt caatgagttgt gccgattcgg 1200
 ccattagatt tctgttcagt ttccgtttat tctatagacc gtccgatttc agatcatccc 1260
 taatccggacg gtggcgttg gatgtatcag tagtgttata ctgtgtttagg tagaagaaaa 1320
 tccacttgtt cttaaattgg cataaaagtc agaagctaattttatgtt gccgcaatca 1380
 atttaatatt ttctgtctaa aaaaaaaaaa 1409

<210> 38

<211> 319

<212> PRT

<213> Arabidopsis thaliana

<220>

<223> G502

<400> 38

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Asp	Glu	Glu	Leu	Val	Met	His	Tyr	Leu	Cys	Arg	Lys	Cys	Ala	Ser	Gln
									25					30	

Ser	Ile	Ala	Val	Pro	Ile	Ile	Ala	Glu	Ile	Asp	Leu	Tyr	Lys	Tyr	Asp
									40				45		

Pro	Trp	Glu	Leu	Pro	Gly	Leu	Ala	Leu	Tyr	Gly	Glu	Lys	Glu	Trp	Tyr
									55			60			

Phe	Phe	Ser	Pro	Arg	Asp	Arg	Lys	Tyr	Pro	Asn	Gly	Ser	Arg	Pro	Asn
65									75				80		

Arg	Ser	Ala	Gly	Ser	Gly	Tyr	Trp	Lys	Ala	Thr	Gly	Ala	Asp	Lys	Pro
									85				95		

Ile	Gly	Leu	Pro	Lys	Pro	Val	Gly	Ile	Lys	Lys	Ala	Leu	Val	Phe	Tyr
									100				110		

Ala	Gly	Lys	Ala	Pro	Lys	Gly	Glu	Lys	Thr	Asn	Trp	Ile	Met	His	Glu
									115			125			

Tyr	Arg	Leu	Ala	Asp	Val	Asp	Arg	Ser	Val	Arg	Lys	Lys	Lys	Asn	Ser
									130			140			

Leu	Arg	Leu	Asp	Asp	Trp	Val	Leu	Cys	Arg	Ile	Tyr	Asn	Lys	Lys	Gly
									145			155		160	

Ala Thr Glu Arg Arg Gly Pro Pro Pro Pro Val Val Tyr Gly Asp Glu
 165 170 175

Ile Met Glu Glu Lys Pro Lys Val Thr Glu Met Val Met Pro Pro Pro
 180 185 190

Pro Gln Gln Thr Ser Glu Phe Ala Tyr Phe Asp Thr Ser Asp Ser Val
 195 200 205

Pro Lys Leu His Thr Thr Asp Ser Ser Cys Ser Glu Gln Val Val Ser
 210 215 220

Pro Glu Phe Thr Ser Glu Val Gln Ser Glu Pro Lys Trp Lys Asp Trp
 225 230 235 240

Ser Ala Val Ser Asn Asp Asn Asn Asn Thr Leu Asp Phe Gly Phe Asn
 245 250 255

Tyr Ile Asp Ala Thr Val Asp Asn Ala Phe Gly Gly Gly Ser Ser
 260 265 270

Asn Gln Met Phe Pro Leu Gln Asp Met Phe Met Tyr Met Gln Lys Pro
 275 280 285

Tyr Lys Gly Ile Pro Phe Leu Pro Pro Lys Arg Asn Ala Lys Arg Pro
 290 295 300

Ser Phe Leu Arg Leu Trp Gln His Glu Thr Val Leu Tyr Gly Gln
 305 310 315

<210> 39
 <211> 1347
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G239

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 ggcgatgctc gttggAACCA catcgctcgt tcctctggc taaagcgaac tggtaagagt 180
 tgttagattaa gatggcttaa ttacttacgt ccagatgtta gaagaggcaa catcactctc 240
 gaagaacaat ttatgatcct caaactccat tctctttggg gcaatagggt gtcgaagatt 300
 gcgcaatatac taccgggaag aacagataat gaaataaaga attattggag aactcgagtc 360
 caaaagcaag ccaaacaccc aagatgcgtat gttAACAGTA atctttcaa ggagactatg 420
 agaaaatgttt ggatGCCGAG attagtggaa cgaatcaacg cccaaatcatt acccaccacg 480
 tgtgaacaag tggagtcaat gatcaccgac ccaagtcaac cagttAACGA accgagtccg 540
 gtcgagccgg gtttcgttca attcagccag aatcatcatc agcaattcgt accggctacg 600
 gaattgtcag caacgtcttc gaattctccg gctgagacgt tttcggacgt tcggaggtggg 660
 gtgggtgaacg ggtcagggtta tgatccgtcg ggtcaaaacgg gtttcggaga gttcaacgt 720
 tggggctgtg ttgggtggga caacatgtgg actgacgagg agagttttg gttcttgcag 780
 gaccagttct gccccgatac gacatcgat tcgtataatt aaggaaatat acgattacta 840
 tacgtAACGA ggaattcaat tgcgtcacgt ttgggtgtat attcattcgt gctgtatgcc 900
 aatttttagat acggccttgg tatacgaatc tttgacttaa ttattatctt ttctttccct 960
 ctcttgcattt aaacccttga ttaaattaag atttgatcat cagacgagga tatttgcgtat 1020

tcactgattt gtgatattga tataatgtgaa ttatttgata taacgtttta aaaaccaaca 1080
 aaaaaaaaaaa atcattccaa ggaaaagttc ttaattttga tactcgaaaa gagcgttagac 1140
 tgactcgaat cagttcatat tttctttgggt tcgttttatt tacgacaaaa ttcaactaaca 1200
 aaaattaaaa aacgacaaaa cgaaaaatatg actaaattta ttttttgcg agttaaccac 1260
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 gaatgctgca tgggtgatcc taagaaaa 1347

<210> 40
 <211> 273
 <212> PRT
 <213> *Arabidopsis thaliana*

<220>
 <223> G239

<400> 40
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 Asp Ser Asp Val Arg Lys Gly Pro Trp Thr Glu Glu Asp Ala Ile
 20 25 30
 Leu Val Asn Phe Val Ser Ile His Gly Asp Ala Arg Trp Asn His Ile
 35 40 45
 Ala Arg Ser Ser Gly Leu Lys Arg Thr Gly Lys Ser Cys Arg Leu Arg
 50 55 60
 Trp Leu Asn Tyr Leu Arg Pro Asp Val Arg Arg Gly Asn Ile Thr Leu
 65 70 75 80
 Glu Glu Gln Phe Met Ile Leu Lys Leu His Ser Leu Trp Gly Asn Arg
 85 90 95
 Trp Ser Lys Ile Ala Gln Tyr Leu Pro Gly Arg Thr Asp Asn Glu Ile
 100 105 110
 Lys Asn Tyr Trp Arg Thr Arg Val Gln Lys Gln Ala Lys His Leu Arg
 115 120 125
 Cys Asp Val Asn Ser Asn Leu Phe Lys Glu Thr Met Arg Asn Val Trp
 130 135 140
 Met Pro Arg Leu Val Glu Arg Ile Asn Ala Gln Ser Leu Pro Thr Thr
 145 150 155 160
 Cys Glu Gln Val Glu Ser Met Ile Thr Asp Pro Ser Gln Pro Val Asn
 165 170 175
 Glu Pro Ser Pro Val Glu Pro Gly Phe Val Gln Phe Ser Gln Asn His
 180 185 190
 His Gln Gln Phe Val Pro Ala Thr Glu Leu Ser Ala Thr Ser Ser Asn
 195 200 205
 Ser Pro Ala Glu Thr Phe Ser Asp Val Arg Gly Gly Val Val Asn Gly
 210 215 220

Ser Gly Tyr Asp Pro Ser Gly Gln Thr Gly Phe Gly Glu Phe Asn Asp
 225 230 235 240

Trp Gly Cys Val Gly Gly Asp Asn Met Trp Thr Asp Glu Glu Ser Phe
 245 250 255

Trp Phe Leu Gln Asp Gln Phe Cys Pro Asp Thr Thr Ser Tyr Ser Tyr
 260 265 270

Asn

<210> 41
 <211> 1360
 <212> DNA
 <213> *Arabidopsis thaliana*

<220>
 <223> G555

<400> 41
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 ccacgtctt gcagttgttag attggaaattt tccgatctt tctctaaatc gcttttctc 120
 cgagcaactt ttgtttgggg ttaagctcaa agaatccgtt ctttcagtc tttactccat 180
 ctagggtacc acgattggat cggttttat ctgtatgattt agtaacagag attttgaaga 240
 aaaagaaaaa tgggagatac tagtccaaga acatcagtct caacagatgg agacactgat 300
 cataataacc taatgttcga tgaagggcat ttgggtatcg gtgcttctga ttcttagtgac 360
 cgttcaaaga gtaaaatgga tcaaaagacg cttcgttaggc tcgctcaaaa ccgtgaggct 420
 gcaaggaaaaa gcagattgag gaagaaaagca tatgttcagc agcttagagaa cagtcgattg 480
 aagctaacac aacttgagca ggagctacaa agagcacggc aacaggggtt ctttatctca 540
 agctctggag accaagccca ttctaccgtt ggagatgggg caatggcatt tgatgttagaa 600
 tacagacgtt ggcaggaaga taaaaacaga cagatgaagg agctgagttc tgctatagat 660
 tctcacgcga ctgattctga gttcggata attgttagatg gagtaatagc tcactatgag 720
 gagcttaca ggataaaaagg caacgcagct aagagtgtt tcttccattt attatcaggg 780
 atgtggaaaa ccccagctga gagatgtttc ttgtggctcg gcgggttccg ttcatcagaa 840
 cttctcaagc ttatagcgtg tcagttggag cccttgacag aacaacaatc gctagacata 900
 aataacttgc aacagtcaac tcagcaagca gaagatgtt tgcgtcaagg gatggacaac 960
 ttacagcaat cactcgctga tactttatcg agtggactc tcgggtcaag ttcatcaggg 1020
 aatgttagcta gctacatggg tcagatggcc atggcgatgg ggaagttagg tacccttgaa 1080
 ggatttatcc gccaggctga taacttaagg ctacaaacat atcaacagat ggtgagacta 1140
 ttaacaaccc gacaatcgcc tcgtgctctc cttgcgtac acaattatac attgcggta 1200
 cgtgctctta gctcttatg gcttgcaga ccaagagagt gaaccatgac tctattatac 1260
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 tagctcgtat acaagattt gattatactg ttttgtgttg 1360

<210> 42
 <211> 330
 <212> PRT
 <213> *Arabidopsis thaliana*

<220>
 <223> G555

<400> 42
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 20 25 30
 Ser Asp Ser Ser Asp Arg Ser Lys Ser Lys Met Asp Gln Lys Thr Leu
 35 40 45
 Arg Arg Leu Ala Gln Asn Arg Glu Ala Ala Arg Lys Ser Arg Leu Arg
 50 55 60
 Lys Lys Ala Tyr Val Gln Gln Leu Glu Asn Ser Arg Leu Lys Leu Thr
 65 70 75 80
 Gln Leu Glu Gln Glu Leu Gln Arg Ala Arg Gln Gln Gly Val Phe Ile
 85 90 95
 Ser Ser Ser Gly Asp Gln Ala His Ser Thr Ala Gly Asp Gly Ala Met
 100 105 110
 Ala Phe Asp Val Glu Tyr Arg Arg Trp Gln Glu Asp Lys Asn Arg Gln
 115 120 125
 Met Lys Glu Leu Ser Ser Ala Ile Asp Ser His Ala Thr Asp Ser Glu
 130 135 140
 Leu Arg Ile Ile Val Asp Gly Val Ile Ala His Tyr Glu Glu Leu Tyr
 145 150 155 160
 Arg Ile Lys Gly Asn Ala Ala Lys Ser Asp Val Phe His Leu Leu Ser
 165 170 175
 Gly Met Trp Lys Thr Pro Ala Glu Arg Cys Phe Leu Trp Leu Gly Gly
 180 185 190
 Phe Arg Ser Ser Glu Leu Leu Lys Leu Ile Ala Cys Gln Leu Glu Pro
 195 200 205
 Leu Thr Glu Gln Gln Ser Leu Asp Ile Asn Asn Leu Gln Gln Ser Thr
 210 215 220
 Gln Gln Ala Glu Asp Ala Leu Ser Gln Gly Met Asp Asn Leu Gln Gln
 225 230 235 240
 Ser Leu Ala Asp Thr Leu Ser Ser Gly Thr Leu Gly Ser Ser Ser Ser
 245 250 255
 Gly Asn Val Ala Ser Tyr Met Gly Gln Met Ala Met Ala Met Gly Lys
 260 265 270
 Leu Gly Thr Leu Glu Gly Phe Ile Arg Gln Ala Asp Asn Leu Arg Leu
 275 280 285
 Gln Thr Tyr Gln Gln Met Val Arg Leu Leu Thr Thr Arg Gln Ser Ala
 290 295 300

Arg Ala Leu Leu Ala Val His Asn Tyr Thr Leu Arg Leu Arg Ala Leu
 305 310 315 320

Ser Ser Leu Trp Leu Ala Arg Pro Arg Glu
 325 330

<210> 43
 <211> 817
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G352

<400> 43
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 cgctcgccct cttctccggc atcgtgaaga aatggagcct gagaatctcg agcaatggc 180
 taaaagaaaa cgaacaaaac gtcaacgtt tgatcacggt catcagaatc aagaaacgaa 240
 caagaacctt cttctgaag aagagtatct cgctcttgc tccctcatgc tcgctcgtgg 300
 ctccgcgtt caatctcctc ctcttcctcc tctaccgtca cgtgcgtcact cgtccgatca 360
 ccgagattac aagtgtacgg tctgtggaa gtcctttcg tcataccaag ccttaggtgg 420
 acacaagacg agtcacccgaa aaccgacgaa cactagtatc acttccggta accaagaact 480
 gtctaataac agtcacagta acagcgggtc cgttgttatt aacgttaccg tgaacactgg 540
 taacgggtt agtcaaagcg gaaagattca cacttgctca atctgtttca agtctgttgc 600
 gtctggtaa gccttaggtg gacacaaaacg gtgtcaactat gacggtgcc acaacggtaa 660
 cggtaacggaa agtagcagca acagcgtaga actcgtcgct ggttagtgc acgtcgatgt 720
 tgataatgag agatggtccg aagaaagtgc gatcggtgcc caccgtggat ttgacctaaa 780
 cttaccggct gatcaagtct cagtgacgac ttcttaa 817

<210> 44
 <211> 245
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G352

<400> 44
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Pro Leu Leu Arg Tyr Arg Glu Glu Met Glu Pro Glu Asn Leu Glu Gln
 20 25 30

Trp Ala Lys Arg Lys Arg Thr Lys Arg Gln Arg Phe Asp His Gly His
 35 40 45

Gln Asn Gln Glu Thr Asn Lys Asn Leu Pro Ser Glu Glu Glu Tyr Leu
 50 55 60

Ala Leu Cys Leu Leu Met Leu Ala Arg Gly Ser Ala Val Gln Ser Pro
 65 70 75 80

Pro Leu Pro Pro Leu Pro Ser Arg Ala Ser Pro Ser Asp His Arg Asp
 85 90 95

Tyr Lys Cys Thr Val Cys Gly Lys Ser Phe Ser Ser Tyr Gln Ala Leu
 100 105 110

Gly Gly His Lys Thr Ser His Arg Lys Pro Thr Asn Thr Ser Ile Thr
 115 120 125

Ser Gly Asn Gln Glu Leu Ser Asn Asn Ser His Ser Asn Ser Gly Ser
 130 135 140

Val Val Ile Asn Val Thr Val Asn Thr Gly Asn Gly Val Ser Gln Ser
 145 150 155 160

Gly Lys Ile His Thr Cys Ser Ile Cys Phe Lys Ser Phe Ala Ser Gly
 165 170 175

Gln Ala Leu Gly Gly His Lys Arg Cys His Tyr Asp Gly Gly Asn Asn
 180 185 190

Gly Asn Gly Asn Gly Ser Ser Ser Asn Ser Val Glu Leu Val Ala Gly
 195 200 205

Ser Asp Val Ser Asp Val Asp Asn Glu Arg Trp Ser Glu Glu Ser Ala
 210 215 220

Ile Gly Gly His Arg Gly Phe Asp Leu Asn Leu Pro Ala Asp Gln Val
 225 230 235 240

Ser Val Thr Thr Ser
 245

<210> 45
 <211> 1001
 <212> DNA
 <213> *Arabidopsis thaliana*

<220>
 <223> G1352

<400> 45
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 aagaagattc atctgatcat ggcctcgaa gcgtgaaca ctccaaacttc ttctttcacc 120
 agaatcgaaa cgaaagaaga tttgatgaac gacggcggtt tcattgagcc gtggcttaaa 180
 cgcaaacgct ccaaaccgtca gcgttctcac agcccttctt cgtttcttc ctcaccgcct 240
 cgatctcgac ccaaatacccga gaatcaagat cttacggaaag aagagtatct cgctctttgt 300
 ctcctcatgc tcgctaaaga tcaaccgtcg caaacgcgtat ttcatcaaca gtcgcaatcg 360
 ttaacccgcg cggcagaatc aaagaacctt ccgtacaagt gtaacgtctg tgaaaaagcgc 420
 tttccttctt atcaggctt aggcggtcac aaagcaagtc accgaatcaa accaccaacc 480
 gtaatctcaa caaccgcga tgattcaaca gtcggacca tctccatcg tgcggagaa 540
 aaacatccga ttgctgcctc cggaaagatc cacgagtgtt caatctgtca taaagtgttt 600
 cccgacgggtc aagctttagg cggtcacaaa cgttgtcact acgaaggcaa cctcggcggc 660
 ggaggaggag gagaaagcaa atcaatcagt cacagtggaa ggcgttcgag cacggtatcg 720
 gaagaaaagga gccaccgtgg attcatcgat ctaaacctac cggcggttacc tgaactcagc 780
 cttcatcaca atccaatcgt cgacgaagag atcttgatgc cgttgaccgg taaaaaaccg 840
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 ttttagttac aaattttaa ttgttctgtat ttggattgaa a 1001

<210> 46
 <211> 273
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G1352

<400> 46
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 35 40 45
 Ser Ser Ser Pro Pro Arg Ser Arg Pro Lys Ser Gln Asn Gln Asp
 50 55 60
 Leu Thr Glu Glu Glu Tyr Leu Ala Leu Cys Leu Leu Met Leu Ala Lys
 65 70 75 80
 Asp Gln Pro Ser Gln Thr Arg Phe His Gln Gln Ser Gln Ser Leu Thr
 85 90 95
 Pro Pro Pro Glu Ser Lys Asn Leu Pro Tyr Lys Cys Asn Val Cys Glu
 100 105 110
 Lys Ala Phe Pro Ser Tyr Gln Ala Leu Gly Gly His Lys Ala Ser His
 115 120 125
 Arg Ile Lys Pro Pro Thr Val Ile Ser Thr Thr Ala Asp Asp Ser Thr
 130 135 140
 Ala Pro Thr Ile Ser Ile Val Ala Gly Glu Lys His Pro Ile Ala Ala
 145 150 155 160
 Ser Gly Lys Ile His Glu Cys Ser Ile Cys His Lys Val Phe Pro Thr
 165 170 175
 Gly Gln Ala Leu Gly Gly His Lys Arg Cys His Tyr Glu Gly Asn Leu
 180 185 190
 Gly Gly Gly Gly Gly Ser Lys Ser Ile Ser His Ser Gly Ser
 195 200 205
 Val Ser Ser Thr Val Ser Glu Glu Arg Ser His Arg Gly Phe Ile Asp
 210 215 220
 Leu Asn Leu Pro Ala Leu Pro Glu Leu Ser Leu His His Asn Pro Ile
 225 230 235 240
 Val Asp Glu Glu Ile Leu Ser Pro Leu Thr Gly Lys Lys Pro Leu Leu
 245 250 255

Leu Thr Asp His Asp Gln Val Ile Lys Lys Glu Asp Leu Ser Leu Lys
 260 265 270

Ile

<210> 47
 <211> 2663
 <212> DNA
 <213> *Arabidopsis thaliana*

<220>
 <223> G1089

<400> 47
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 cgttaacgctt tcgccccgc tcactcagct tacgctatgg ctcttaaaaa caccggagct 180
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 gctgcagcaa tcgcttctac ttcttctt cccactgcta tatctccctt tcttccttct 300
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 35 40 45
 Gly Ala Ala Leu Ser Asp Tyr Ser His Gly Glu Phe Leu Val Ser Asn
 50 55 60
 His Ser Ser Ser Ser Ala Ala Ala Ile Ala Ser Thr Ser Ser Leu
 65 70 75 80
 Pro Thr Ala Ile Ser Pro Pro Leu Pro Ser Ser Thr Ala Pro Val Ser
 85 90 95
 Asn Ser Thr Ala Ser Ser Ser Ala Ala Val Pro Gln Pro Ile Pro
 100 105 110
 Asp Thr Leu Pro Pro Pro Pro Pro Pro Leu Pro Leu Gln Arg
 115 120 125
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 Gly Ser Gly Leu Asn Gly Ile Glu Glu Asp Gly Ala Leu Asp Asn Asp
 145 150 155 160
 Asp Asp Asp Asp Asp Asp Asp Asp Ser Glu Met Glu Asn Arg Asp
 165 170 175
 Arg Leu Ile Arg Lys Ser Arg Ser Arg Gly Gly Ser Thr Arg Gly Asn
 180 185 190
 Arg Thr Thr Ile Glu Asp His His Leu Gln Glu Glu Lys Ala Pro Pro
 195 200 205
 Pro Pro Pro Leu Ala Asn Ser Arg Pro Ile Pro Pro Pro Arg Gln His
 210 215 220

Gln His Gln His Gln Gln Gln Gln Pro Phe Tyr Asp Tyr Phe
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 Phe Pro Asn Val Glu Asn Met Pro Gly Thr Thr Leu Glu Asp Thr Pro
 245 250 255
 Pro Gln Pro Gln Pro Gln Pro Thr Arg Pro Val Pro Pro Gln Pro His
 260 265 270
 Ser Pro Val Val Thr Glu Asp Asp Glu Asp Glu Glu Glu Glu Glu
 275 280 285
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 Glu Glu Arg Pro Lys Arg Val Glu Glu Val Thr Ile Glu Leu Glu Lys
 305 310 315 320
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 325 330 335
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 405 410 415
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 420 425 430
 Thr Val Leu Asp Lys Leu Leu Ala Trp Glu Lys Lys Leu Tyr Asp Glu
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 Val Lys Ala Gly Glu Leu Met Lys Ile Glu Tyr Gln Lys Lys Val Ala
 450 455 460
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 465 470 475 480
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 485 490 495
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 515 520 525

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 545 550 555 560
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 565 570 575
 Trp His Thr Gln Phe Cys Arg Met Ile Asp His Gln Lys Glu Tyr Ile
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 Lys Ala Leu Gly Gly Trp Leu Lys Leu Asn Leu Ile Pro Ile Glu Ser
 595 600 605
 Thr Leu Lys Glu Lys Val Ser Ser Pro Pro Arg Val Pro Asn Pro Ala
 610 615 620
 Ile Gln Lys Leu Leu His Ala Trp Tyr Asp Arg Leu Asp Lys Ile Pro
 625 630 635 640
 Asp Glu Met Ala Lys Ser Ala Ile Ile Asn Phe Ala Ala Val Val Ser
 645 650 655
 Thr Ile Met Gln Gln Gln Glu Asp Glu Ile Ser Leu Arg Asn Lys Cys
 660 665 670
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 675 680 685
 Trp Tyr His Lys Tyr Ile Gln Lys Arg Gly Pro Glu Gly Met Asn Pro
 690 695 700
 Asp Glu Ala Asp Asn Asp His Asn Asp Glu Val Ala Val Arg Gln Phe
 705 710 715 720
 Asn Val Glu Gln Ile Lys Lys Arg Leu Glu Glu Glu Glu Ala Tyr
 725 730 735
 His Arg Gln Ser His Gln Val Arg Glu Lys Ser Leu Ala Ser Leu Arg
 740 745 750
 Thr Arg Leu Pro Glu Leu Phe Gln Ala Met Ser Glu Val Ala Tyr Ser
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tctgtgtata ataactctt tgaaggcagaa cgcgcgactataatgatca ggacgaagac 360
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Asn Gln Ser Ser Thr Thr Leu Glu Val Asp Ala Arg Pro Glu Ala
50 55 60
Asp Asp Asn Asn Arg Val Asn Tyr Thr Ser Val Tyr Asn Asn Ser Leu
65 70 75 80
Glu Ala Glu Pro Ser Ser Asn Asn Asp Gln Asp Glu Asp Arg Ile Asn
85 90 95

Asp Lys Met Lys Arg Arg Leu Ala Gln Asn Arg Glu Ala Ala Arg Lys
 100 105 110
 Ser Arg Leu Arg Lys Lys Ala His Val Gln Gln Leu Glu Glu Ser Arg
 115 120 125
 Leu Lys Leu Ser Gln Leu Glu Gln Glu Leu Val Arg Ala Arg Gln Gln
 130 135 140
 Gly Leu Cys Val Arg Asn Ser Ser Asp Thr Ser Tyr Leu Gly Pro Ala
 145 150 155 160
 Gly Asn Met Asn Ser Gly Ile Ala Ala Phe Glu Met Glu Tyr Thr His
 165 170 175
 Trp Leu Glu Glu Gln Asn Arg Arg Val Ser Glu Ile Arg Thr Ala Leu
 180 185 190
 Gln Ala His Ile Gly Asp Ile Glu Leu Lys Met Leu Val Asp Ser Cys
 195 200 205
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 Ala Asp Val Phe Phe Leu Met Ser Gly Met Trp Arg Thr Ser Thr Glu
 225 230 235 240
 Arg Phe Phe Gln Trp Ile Gly Gly Phe Arg Pro Ser Glu Leu Leu Asn
 245 250 255
 Val Val Met Pro Tyr Val Glu Pro Leu Thr Asp Gln Gln Leu Leu Glu
 260 265 270
 Val Arg Asn Leu Gln Gln Ser Ser Gln Gln Ala Glu Glu Ala Leu Ser
 275 280 285
 Gln Gly Leu Asp Lys Leu Gln Gln Gly Leu Val Glu Ser Ile Ala Ile
 290 295 300
 Gln Ile Lys Val Val Glu Ser Val Asn His Gly Ala Pro Met Ala Ser
 305 310 315 320
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 325 330 335
 His Leu Arg Gln Gln Thr Leu Gln Gln Met Ser Lys Ile Leu Thr Thr
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 Ala Ser Leu Ile Ile Gly Ile Ile Tyr Ser Val Ile Val Met Lys Leu
 50 55 60
 Asn Leu Thr Thr Gly Leu Val Pro Asn Leu Asn Val Ser Ala Ala Leu
 65 70 75 80
 Leu Ala Phe Val Phe Leu Arg Ser Trp Thr Lys Leu Leu Thr Lys Ala
 85 90 95
 Gly Ile Val Thr Lys Pro Phe Thr Lys Gln Glu Asn Thr Val Val Gln
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 115 120 125
 Ser Tyr Leu Leu Gly Leu Asn Arg Ile Thr Tyr Glu Gln Ser Gly Gly
 130 135 140
 Thr His Thr Asp Gly Asn Tyr Pro Glu Gly Thr Lys Glu Pro Gly Ile
 145 150 155 160
 Gly Trp Met Thr Ala Phe Leu Phe Phe Thr Cys Phe Val Gly Leu Leu
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 Ala Leu Val Pro Leu Arg Lys Ile Met Ile Ile Asp Tyr Lys Leu Thr
 180 185 190
 Tyr Pro Ser Gly Thr Ala Thr Ala Val Leu Ile Asn Gly Phe His Thr
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 Pro Lys Gly Asn Lys Met Ala Lys Lys Gln Val Phe Gly Phe Val Lys
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 Tyr Phe Ser Phe Ser Phe Ile Trp Ala Phe Phe Gln Trp Phe Phe Ser
 225 230 235 240
 Gly Gly Thr Glu Cys Gly Phe Ile Gln Phe Pro Thr Phe Gly Leu Glu
 245 250 255
 Ala Leu Lys Asn Thr Phe Tyr Phe Asp Phe Ser Met Thr Tyr Val Gly
 260 265 270
 Ala Gly Met Ile Cys Pro His Ile Val Asn Ile Ser Leu Leu Phe Gly
 275 280 285
 Ala Val Leu Ser Trp Gly Ile Met Trp Pro Leu Ile Lys Gly Leu Lys
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Gly Asp Trp Phe Pro Ser Thr Leu Pro Glu Asn Ser Met Lys Ser Leu
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Asn Gly Tyr Lys Val Phe Ile Ser Ile Ser Leu Ile Leu Gly Asp Gly
 325 330 335

Leu Tyr Gln Phe Ile Lys Ile Leu Phe Lys Thr Gly Ile Asn Met Tyr
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Val Lys Leu Asn Asn Arg Asn Ser Gly Lys Ser Asn Ser Glu Lys Asp
 355 360 365

Lys Gln Ser Ile Ala Asp Leu Lys Arg Asp Glu Ile Phe Val Arg Asp
 370 375 380

Ser Ile Pro Leu Trp Val Ala Ala Val Gly Asn Ala Ala Phe Ser Val
 385 390 395 400

Val Ser Ile Ile Ala Ile Pro Ile Met Phe Pro Glu Leu Lys Trp Tyr
 405 410 415

Phe Ile Val Val Ala Tyr Met Leu Ala Pro Ser Leu Gly Phe Ser Asn
 420 425 430

Ala Tyr Gly Ala Gly Leu Thr Asp Met Asn Met Ala Tyr Asn Tyr Gly
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Lys Val Ala Leu Phe Ile Leu Ala Ala Met Ala Gly Lys Gln Asn Gly
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Val Val Ala Gly Leu Val Gly Cys Gly Leu Ile Lys Ser Ile Val Ser
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Ile Ser Ser Asp Leu Met His Asp Phe Lys Thr Gly His Leu Thr Leu
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Thr Ser Pro Arg Ser Met Leu Val Ser Gln Ala Ile Gly Thr Ala Ile
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 515 520 525

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Gln His Cys Leu Gln Leu Cys Tyr Gly Phe Phe Ala Phe Ala Val Ala
 565 570 575

Ala Asn Leu Val Arg Asp Arg Leu Pro Asp Lys Ile Gly Asn Trp Val
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Pro Leu Pro Met Ala Met Ala Val Pro Phe Leu Val Gly Gly Tyr Phe
 595 600 605

Ala Ile Asp Met Cys Val Gly Ser Leu Ile Val Phe Ala Trp Asn Met
 610 615 620

Arg Asp Arg Val Lys Ala Gly Leu Met Val Pro Ala Val Ala Ser Gly
 625 630 635 640

Leu Ile Cys Gly Asp Gly Leu Trp Ile Leu Pro Ser Ser Val Leu Ala
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 35 40 45

Leu Gly Ser Leu His Tyr His Arg Gln Leu Asn Ile Gly His Glu Pro
 50 55 60

Met Leu Lys Asn Gln Asn Pro Asn Asn Ser Ile Phe Gln Asp Phe Leu
 65 70 75 80

Asn Met Pro Leu Asn Gln Pro Pro Pro Pro Pro Pro Pro Pro Ser Ser
 85 90 95

Ser Thr Ile Val Thr Ala Leu Tyr Gly Ser Leu Pro Leu Pro Pro Pro
 100 105 110

Ala Thr Val Leu Ser Leu Asn Ser Gly Val Gly Phe Glu Phe Leu Asp
 115 120 125

Thr Thr Glu Asn Leu Leu Ala Ser Asn Pro Arg Ser Phe Glu Glu Ser
 130 135 140

Ala Lys Phe Gly Cys Leu Gly Lys Lys Arg Gly Gln Asp Ser Asp Asp
 145 150 155 160

Thr Arg Gly Asp Arg Arg Tyr Lys Arg Met Ile Lys Asn Arg Glu Ser
 165 170 175

Ala Ala Arg Ser Arg Ala Arg Lys Gln Ala Tyr Thr Asn Glu Leu Glu
 180 185 190

Leu Glu Ile Ala His Leu Gln Thr Glu Asn Ala Arg Leu Lys Ile Gln
 195 200 205

Gln Glu Gln Leu Lys Ile Ala Glu Ala Thr Gln Asn Gln Val Lys Lys
 210 215 220

Thr Leu Gln Arg Ser Ser Thr Ala Pro Phe
 225 230

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 ctccccgatg ccggcgatga tttcattgtc ggtgactgtc tcgtctacga ggacggcgatc 240
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 aatccggcgtg gcggggctaa gagattagat gaatccgaga ttgagccgaa gaacctcgtg 360

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 ctgaggaag ttgacttgaa tgactttctc acgtacaagg aagccaagtt ggctcaattg 540
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 35 40 45

Asp Ala Gly Asp Asp Phe Ile Val Gly Asp Cys Leu Val Tyr Glu Asp
 50 55 60

Gly Val Phe Glu Asp Pro Tyr Leu Asp Lys Glu Val Thr Gln Val Ala
 65 70 75 80

Lys Gln Glu Arg Lys Lys Asn Arg Arg Gly Gly Ala Lys Arg Leu Asp
 85 90 95

Glu Ser Glu Ile Glu Pro Glu Asn Leu Val Pro Glu Glu Trp Arg Asp
 100 105 110

Ile Gln Ala Glu Val Asn Leu Thr Lys Lys Asp Lys Arg Lys Ile Ala
 115 120 125

Gln Glu Met Glu Phe Gly Val Arg Val Glu Lys Lys Arg Gln Gly Leu
 130 135 140

Ile Pro Leu Arg Lys Val Asp Leu Asn Asp Phe Leu Thr Tyr Lys Glu
 145 150 155 160
 Ala Lys Leu Ala Gln Leu Arg Pro Val Ile Leu Asp Lys Pro Gly Asn
 165 170 175
 Phe Ser Asp Asp Ser Gly Ala Ser Ser Asp Gly Glu Thr Ala Val Ser
 180 185 190
 Ser Pro Ser Glu Arg Val Ala Pro Lys Asn Pro Arg Trp Ala Val Tyr
 195 200 205
 Gly Lys Gly Phe Asp His Val Ala Lys Phe Phe Asn Ser Asp Lys Tyr
 210 215 220
 Asp Pro Ser Asp Lys Lys Ser Asp Gly Pro Arg Lys Leu Leu Ser Lys
 225 230 235 240
 Glu Glu Lys Phe Met Leu Asn Ser Arg Asn Pro Asp Leu Ala Val Ala
 245 250 255
 Thr Ser Lys Lys Trp Leu Pro Leu His Thr Leu Ala Ala Cys Gly Glu
 260 265 270
 Phe Tyr Leu Val Asp Ser Leu Leu Lys His Asn Leu Asp Ile Asn Ala
 275 280 285
 Thr Asp Val Gly Gly Leu Thr Val Leu His Arg Ala Ile Ile Gly Lys
 290 295 300
 Lys Gln Ala Ile Thr Asn Tyr Leu Leu Arg Glu Ser Ala Asn Pro Phe
 305 310 315 320
 Val Leu Asp Asp Glu Gly Ala Thr Leu Met His Tyr Ala Val Gln Thr
 325 330 335
 Ala Ser Ala Pro Thr Ile Lys Leu Leu Leu Tyr Asn Ala Asp Ile
 340 345 350
 Asn Ala Gln Asp Arg Asp Gly Trp Thr Pro Leu His Val Ala Val Gln
 355 360 365
 Ala Arg Arg Ser Asp Ile Val Lys Leu Leu Leu Ile Lys Gly Ala Asp
 370 375 380
 Ile Glu Val Lys Asn Lys Asp Gly Leu Thr Pro Leu Gly Leu Cys Leu
 385 390 395 400
 Tyr Leu Gly Arg Glu Ile Arg Thr Tyr Glu Val Met Lys Leu Leu Lys
 405 410 415
 Glu Phe Pro Leu Ser Arg His Lys Lys Arg Leu Val Thr Thr Asp Glu
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 Asp Ile Glu
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ccccaaaaag	ctggactaaa	acgatgtgga	aagagttgt	gattgcgtat	ggctaactat	180
ttgaaacctg	acatcaagag	aggagagttt	agctatgagg	aggaacacat	tatcatcatg	240
ctcacacgctt	ctcgccgcaa	caagtgtca	gtcatagcga	gacatttgcc	caaaagaaca	300
gataacgaga	ttaagaacta	ctggaacacg	catctcaaaa	agctcctgtat	cgataaggg	360
atcgatcccc	tgacccacaa	gccacttgcc	tatgactcaa	acccgatgt	gcaatcgcaa	420
tcgggttcca	tctctccaaa	gtctcttcct	ccttcaagct	ccaaaaatgt	acccggagata	480
accagcagt	acgagacacc	gaaatatgtat	gcttccttga	gctccaagaa	acgttgttt	540
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ggaactatac	taggcgcctc	catcgaagga	accttgcata	gctctacacc	gttgtcttca	660
tgtctaaatg	atgactttt	tgaaacaagt	caatttcaga	tggaaagaatt	tgatccattc	720
tatcagtcat	ctgaacacat	aattgtatcat	atgaaaagaag	atatcagcat	caacaattcc	780
gaatacaatt	tctcgcagtt	tctcgagcag	tttagtaaca	acgaagggga	agaagctgac	840
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gatgaagacg	agatgtatgca	aaacataact	ggttggtcaa	attatcttc	tgaccattcc	960
gatttcaatt	atgacacgag	ccaagattat	gacgacaaga	acttcataatg	atccgttcat	1020
tgcttaccgg	actagagttg	accggtaat	gtcatatgg	tctcttagat	atttgcataat	1080
ttatagtaaa	ggtccactat	agggtcaacta	tatattaata	ttcagtaatg	gattctctt	1140
gttagagaac	cttgtgtatgc	cgtggatcaa	ttagtattt	atttgcggga	gacacgagg	1200
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<211> 336

<212> PRT

<213> *Arabidopsis thaliana*

<220>

<223> G201

<400> 58

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Gly Gly Gly Gly Trp Arg Asp Ile Pro Gln Lys Ala Gly Leu Lys Arg
35 40 45

Cys Gly Lys Ser Cys Arg Leu Arg Trp Ala Asn Tyr Leu Lys Pro Asp
 50 55 60

Ile Lys Arg Gly Glu Phe Ser Tyr Glu Glu Glu Gln Ile Ile Ile Met
65 70 75 80

Leu His Ala Ser Arg Gly Asn Lys Trp Ser Val Ile Ala Arg His Leu
 85 90 95
 Pro Lys Arg Thr Asp Asn Glu Ile Lys Asn Tyr Trp Asn Thr His Leu
 100 105 110
 Lys Lys Leu Leu Ile Asp Lys Gly Ile Asp Pro Val Thr His Lys Pro
 115 120 125
 Leu Ala Tyr Asp Ser Asn Pro Asp Glu Gln Ser Gln Ser Gly Ser Ile
 130 135 140
 Ser Pro Lys Ser Leu Pro Pro Ser Ser Ser Lys Asn Val Pro Glu Ile
 145 150 155 160
 Thr Ser Ser Asp Glu Thr Pro Lys Tyr Asp Ala Ser Leu Ser Ser Lys
 165 170 175
 Lys Arg Cys Phe Lys Arg Ser Ser Ser Thr Ser Lys Leu Leu Asn Lys
 180 185 190
 Val Ala Ala Arg Ala Ser Ser Met Gly Thr Ile Leu Gly Ala Ser Ile
 195 200 205
 Glu Gly Thr Leu Ile Ser Ser Thr Pro Leu Ser Ser Cys Leu Asn Asp
 210 215 220
 Asp Phe Ser Glu Thr Ser Gln Phe Gln Met Glu Glu Phe Asp Pro Phe
 225 230 235 240
 Tyr Gln Ser Ser Glu His Ile Ile Asp His Met Lys Glu Asp Ile Ser
 245 250 255
 Ile Asn Asn Ser Glu Tyr Asn Phe Ser Gln Phe Leu Glu Gln Phe Ser
 260 265 270
 Asn Asn Glu Gly Glu Ala Asp Asn Thr Gly Gly Tyr Asn Gln
 275 280 285
 Asp Leu Leu Met Ser Asp Val Ser Ser Thr Ser Val Asp Glu Asp Glu
 290 295 300
 Met Met Gln Asn Ile Thr Gly Trp Ser Asn Tyr Leu Leu Asp His Ser
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 325 330 335

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 <223> G1417
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 tggcgatgac aaaaccaaaa ctcaaattag tagactgaag ttggagctag agaggcttca 360
 cgaggagaat cacaactga agcatttatt agatgaggtc agtgagagtt acaacgacct 420
 ccaaagaaga gttttgttag caagacaac acaagtggaa ggtcttcata ataaacaaca 480
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 <213> *Arabidopsis thaliana*

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 <223> G1417

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 35 40 45
 Phe Phe Ala Ala Lys Ser Gln Pro Phe Asp Leu Gly His Val Arg Thr
 50 55 60
 Thr Thr Ile Val Gly Ser Ser Gly Phe Asn Asp Gly Leu Gly Leu Val
 65 70 75 80
 Asn Ser Cys His Gly Thr Ser Ser Asn Asp Gly Asp Asp Lys Thr Lys
 85 90 95

Thr Gln Ile Ser Arg Leu Lys Leu Glu Leu Glu Arg Leu His Glu Glu
 100 105 110

Asn His Lys Leu Lys His Leu Leu Asp Glu Val Ser Glu Ser Tyr Asn
 115 120 125

Asp Leu Gln Arg Arg Val Leu Leu Ala Arg Gln Thr Gln Val Glu Gly
 130 135 140

Leu His His Lys Gln His Glu Asp Val Pro Gln Ala Gly Ser Ser Gln
 145 150 155 160

Ala Leu Glu Asn Arg Arg Pro Lys Asp Met Asn His Glu Thr Pro Ala
 165 170 175

Thr Thr Leu Lys Arg Arg Ser Pro Asp Asp Val Asp Gly Arg Asp Met
 180 185 190

His Arg Gly Ser Pro Lys Thr Pro Arg Ile Asp Gln Asn Lys Ser Thr
 195 200 205

Asn His Glu Glu Gln Gln Asn Pro His Asp Gln Leu Pro Tyr Arg Lys
 210 215 220

Ala Arg Val Ser Val Arg Ala Arg Ser Asp Ala Thr Thr Val Asn Asp
 225 230 235 240

Gly Cys Gln Trp Arg Lys Tyr Gly Gln Lys Met Ala Lys Gly Asn Pro
 245 250 255

Cys Pro Arg Ala Tyr Tyr Arg Cys Thr Met Ala Val Gly Cys Pro Val
 260 265 270

Arg Lys Gln Val Gln Arg Cys Ala Glu Asp Thr Thr Ile Leu Thr Thr
 275 280 285

Thr Tyr Glu Gly Asn His Asn His Pro Leu Pro Pro Ser Ala Thr Ala
 290 295 300

Met Ala Ala Thr Thr Ser Ala Ala Ala Ala Met Leu Leu Ser Gly Ser
 305 310 315 320

Ser Ser Ser Asn Leu His Gln Thr Leu Ser Ser Pro Ser Ala Thr Ser
 325 330 335

Ser Ser Ser Phe Tyr His Asn Phe Pro Tyr Thr Ser Thr Ile Ala Thr
 340 345 350

Leu Ser Ala Ser Ala Pro Phe Pro Thr Ile Thr Leu Asp Leu Thr Asn
 355 360 365

Pro Pro Arg Pro Leu Gln Pro Pro Pro Gln Phe Leu Ser Gln Tyr Gly
 370 375 380

Pro Ala Ala Phe Leu Pro Asn Ala Asn Gln Ile Arg Ser Met Asn Asn
 385 390 395 400

Asn Asn Gln Gln Leu Leu Ile Pro Asn Leu Phe Gly Pro Gln Ala Pro
 405 410 415

Pro Arg Glu Met Val Asp Ser Val Arg Ala Ala Ile Ala Met Asp Pro
 420 425 430

Asn Phe Thr Ala Ala Leu Ala Ala Ile Ser Asn Ile Ile Gly Gly
 435 440 445

Gly Asn Asn Asp Asn Asn Asn Thr Asp Ile Asn Asp Asn Lys Val
 450 455 460

Asp Ala Lys Ser Gly Gly Ser Ser Asn Gly Asp Ser Pro Gln Leu Pro
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Gln Ser Cys Thr Thr Phe Ser Thr Asn
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<211> 1046

<212> DNA

<213> Arabidopsis thaliana

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<223> G233

<400> 61

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<211> 273

<212> PRT

<213> Arabidopsis thaliana

<220>

<223> G233

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 35 40 45
 Cys Gly Lys Ser Cys Arg Leu Arg Trp Met Asn Tyr Leu Lys Pro Asp
 50 55 60
 Ile Lys Arg Gly Asn Phe Thr Lys Glu Glu Asp Ala Ile Ile Ser
 65 70 75 80
 Leu His Gln Ile Leu Gly Asn Arg Trp Ser Ala Ile Ala Ala Lys Leu
 85 90 95
 Pro Gly Arg Thr Asp Asn Glu Ile Lys Asn Val Trp His Thr His Leu
 100 105 110
 Lys Lys Arg Leu Glu Asp Tyr Gln Pro Ala Lys Pro Lys Thr Ser Asn
 115 120 125
 Lys Lys Lys Gly Thr Lys Pro Lys Ser Glu Ser Val Ile Thr Ser Ser
 130 135 140
 Asn Ser Thr Arg Ser Glu Ser Glu Leu Ala Asp Ser Ser Asn Pro Ser
 145 150 155 160
 Gly Glu Ser Leu Phe Ser Thr Ser Pro Ser Thr Ser Glu Val Ser Ser
 165 170 175
 Met Thr Leu Ile Ser His Asp Gly Tyr Ser Asn Glu Ile Asn Met Asp
 180 185 190
 Asn Lys Pro Gly Asp Ile Ser Thr Ile Asp Gln Glu Cys Val Ser Phe
 195 200 205
 Glu Thr Phe Gly Ala Asp Ile Asp Glu Ser Phe Trp Lys Glu Thr Leu
 210 215 220
 Tyr Ser Gln Asp Glu His Asn Tyr Val Ser Asn Asp Leu Glu Val Ala
 225 230 235 240
 Gly Leu Val Glu Ile Gln Gln Glu Phe Gln Asn Leu Gly Ser Ala Asn
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<212> DNA
 <213> Arabidopsis thaliana

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 <223> G920

<400> 63

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 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G920

<400> 64

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 35 40 45

Val Ser Gly Ser Gly Ser Val Ser Gly Gly Pro Asp Pro Val Asp Glu
 50 55 60

Leu Met Ser Lys Ile Leu Gly Ser Phe His Lys Thr Ile Ser Val Leu
 65 70 75 80

Asp Ser Phe Asp Pro Val Ala Val Ser Val Pro Ile Ala Val Glu Gly
 85 90 95

Ser Trp Asn Ala Ser Cys Gly Asp Asp Ser Ala Thr Pro Val Ser Cys
 100 105 110
 Asn Gly Gly Asp Ser Gly Glu Ser Lys Lys Lys Arg Leu Gly Val Gly
 115 120 125
 Lys Gly Lys Arg Gly Cys Tyr Thr Arg Lys Thr Arg Ser His Thr Arg
 130 135 140
 Ile Val Glu Ala Lys Ser Ser Glu Asp Arg Tyr Ala Trp Arg Lys Tyr
 145 150 155 160
 Gly Gln Lys Glu Ile Leu Asn Thr Thr Phe Pro Arg Ser Tyr Phe Arg
 165 170 175
 Cys Thr His Lys Pro Thr Gln Gly Cys Lys Ala Thr Lys Gln Val Gln
 180 185 190
 Lys Gln Asp Gln Asp Ser Glu Met Phe Gln Ile Thr Tyr Ile Gly Tyr
 195 200 205
 His Thr Cys Thr Ala Asn Asp Gln Thr His Ala Lys Thr Glu Pro Phe
 210 215 220
 Asp Gln Glu Ile Ile Met Asp Ser Glu Lys Thr Leu Ala Ala Ser Thr
 225 230 235 240
 Ala Gln Asn His Val Asn Ala Met Val Gln Glu Gln Glu Asn Asn Thr
 245 250 255
 Ser Ser Val Thr Ala Ile Asp Ala Gly Met Val Lys Glu Glu Gln Asn
 260 265 270
 Asn Asn Gly Asp Gln Ser Lys Asp Tyr Tyr Glu Gly Ser Ser Thr Gly
 275 280 285
 Glu Asp Leu Ser Leu Val Trp Gln Glu Thr Met Met Phe Asp Asp His
 290 295 300
 Gln Asn His Tyr Tyr Cys Gly Glu Thr Ser Thr Thr Ser His Gln Phe
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 <212> DNA
 <213> *Arabidopsis thaliana*

<220>
 <223> G867

<400> 65

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 cccgcgataa ctccggcgaa aaagtcgtcg gtaggtaact tatacaggat gggaaagcgga 180
 tcaagcgtt tgtagattc agagaacgac gtagaagctg aatcttaggaa gcttccgtcg 240
 tcaaaaataca aagggtgtggt gccacaacca aacgaaagat ggggagctca gatttacgag 300
 aaacaccagc gcgtgtggct cgggacattc aacgaaagaag acgaagccgc tcgtgcctac 360
 gacgtcgcgg ttcacagggtt ccgtccgtg gacgccgtca caaatttcaa agacgtgaag 420
 atggacgaag acgaggtcga tttcttgaat tctcattcga aatctgagat cgttgatatg 480
 ttgaggaaac atacttataa cgaagagttt gaggcagatc aacggcgtcg taatggtaac 540
 ggaaacatga ctaggacgtt gtttaacgtcg gggttgagta atgatgggtt ttctacgacg 600
 gggtttagat cggcggaggc actgttttagaa aagcggtaa cggcaagcga cgttggaaag 660
 ctaaaaccgtt tggttatacc gaaacatcac gcagagaaac attttccgtt accgtcaagt 720
 aacgtttccg tgaaaggagt gttgttgaac tttgaggacg ttaacgggaa agtgtggagg 780
 ttccgttact cgtattggaa cagtagtcag agttatgttt tgactaaagg ttggagcagg 840
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 ggaaacaaaaa gagtgaacga tactgagatg ttatcggtt ggtgttagcaa gaagcaacgc 1080
 atcttcacg cctcgtaaca actcttcttc ttttttttc ttttgggtt ttaataattt 1140
 ttaaaaactc catttcgtt ttcttttattt gcatcggtt ctttcttctt gtttaccaaa 1200
 gtttcatgag ttgttttgg ttttgggtt gtttaccaaa 1260
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<210> 66
 <211> 344
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G867

<400> 66
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 Cys Glu Thr Pro Ala Ile Thr Pro Ala Lys Lys Ser Ser Val Gly Asn
 20 25 30
 Leu Tyr Arg Met Gly Ser Gly Ser Ser Val Val Leu Asp Ser Glu Asn
 35 40 45
 Gly Val Glu Ala Glu Ser Arg Lys Leu Pro Ser Ser Lys Tyr Lys Gly
 50 55 60
 Val Val Pro Gln Pro Asn Gly Arg Trp Gly Ala Gln Ile Tyr Glu Lys
 65 70 75 80
 His Gln Arg Val Trp Leu Gly Thr Phe Asn Glu Glu Asp Glu Ala Ala
 85 90 95
 Arg Ala Tyr Asp Val Ala Val His Arg Phe Arg Arg Arg Asp Ala Val
 100 105 110
 Thr Asn Phe Lys Asp Val Lys Met Asp Glu Asp Glu Val Asp Phe Leu
 115 120 125

Asn Ser His Ser Lys Ser Glu Ile Val Asp Met Leu Arg Lys His Thr
 130 135 140

Tyr Asn Glu Glu Leu Glu Gln Ser Lys Arg Arg Arg Asn Gly Asn Gly
 145 150 155 160

Asn Met Thr Arg Thr Leu Leu Thr Ser Gly Leu Ser Asn Asp Gly Val
 165 170 175

Ser Thr Thr Gly Phe Arg Ser Ala Glu Ala Leu Phe Glu Lys Ala Val
 180 185 190

Thr Pro Ser Asp Val Gly Lys Leu Asn Arg Leu Val Ile Pro Lys His
 195 200 205

His Ala Glu Lys His Phe Pro Leu Pro Ser Ser Asn Val Ser Val Lys
 210 215 220

Gly Val Leu Leu Asn Phe Glu Asp Val Asn Gly Lys Val Trp Arg Phe
 225 230 235 240

Arg Tyr Ser Tyr Trp Asn Ser Ser Gln Ser Tyr Val Leu Thr Lys Gly
 245 250 255

Trp Ser Arg Phe Val Lys Glu Lys Asn Leu Arg Ala Gly Asp Val Val
 260 265 270

Ser Phe Ser Arg Ser Asn Gly Gln Asp Gln Gln Leu Tyr Ile Gly Trp
 275 280 285

Lys Ser Arg Ser Gly Ser Asp Leu Asp Ala Gly Arg Val Leu Arg Leu
 290 295 300

Phe Gly Val Asn Ile Ser Pro Glu Ser Ser Arg Asn Asp Val Val Gly
 305 310 315 320

Asn Lys Arg Val Asn Asp Thr Glu Met Leu Ser Leu Val Cys Ser Lys
 325 330 335

Lys Gln Arg Ile Phe His Ala Ser
 340

<210> 67

<211> 984

<212> DNA

<213> Arabidopsis thaliana

<220>

<223> G659

<400> 67

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 tctctcccca aacaatctgg tatgtcatg ctttgcattt cacaatcaaa gcaaaaggct 180
 cttcaattgt tttttctttt ctttatgatt ctgaatgtat atatatgcaa aaatgaaggg 240
 ctattgaggt gtgggaagag ttgtcgatca aggtggatta actatcttag gccagatctg 300
 aagcgtggca acttcacttc agaggagaa gaaacaatca ttaagcttca ccacaactat 360

gggaacaagt ggtcgaaaat cgcttctcaa cttccaggta gaacagataa cgagatcaag 420
 aatgtgtggc acactcatct aaagaaaaga ctggctcaga gctcaggaac tgcagatgaa 480
 cccgcctcgc cttgttcgag tgattctgtt tctcggtggaa aagatgataa gtcatctcac 540
 gtagaagatt ctttgaacag agagactaat cataggaatg agttgtctac atctatgtct 600
 tctgggggtt ccaaccaaca agatgatcca aagatagacg aactcagggtt tgagtatata 660
 gaagaagctt atagcgagtt taacgacatt attattcaag aggtagacaa acccgatctg 720
 ctggagatac catttggattc agatcctgac atttgaggtt tcttagatac ttcaaactca 780
 tttcaacaat ccactgcaaa tgagaacagc tcaggtcaa gagcaacaac agaagaagag 840
 tctgatgagg atgaggttaa gaaatggttc aagcacctag aaagcgaact cgggtagaa 900
 gaagacgata atcaacaaca atacaagaa gaagaatcat catcatcatc actcttgaag 960
 aactacgagc tcatgataca ttga 984

<210> 68
 <211> 327
 <212> PRT
 <213> *Arabidopsis thaliana*

<220>
 <223> G659

<400> 68
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 Gly Pro Trp Ser Pro Glu Glu Asp Ile Lys Leu Ile Ser Phe Ile Gln
 20 25 30
 Lys Phe Gly His Glu Asn Trp Arg Ser Leu Pro Lys Gln Ser Gly Met
 35 40 45
 Ser Leu Leu Leu Ser Ser Gln Ser Lys Gln Lys Pro Leu Gln Leu Phe
 50 55 60
 Phe Leu Phe Phe Met Ile Leu Asn Val Tyr Ile Cys Lys Asn Glu Gly
 65 70 75 80
 Leu Leu Arg Cys Gly Lys Ser Cys Arg Leu Arg Trp Ile Asn Tyr Leu
 85 90 95
 Arg Pro Asp Leu Lys Arg Gly Asn Phe Thr Ser Glu Glu Glu Thr
 100 105 110
 Ile Ile Lys Leu His His Asn Tyr Gly Asn Lys Trp Ser Lys Ile Ala
 115 120 125
 Ser Gln Leu Pro Gly Arg Thr Asp Asn Glu Ile Lys Asn Val Trp His
 130 135 140
 Thr His Leu Lys Lys Arg Leu Ala Gln Ser Ser Gly Thr Ala Asp Glu
 145 150 155 160
 Pro Ala Ser Pro Cys Ser Ser Asp Ser Val Ser Arg Gly Lys Asp Asp
 165 170 175
 Lys Ser Ser His Val Glu Asp Ser Leu Asn Arg Glu Thr Asn His Arg
 180 185 190

Asn Glu Leu Ser Thr Ser Met Ser Ser Gly Gly Ser Asn Gln Gln Asp
 195 200 205

Asp Pro Lys Ile Asp Glu Leu Arg Phe Glu Tyr Ile Glu Glu Ala Tyr
 210 215 220

Ser Glu Phe Asn Asp Ile Ile Ile Gln Glu Val Asp Lys Pro Asp Leu
 225 230 235 240

Leu Glu Ile Pro Phe Asp Ser Asp Pro Asp Ile Trp Ser Phe Leu Asp
 245 250 255

Thr Ser Asn Ser Phe Gln Gln Ser Thr Ala Asn Glu Asn Ser Ser Gly
 260 265 270

Ser Arg Ala Thr Thr Glu Glu Glu Ser Asp Glu Asp Glu Val Lys Lys
 275 280 285

Trp Phe Lys His Leu Glu Ser Glu Leu Gly Leu Glu Glu Asp Asp Asn
 290 295 300

Gln Gln Gln Tyr Lys Glu Glu Glu Ser Ser Ser Ser Ser Leu Leu Lys
 305 310 315 320

Asn Tyr Glu Leu Met Ile His
 325

<210> 69

<211> 826

<212> DNA

<213> *Arabidopsis thaliana*

<220>

<223> G620

<400> 69

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 cgtgagcaag accaatacat gccaatcgca aacgtcataa gaatcatgctg taaaaccta 180
 ccgtctcacg ccaaaatctc tgacgacgccc aaagaaaacgta ttcaagaatgtgtctccgag 240
 tacatcatcgat tcgtgaccgg tgaagccaaac gagcgttgcc aacgtgagca acgtaagacc 300
 ataactgctg aagatatacct ttgggctatg agcaagctt ggttcgataa ctacgtggac 360
 cccctcaccg tggcattaa ccggtaccgt gagatagaga ccgatcgtgg ttctgcactt 420
 agaggtgagc caccgtcggtt gagacaaaacc tatggaggaa atggattttt gtttacggc 480
 ccatctcatg gcctacacctc tccgggtcct tatgttatg gtatgttggc ccaatccatg 540
 gttatgggag gtggctggta ctacaaaac gggtcgtcggtt gtcagatgtt atccagtgtt 600
 ggtgggtggctt cttcgcttcc cattaacgga atgcggctt ttgaccattt tggcgttat 660
 aagtgaagaa ggagttattt ttcatttttatacttca aaacatgtgtt ttcgtatgtt 720
 attttatattt tatgtcttat caataaacatt tctatataat gttgcttctt taaggaaaag 780
 tggatgtatgtt caataacttta tgagaaactg atttatatat gcaat 826

<210> 70

<211> 208

<212> PRT

<213> *Arabidopsis thaliana*

<220>
<223> G620

<400> 70
Met Thr Ser Ser Val Ile Val Ala Gly Ala Gly Asp Lys Asn Asn Gly
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Ile Val Val Gln Gln Gln Pro Pro Cys Val Ala Arg Glu Gln Asp Gln
20 25 30

Tyr Met Pro Ile Ala Asn Val Ile Arg Ile Met Arg Lys Thr Leu Pro
35 40 45

Ser His Ala Lys Ile Ser Asp Asp Ala Lys Glu Thr Ile Gln Glu Cys
50 55 60

Val Ser Glu Tyr Ile Ser Phe Val Thr Gly Glu Ala Asn Glu Arg Cys
65 70 75 80

Gln Arg Glu Gln Arg Lys Thr Ile Thr Ala Glu Asp Ile Leu Trp Ala
85 90 95

Met Ser Lys Leu Gly Phe Asp Asn Tyr Val Asp Pro Leu Thr Val Phe
100 105 110

Ile Asn Arg Tyr Arg Glu Ile Glu Thr Asp Arg Gly Ser Ala Leu Arg
115 120 125

Gly Glu Pro Pro Ser Leu Arg Gln Thr Tyr Gly Gly Asn Gly Ile Gly
130 135 140

Phe His Gly Pro Ser His Gly Leu Pro Pro Pro Gly Pro Tyr Gly Tyr
145 150 155 160

Gly Met Leu Asp Gln Ser Met Val Met Gly Gly Arg Tyr Tyr Gln
165 170 175

Asn Gly Ser Ser Gly Gln Asp Glu Ser Ser Val Gly Gly Ser Ser
180 185 190

Ser Ser Ile Asn Gly Met Pro Ala Phe Asp His Tyr Gly Gln Tyr Lys
195 200 205

<210> 71
<211> 1394
<212> DNA
<213> Arabidopsis thaliana

<220>
<223> G596

<400> 71
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cctcaagcta agattctggt tttgtgagtt gagtggatga gaagaggaga gattaactaa 120
attagggttt caattgttta cttttgttt gcttttata tcaagtaatg gatcagggtct 180
ctcgctctc tcctccaccc tttctctcaa gagatctcca tcttcaccca caccatcaat 240
tccagcatca gcagcagcag cagcaacaga atcacggcca cgatataagac cagcaccgaa 300

tcgggtggct aaaacgtgac cgagatgctg atatcgatcc caacgagcac tcttcagccg 360
 gaaaagatca aagtactcct ggctccggtg gagaaagccg cggcggagga ggaggagata 420
 atcacatcac gagaaggcca cgtggcagac cagcgggatc taagaacaaa ccaaaaccgc 480
 caatcatcat cactcgagac agcgcaaacg ctctcaaatac tcatgtcatg gaagtagcaa 540
 acggatgtga cgtcatggaa agtgtcaccg tcttcgctcg ccgtcgccaa cgtggcatct 600
 gcgttttagg cggaaacggc gccgttacca acgttaccat aagacaacca gcttcagtag 660
 ctgggtgtgg ctcatctgtc gttaacttac acggacgtt cgagatttct tctctctcg 720
 gatcattcct tcctccctcg gctccaccag ctgcgtcagg tctaacgatt tacttagccg 780
 gtggtcaggg acagggtgtt ggaggaagcg tggttggtcc actcatggct tcaggacctg 840
 tagtgattat ggcagcttcg tttggaaacg ctgcgtatga gagactgccc ttggaggaag 900
 acgatcaaga agagcaaaca gctggagccg ttgctaataa tatcgatgga aacgcaacaa 960
 tgggtgtgg aacgcaaacg caaactcaga cgcagcagca acagcaacaa cagttgatgc 1020
 aagatccgac gtcgttata caagggttgc ctccgaatct tatgaattct gttcaattgc 1080
 cagctgaagc ttattggga actccgagac catcttcta aatcgcaag aaaaaacaag 1140
 ttagatacgt tcgttgttt taatttataa tctctttct gtcaagttt aattttctt 1200
 ttcttcttct ttgtttctta aagataattg tagtcttga cgaagattcg tggtagt 1260
 gaatcgaaga gaatcgaaaa ggtcatggga ttgctcgatc tattagttt gagaggggt 1320
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 gtgttggtga taaa 1394

<210> 72
 <211> 317
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G596

<400> 72
 Met Asp Gln Val Ser Arg Ser Leu Pro Pro Pro Phe Leu Ser Arg Asp
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Leu His Leu His Pro His His Gln Phe Gln His Gln Gln Gln Gln
 20 25 30

Gln Gln Asn His Gly His Asp Ile Asp Gln His Arg Ile Gly Gly Leu
 35 40 45

Lys Arg Asp Arg Asp Ala Asp Ile Asp Pro Asn Glu His Ser Ser Ala
 50 55 60

Gly Lys Asp Gln Ser Thr Pro Gly Ser Gly Gly Glu Ser Gly Gly Gly
 65 70 75 80

Gly Gly Gly Asp Asn His Ile Thr Arg Arg Pro Arg Gly Arg Pro Ala
 85 90 95

Gly Ser Lys Asn Lys Pro Lys Pro Pro Ile Ile Ile Thr Arg Asp Ser
 100 105 110

Ala Asn Ala Leu Lys Ser His Val Met Glu Val Ala Asn Gly Cys Asp
 115 120 125

Val Met Glu Ser Val Thr Val Phe Ala Arg Arg Gln Arg Gly Ile
 130 135 140

Cys Val Leu Ser Gly Asn Gly Ala Val Thr Asn Val Thr Ile Arg Gln
 145 150 155 160

Pro Ala Ser Val Pro Gly Gly Ser Ser Val Val Asn Leu His Gly
 165 170 175

Arg Phe Glu Ile Leu Ser Leu Ser Gly Ser Phe Leu Pro Pro Pro Ala
 180 185 190

Pro Pro Ala Ala Ser Gly Leu Thr Ile Tyr Leu Ala Gly Gly Gln Gly
 195 200 205

Gln Val Val Gly Gly Ser Val Val Gly Pro Leu Met Ala Ser Gly Pro
 210 215 220

Val Val Ile Met Ala Ala Ser Phe Gly Asn Ala Ala Tyr Glu Arg Leu
 225 230 235 240

Pro Leu Glu Glu Asp Asp Gln Glu Glu Gln Thr Ala Gly Ala Val Ala
 245 250 255

Asn Asn Ile Asp Gly Asn Ala Thr Met Gly Gly Gly Thr Gln Thr Gln
 260 265 270

Thr Gln Thr Gln Gln Gln Gln Gln Gln Leu Met Gln Asp Pro Thr
 275 280 285

Ser Phe Ile Gln Gly Leu Pro Pro Asn Leu Met Asn Ser Val Gln Leu
 290 295 300

Pro Ala Glu Ala Tyr Trp Gly Thr Pro Arg Pro Ser Phe
 305 310 315

<210> 73

<211> 913

<212> DNA

<213> Arabidopsis thaliana

<220>

<223> G511

<400> 73

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 ttctatccca cggaaagaaga actggtttcg ttctacctac gaaaccagct cgaaggaagg 120
 agtgtatgact caatgcattcg tgtcattccc gtacttgacg tctttgaggt cgagcctagt 180
 catcttccaa atgttgctgg agtgagatgt cgaggagacg ctgagcaatg gttcttcttc 240
 gtgccacgac aagaacgcga agcaagagga ggcagacccg gtagaactac tggttcagga 300
 tactggaaag caactggatc acctggtcca gtctttccaa aagacaacaa aatgatttgg 360
 gcaaaagaaaa ctatggttt ctacactgga aaagcaccctt caggaagaaaa aactaaatgg 420
 aaaatgaatg agtaccacgc cgttgcacgaa acagtcaacg cttccacaat ccctaagctg 480
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 agacgtcctg agggagttt gcagacagag agaatgctaa caagtgtatgt tgcagtagct 600
 gagacatcgt tccgtgtgga aagctcactg gaaacttcga tttcaggagg agaacatatt 660
 gatgtctcta tgaacacaga gtttggatggactatcag aaccgatgtg ggactggaa 720
 cagctgactt ggccttgaag ctatatacat tttataatca agcaaattta aacttggttc 780
 aattgcttat tgtagttt aattttatga cccgaaagat tcttttctt tctttacctt 840
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aaaaaaaaaaa aaa

913

<210> 74
<211> 235
<212> PRT
<213> *Arabidopsis thaliana*

<220>
<223> G511

<400> 74
 Met Ala Asp Glu Val Thr Ile Gly Phe Arg Phe Tyr Pro Thr Glu Glu
 1 5 10 15

 Glu Leu Val Ser Phe Tyr Leu Arg Asn Gln Leu Glu Gly Arg Ser Asp
 20 25 30

 Asp Ser Met His Arg Val Ile Pro Val Leu Asp Val Phe Glu Val Glu
 35 40 45

 Pro Ser His Leu Pro Asn Val Ala Gly Val Arg Cys Arg Gly Asp Ala
 50 55 60

 Glu Gln Trp Phe Phe Phe Val Pro Arg Gln Glu Arg Glu Ala Arg Gly
 65 70 75 80

 Gly Arg Pro Ser Arg Thr Thr Gly Ser Gly Tyr Trp Lys Ala Thr Gly
 85 90 95

 Ser Pro Gly Pro Val Phe Ser Lys Asp Asn Lys Met Ile Gly Ala Lys
 100 105 110

 Lys Thr Met Val Phe Tyr Thr Gly Lys Ala Pro Thr Gly Arg Lys Thr
 115 120 125

 Lys Trp Lys Met Asn Glu Tyr His Ala Val Asp Glu Thr Val Asn Ala
 130 135 140

 Ser Thr Ile Pro Lys Leu Arg Arg Glu Phe Ser Leu Cys Arg Val Tyr
 145 150 155 160

 Ile Thr Thr Gly Ser Ser Arg Ala Phe Asp Arg Arg Pro Glu Gly Val
 165 170 175

 Leu Gln Thr Glu Arg Met Leu Thr Ser Asp Val Ala Val Ala Glu Thr
 180 185 190

 Ser Phe Arg Val Glu Ser Ser Leu Glu Thr Ser Ile Ser Gly Gly Glu
 195 200 205

 His Ile Asp Val Ser Met Asn Thr Glu Phe Val Asp Gly Leu Ser Glu
 210 215 220

 Pro Met Trp Asp Trp Glu Gln Leu Thr Trp Pro
 225 230 235

<210> 75
 <211> 2332
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G471

<400> 75
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 ctctggcatg cctgtgttgg acctcttggta accctacctc gtgaagggga acgagttat 240
 tatttccctg aaggccacat ggagcagctc gaggcatcaa tgcaccaagg tttggagcaa 300
 cagatgcctt cttcaaccc cccatctaag atcctctgtt aagttatcaa catccagcgc 360
 agggcagagc cccagactga cgaagtataat ggcgaaataa ctttattgcc agaactggat 420
 caaagcgaac ccactagccc agatgcccgtt gttcaagaac ctgaaaagtg caccgtacat 480
 tcattttgca agacactaac tgcttcagac acaagcacac atgggtggctt ctcgggtcta 540
 cggagacatg cagatgattt tctcccaccc ttggatatgt cccaacaacc accgtggcaa 600
 gaatttgggtt caactgattt gcacaataatgtaatgatggcatt tttaggcacat tttccgaggc 660
 caaccaaggc gtcatttgctt aacaacttggta tggagtgttt ttgttagtgc gaagaaacta 720
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 cgtcctgaaa gagtttccacc ttgggaactt gagccctag ttgcaaatag tactccgtct 1200
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 aacaaatcat ttggagtatc tattggatca gcctttggc ccaccaatgc agatagtgc 1440
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 aatgtctgtt ggcttttgg gtttggatca gttggaaatg ttaatgttgg tgaatgtttc 1560
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 gaaaggcagaa gcaagagatc gtttggatcaat atggatgttgg agatgtgtt gttatgttgc 2220
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<210> 76
 <211> 665
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G471

<400> 76
 Met Ala Ala Ser Asn His Ser Ser Gly Lys Pro Gly Gly Val Leu Ser
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 Asp Ala Leu Cys Arg Glu Leu Trp His Ala Cys Ala Gly Pro Leu Val
 20 25 30
 Thr Leu Pro Arg Glu Gly Glu Arg Val Tyr Tyr Phe Pro Glu Gly His
 35 40 45
 Met Glu Gln Leu Glu Ala Ser Met His Gln Gly Leu Glu Gln Gln Met
 50 55 60
 Pro Ser Phe Asn Leu Pro Ser Lys Ile Leu Cys Lys Val Ile Asn Ile
 65 70 75 80
 Gln Arg Arg Ala Glu Pro Glu Thr Asp Glu Val Tyr Ala Gln Ile Thr
 85 90 95
 Leu Leu Pro Glu Leu Asp Gln Ser Glu Pro Thr Ser Pro Asp Ala Pro
 100 105 110
 Val Gln Glu Pro Glu Lys Cys Thr Val His Ser Phe Cys Lys Thr Leu
 115 120 125
 Thr Ala Ser Asp Thr Ser Thr His Gly Gly Phe Ser Val Leu Arg Arg
 130 135 140
 His Ala Asp Asp Cys Leu Pro Pro Leu Asp Met Ser Gln Gln Pro Pro
 145 150 155 160
 Trp Gln Glu Leu Val Ala Thr Asp Leu His Asn Ser Glu Trp His Phe
 165 170 175
 Arg His Ile Phe Arg Gly Gln Pro Arg Arg His Leu Leu Thr Thr Gly
 180 185 190
 Trp Ser Val Phe Val Ser Ser Lys Lys Leu Val Ala Gly Asp Ala Phe
 195 200 205
 Ile Phe Leu Arg Gly Glu Asn Glu Glu Leu Arg Val Gly Val Arg Arg
 210 215 220
 His Met Arg Gln Gln Thr Asn Ile Pro Ser Ser Val Ile Ser Ser His
 225 230 235 240
 Ser Met His Ile Gly Val Leu Ala Thr Ala Ala His Ala Ile Thr Thr
 245 250 255
 Gly Thr Ile Phe Ser Val Phe Tyr Lys Pro Arg Thr Ser Arg Ser Glu
 260 265 270
 Phe Ile Val Ser Val Asn Arg Tyr Leu Glu Ala Lys Thr Gln Lys Leu
 275 280 285
 Ser Val Gly Met Arg Phe Lys Met Arg Phe Glu Gly Glu Glu Ala Pro
 290 295 300

Glu Lys Arg Phe Ser Gly Thr Ile Val Gly Val Gln Glu Asn Lys Ser
 305 310 315 320
 Ser Val Trp His Asp Ser Glu Trp Arg Ser Leu Lys Val Gln Trp Asp
 325 330 335
 Glu Pro Ser Ser Val Phe Arg Pro Glu Arg Val Ser Pro Trp Glu Leu
 340 345 350
 Glu Pro Leu Val Ala Asn Ser Thr Pro Ser Ser Gln Pro Gln Pro Pro
 355 360 365
 Gln Arg Asn Lys Arg Pro Arg Pro Pro Gly Leu Pro Ser Pro Ala Thr
 370 375 380
 Gly Pro Ser Gly Pro Val Thr Pro Asp Gly Val Trp Lys Ser Pro Ala
 385 390 395 400
 Asp Thr Pro Ser Ser Val Pro Leu Phe Ser Pro Pro Ala Lys Ala Ala
 405 410 415
 Thr Phe Gly His Gly Gly Asn Lys Ser Phe Gly Val Ser Ile Gly Ser
 420 425 430
 Ala Phe Trp Pro Thr Asn Ala Asp Ser Ala Ala Glu Ser Phe Ala Ser
 435 440 445
 Ala Phe Asn Asn Glu Ser Thr Glu Lys Lys Gln Thr Asn Gly Asn Val
 450 455 460
 Cys Arg Leu Phe Gly Phe Glu Leu Val Glu Asn Val Asn Val Asp Glu
 465 470 475 480
 Cys Phe Ser Ala Ala Ser Val Ser Gly Ala Val Ala Val Asp Gln Pro
 485 490 495
 Val Pro Ser Asn Glu Phe Asp Ser Gly Gln Gln Ser Glu Pro Leu Asn
 500 505 510
 Ile Asn Gln Ser Asp Ile Pro Ser Gly Ser Gly Asp Pro Glu Lys Ser
 515 520 525
 Ser Leu Arg Ser Pro Gln Glu Ser Gln Ser Arg Gln Ile Arg Ser Cys
 530 535 540
 Thr Lys Val His Met Gln Gly Ser Ala Val Gly Arg Ala Ile Asp Leu
 545 550 555 560
 Thr Arg Ser Glu Cys Tyr Glu Asp Leu Phe Lys Lys Leu Glu Glu Met
 565 570 575
 Phe Asp Ile Lys Gly Glu Leu Leu Glu Ser Thr Lys Lys Trp Gln Val
 580 585 590
 Val Tyr Thr Asp Asp Glu Asp Asp Met Met Met Val Gly Asp Asp Pro
 595 600 605

Trp Asn Glu Phe Cys Gly Met Val Arg Lys Ile Phe Ile Tyr Thr Pro
 610 615 620

Glu Glu Val Lys Lys Leu Ser Pro Lys Asn Lys Leu Ala Val Asn Ala
 625 630 635 640

Arg Met Gln Leu Lys Ala Asp Ala Glu Glu Asn Gly Asn Thr Glu Gly
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<211> 2217

<212> DNA

<213> Arabidopsis thaliana

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<223> G385

<400> 77

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 gacgaacacc aactccgtct cgaaaatgtct cgattaagg aagagatcga ccgtatatcc 600
 gcaatcgca gctaaatacgt aggcaagcca gtctcaaact atccacttat gtctcctcct 660
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 <213> Arabidopsis thaliana

<220>
 <223> G385

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 Arg Asp Asp Glu Phe Asp Ser Pro Asn Thr Lys Ser Gly Ser Glu Asn
 35 40 45
 Gln Glu Gly Gly Ser Gly Asn Asp Gln Asp Pro Leu His Pro Asn Lys
 50 55 60
 Lys Lys Arg Tyr His Arg His Thr Gln Leu Gln Ile Gln Glu Met Glu
 65 70 75 80
 Ala Phe Phe Lys Glu Cys Pro His Pro Asp Asp Lys Gln Arg Lys Gln
 85 90 95
 Leu Ser Arg Glu Leu Asn Leu Glu Pro Leu Gln Val Lys Phe Trp Phe
 100 105 110
 Gln Asn Lys Arg Thr Gln Met Lys Asn His His Glu Arg His Glu Asn
 115 120 125
 Ser His Leu Arg Ala Glu Asn Glu Lys Leu Arg Asn Asp Asn Leu Arg
 130 135 140
 Tyr Arg Glu Ala Leu Ala Asn Ala Ser Cys Pro Asn Cys Gly Gly Pro
 145 150 155 160
 Thr Ala Ile Gly Glu Met Ser Phe Asp Glu His Gln Leu Arg Leu Glu
 165 170 175
 Asn Ala Arg Leu Arg Glu Glu Ile Asp Arg Ile Ser Ala Ile Ala Ala
 180 185 190
 Lys Tyr Val Gly Lys Pro Val Ser Asn Tyr Pro Leu Met Ser Pro Pro
 195 200 205
 Pro Leu Pro Pro Arg Pro Leu Glu Leu Ala Met Gly Asn Ile Gly Gly
 210 215 220
 Glu Ala Tyr Gly Asn Asn Pro Asn Asp Leu Leu Lys Ser Ile Thr Ala
 225 230 235 240

Pro Thr Glu Ser Asp Lys Pro Val Ile Ile Asp Leu Ser Val Ala Ala
 245 250 255
 Met Glu Glu Leu Met Arg Met Val Gln Val Asp Glu Pro Leu Trp Lys
 260 265 270
 Ser Leu Ala Leu Asp Glu Glu Tyr Ala Arg Thr Phe Pro Arg Gly
 275 280 285
 Ile Gly Pro Arg Pro Ala Gly Tyr Arg Ser Glu Ala Ser Arg Glu Ser
 290 295 300
 Ala Val Val Ile Met Asn His Val Asn Ile Val Glu Ile Leu Met Asp
 305 310 315 320
 Val Asn Gln Trp Ser Thr Ile Phe Ala Gly Met Val Ser Arg Ala Met
 325 330 335
 Thr Leu Ala Val Leu Ser Thr Gly Val Ala Gly Asn Tyr Asn Gly Ala
 340 345 350
 Leu Gln Val Met Ser Ala Glu Phe Gln Val Pro Ser Pro Leu Val Pro
 355 360 365
 Thr Arg Glu Thr Tyr Phe Ala Arg Tyr Cys Lys Gln Gln Gly Asp Gly
 370 375 380
 Ser Trp Ala Val Val Asp Ile Ser Leu Asp Ser Leu Gln Pro Asn Pro
 385 390 395 400
 Pro Ala Arg Cys Arg Arg Ala Ser Gly Cys Leu Ile Gln Glu Leu
 405 410 415
 Pro Asn Gly Tyr Ser Lys Val Thr Trp Val Glu His Val Glu Val Asp
 420 425 430
 Asp Arg Gly Val His Asn Leu Tyr Lys His Met Val Ser Thr Gly His
 435 440 445
 Ala Phe Gly Ala Lys Arg Trp Val Ala Ile Leu Asp Arg Gln Cys Glu
 450 455 460
 Arg Leu Ala Ser Val Met Ala Thr Asn Ile Ser Ser Gly Glu Val Gly
 465 470 475 480
 Val Ile Thr Asn Gln Glu Gly Arg Arg Ser Met Leu Lys Leu Ala Glu
 485 490 495
 Arg Met Val Ile Ser Phe Cys Ala Gly Val Ser Ala Ser Thr Ala His
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 Thr Trp Thr Thr Leu Ser Gly Thr Gly Ala Glu Asp Val Arg Val Met
 515 520 525
 Thr Arg Lys Ser Val Asp Asp Pro Gly Arg Ser Pro Gly Ile Val Leu
 530 535 540

Ser Ala Ala Thr Ser Phe Trp Ile Pro Val Pro Pro Lys Arg Val Phe
 545 550 555 560
 Asp Phe Leu Arg Asp Glu Asn Ser Arg Asn Glu Trp Asp Ile Leu Ser
 565 570 575
 Asn Gly Gly Val Val Gln Glu Met Ala His Ile Ala Asn Gly Arg Asp
 580 585 590
 Thr Gly Asn Cys Val Ser Leu Leu Arg Val Asn Ser Ala Asn Ser Ser
 595 600 605
 Gln Ser Asn Met Leu Ile Leu Gln Glu Ser Cys Ile Asp Pro Thr Ala
 610 615 620
 Ser Phe Val Ile Tyr Ala Pro Val Asp Ile Val Ala Met Asn Ile Val
 625 630 635 640
 Leu Asn Gly Gly Asp Pro Asp Tyr Val Ala Leu Leu Pro Ser Gly Phe
 645 650 655
 Ala Ile Leu Pro Asp Gly Asn Ala Asn Ser Gly Ala Pro Gly Gly Asp
 660 665 670
 Gly Gly Ser Leu Leu Thr Val Ala Phe Gln Ile Leu Val Asp Ser Val
 675 680 685
 Pro Thr Ala Lys Leu Ser Leu Gly Ser Val Ala Thr Val Asn Asn Leu
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 Ile Ala Cys Thr Val Glu Arg Ile Lys Ala Ser Met Ser Cys Glu Thr
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 <211> 1857
 <212> DNA
 <213> *Arabidopsis thaliana*

<220>
 <223> G261

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 tcaagttcga agctttttt ggagggaatt ttgggcttct gattttgatc gaaaacttact 240
 gatagttgtt tctttgatc ctccttaact gtatgttctg tttttttttt tttttttttt 300
 gaaagttttt atctttttt gttattgaaa ctttcataat ttgtatcaaaa gagtctttt 360
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 atccgattct atcgtctttt ggagtcaagag caataagatg tttcatcgat tttttttttt 600
 ggagtttttctt agagatcttcc ttccatgtt gttttttttt tttttttttt tttttttttt 660
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 gttacataaa caagacgagg aacgagaatg gttttagatg caagtgaaag aactaaaga 960
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 agtacttagg tatggttcag ctgtttattt atcacttgta tgatcttcc cagttcattt 1800
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 <211> 401
 <212> PRT
 <213> *Arabidopsis thaliana*

<220>
 <223> G261

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Val Ser Trp Ser Gln Ser Asn Lys Ser Phe Ile Val Trp Asn Pro Pro
 35 40 45

Glu Phe Ser Arg Asp Leu Leu Pro Arg Phe Phe Lys His Asn Asn Phe
 50 55 60

Ser Ser Phe Ile Arg Gln Leu Asn Thr Tyr Gly Phe Arg Lys Ala Asp
 65 70 75 80

Pro Glu Gln Trp Glu Phe Ala Asn Asp Asp Phe Val Arg Gly Gln Pro
 85 90 95

His Leu Met Lys Asn Ile His Arg Arg Lys Pro Val His Ser His Ser
 100 105 110

Leu Pro Asn Leu Gln Ala Gln Leu Asn Pro Leu Thr Asp Ser Glu Arg
 115 120 125

Val Arg Met Asn Asn Gln Ile Glu Arg Leu Thr Lys Glu Lys Glu Gly
 130 135 140

Leu Leu Glu Glu Leu His Lys Gln Asp Glu Glu Arg Glu Val Phe Glu
 145 150 155 160
 Met Gln Val Lys Glu Leu Lys Glu Arg Leu Gln His Met Glu Lys Arg
 165 170 175
 Gln Lys Thr Met Val Ser Phe Val Ser Gln Val Leu Glu Lys Pro Gly
 180 185 190
 Leu Ala Leu Asn Leu Ser Pro Cys Val Pro Glu Thr Asn Glu Arg Lys
 195 200 205
 Arg Arg Phe Pro Arg Ile Glu Phe Phe Pro Asp Glu Pro Met Leu Glu
 210 215 220
 Glu Asn Lys Thr Cys Val Val Val Arg Glu Glu Gly Ser Thr Ser Pro
 225 230 235 240
 Ser Ser His Thr Arg Glu His Gln Val Glu Gln Leu Glu Ser Ser Ile
 245 250 255
 Ala Ile Trp Glu Asn Leu Val Ser Asp Ser Cys Glu Ser Met Leu Gln
 260 265 270
 Ser Arg Ser Met Met Thr Leu Asp Val Asp Glu Ser Ser Thr Phe Pro
 275 280 285
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 290 295 300
 Lys Ser Pro Pro Ser Pro Arg Ile Ile Asp Met Asn Cys Glu Pro Asp
 305 310 315 320
 Gly Ser Lys Glu Gln Asn Thr Val Ala Ala Pro Pro Pro Pro Val
 325 330 335
 Ala Gly Ala Asn Asp Gly Phe Trp Gln Gln Phe Phe Ser Glu Asn Pro
 340 345 350
 Gly Ser Thr Glu Gln Arg Glu Val Gln Leu Glu Arg Lys Asp Asp Lys
 355 360 365
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 <211> 751
 <212> DNA
 <213> Arabidopsis thaliana

<220>

<223> G25

<400> 81

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 ctcagtgagt gaagaaaagag atgggaaacg agagaggaag aatctgtaca gagggataag 240
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 cgatactaaa ccagggggga atcaaaaatga gctgattcg gaaaaccaag tagagagctt 480
 atcgaggac ctgatggcat tggaggatta catgagattc tatcagattc cggttgcga 540
 cgaccaatcg gcgaccgata ttggaaattt atggagctat caagactcca attaaatctc 600
 ttatcccgc gccgggttgc tcactcatatgcgtctt aatttacttg tttttactt 660
 aacaatcaag tctaatttgt ttccatcaat atttcagata agagtaaagc ttcaattgtc 720
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<212> PRT

<213> Arabidopsis thaliana

<220>

<223> G25

<400> 82

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					20			25				30			

Pro	Val	Ser	Val	Ser	Glu	Glu	Arg	Asp	Gly	Lys	Arg	Glu	Arg	Lys	Asn
					35		40				45				

Leu	Tyr	Arg	Gly	Ile	Arg	Gln	Arg	Pro	Trp	Gly	Lys	Trp	Ala	Ala	Glu
				50		55				60					

Ile	Arg	Asp	Pro	Ser	Lys	Gly	Val	Arg	Val	Trp	Leu	Gly	Thr	Phe	Lys
					65		70		75			80			

Thr	Ala	Asp	Glu	Ala	Ala	Arg	Ala	Tyr	Asp	Val	Ala	Ala	Ile	Lys	Ile
					85			90				95			

Arg	Gly	Arg	Lys	Ala	Lys	Leu	Asn	Phe	Pro	Asn	Thr	Gln	Val	Glu	Glu
				100			105				110				

Glu	Ala	Asp	Thr	Lys	Pro	Gly	Gly	Asn	Gln	Asn	Glu	Leu	Ile	Ser	Glu
				115			120				125				

Asn	Gln	Val	Glu	Ser	Leu	Ser	Glu	Asp	Leu	Met	Ala	Leu	Glu	Asp	Tyr
					130		135			140					

Met	Arg	Phe	Tyr	Gln	Ile	Pro	Val	Ala	Asp	Asp	Gln	Ser	Ala	Thr	Asp
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Ile Gly Asn Leu Trp Ser Tyr Gln Asp Ser Asn
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<213> Arabidopsis thaliana
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<223> G610

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<212> PRT
<213> Arabidopsis thaliana
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<220>

<223> G610

<400> 84

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									25					30	

Gly	Lys	Arg	Ser	Asp	Asp	Glu	Ser	Glu	Ile	Cys	Ala	Ile	Asp	Leu	Leu
									35				40	45	

Ala	Ser	Leu	Ala	Gly	Lys	Leu	Leu	Glu	Ser	Glu	Ser	Ser	Ser	Thr	
									50				55	60	

Ser	Thr	Tyr	Ala	Ser	Glu	Ala	Asp	Asn	Leu	Asp	His	Leu	Gly	Gly	Leu
									65			75		80	

Ile	Lys	Gln	Glu	Leu	Glu	Asp	Gly	Tyr	Thr	Thr	Lys	Pro	Cys	Lys	Ser
									85			90		95	

Glu	Phe	Phe	Asp	Pro	Gly	Asn	Pro	Ala	Ser	Lys	Ser	Thr	Ser	Glu	Asn
									100			105		110	

Thr	Ser	Val	Thr	Cys	Leu	Pro	Phe	Ser	Ser	Phe	Glu	Asn	Asp	Cys	Ile
									115			120		125	

Leu	Glu	Gln	Thr	Pro	Val	Ser	Asp	Cys	Lys	Arg	Ala	Ser	Gly	Leu	Lys
									130			135		140	

Ser	Leu	Val	Gly	Ser	Ile	Thr	Glu	Glu	Thr	Cys	Val	Val	Asn	Glu	Asp
									145			150		155	

Ala	Gly	Ser	Glu	Gln	Gly	Ala	Asn	Thr	Phe	Ser	Leu	Lys	Asp	Pro	Ser
									165			170		175	

Gln	Leu	His	Ser	Gln	Ser	Pro	Glu	Ser	Val	Leu	Leu	Asp	Gly	Asp	Val
									180			185		190	

Lys	Leu	Ala	Pro	Cys	Thr	Asp	Gln	Val	Pro	Asn	Asp	Ser	Phe	Lys	Gly
									195			200		205	

Tyr	Arg	Asn	His	Ser	Lys	Leu	Val	Cys	Arg	Asp	Asp	Asp	Glu	Asn	Tyr
									210			215		220	

Cys	Lys	Tyr	Tyr	Lys	Phe	Ser	Asp	Lys	Cys	Lys	Ser	Tyr	Arg	Pro	Leu
									225			230		235	

Ser	Arg	Val	Gly	Asn	Arg	Arg	Ile	Met	Gln	Ser	Val	Arg	Ala	Ile	Ser
									245			250		255	

Lys	Leu	Lys	Cys	Phe	Glu	Asp	Thr	Arg	Thr	Asp	Gly	Arg	Leu	Lys	Ala
									260			265		270	

Leu	Tyr	Arg	Lys	Arg	Lys	Leu	Cys	Tyr	Gly	Tyr	Asn	Pro	Trp	Lys	Arg
									275			280		285	

Glu Thr Ile His Arg Lys Arg Arg Leu Ser Asp Lys Gly Leu Val Val
 290 295 300
 Asn Tyr Asp Gly Gly Leu Ser Ser Glu Ser Val Ser Asn Ser Pro Glu
 305 310 315 320
 Lys Gly Glu Ser Glu Asn Gly Asp Phe Ser Ala Ala Lys Ile Gly Leu
 325 330 335
 Leu Ser Lys Asp Ser Arg Val Lys Phe Ser Ile Lys Ser Leu Arg Ile
 340 345 350
 Pro Glu Leu Val Ile Glu Val Pro Glu Thr Ala Thr Val Gly Leu Leu
 355 360 365
 Lys Arg Thr Val Lys Glu Ala Val Thr Ala Leu Leu Gly Gly Ile
 370 375 380
 Arg Ile Gly Val Leu Val Gln Gly Lys Lys Val Arg Asp Asp Asn Asn
 385 390 395 400
 Thr Leu Ser Gln Thr Gly Leu Ser Cys Arg Glu Asn Leu Gly Asn Leu
 405 410 415
 Gly Phe Thr Leu Glu Pro Gly Leu Glu Thr Leu Pro Val Pro Leu Cys
 420 425 430
 Ser Glu Thr Pro Val Leu Ser Leu Pro Thr Asp Ser Thr Lys Leu Ser
 435 440 445
 Glu Arg Ser Ala Ala Ser Pro Ala Leu Glu Thr Gly Ile Pro Leu Pro
 450 455 460
 Pro Gln Asp Glu Asp Tyr Leu Ile Asn Leu Gly Asn Ser Val Glu Asn
 465 470 475 480
 Asn Asp Glu Leu Val Pro His Leu Ser Asp Ile Pro Ala Asp Glu Gln
 485 490 495
 Pro Ser Ser Asp Ser Arg Ala Leu Val Pro Val Leu Ala Leu Glu Ser
 500 505 510
 Asp Ala Leu Ala Leu Val Pro Val Asn Glu Lys Pro Lys Arg Thr Glu
 515 520 525
 Leu Ser Gln Arg Arg Thr Arg Arg Leu Phe Ser Val Thr Glu Val Glu
 530 535 540
 Ala Leu Val Ser Ala Val Glu Glu Val Gly Thr Gly Arg Trp Arg Asp
 545 550 555 560
 Val Lys Leu Arg Ser Phe Glu Asn Ala Ser His Arg Thr Tyr Val Asp
 565 570 575
 Leu Lys Asp Lys Trp Lys Thr Leu Val His Thr Ala Ser Ile Ser Pro
 580 585 590

Gln Gln Arg Arg Gly Glu Pro Val Pro Gln Glu Leu Leu Asp Arg Val
 595 600 605

Leu Gly Ala His Arg Tyr Trp Thr Gln His Gln Met Lys Gln Asn Gly
 610 615 620

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 <212> DNA
 <213> *Arabidopsis thaliana*

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 <223> G229

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 aaggtgtgga aagagctgta gattgagatg gataaactat ctaagatcag acctcaagcg 240
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 <211> 371
 <212> PRT
 <213> *Arabidopsis thaliana*

<220>
 <223> G229

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Trp Thr Ala Glu Glu Asp Gln Ile Leu Ser Asn Tyr Ile Gln Ser Asn
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Gly Glu Gly Ser Trp Arg Ser Leu Pro Lys Asn Ala Gly Leu Lys Arg
 35 40 45

Cys Gly Lys Ser Cys Arg Leu Arg Trp Ile Asn Tyr Leu Arg Ser Asp
 50 55 60

Leu Lys Arg Gly Asn Ile Thr Pro Glu Glu Glu Leu Val Val Lys
 65 70 75 80

Leu His Ser Thr Leu Gly Asn Arg Trp Ser Leu Ile Ala Gly His Leu
 85 90 95

Pro Gly Arg Thr Asp Asn Glu Ile Lys Asn Tyr Trp Asn Ser His Leu
 100 105 110

Ser Arg Lys Leu His Asn Phe Ile Arg Lys Pro Ser Ile Ser Gln Asp
 115 120 125

Val Ser Ala Val Ile Met Ala Asn Ala Ser Ser Ala Pro Pro Pro Pro
 130 135 140

Gln Ala Lys Arg Arg Leu Gly Arg Thr Ser Arg Ser Ala Met Lys Pro
 145 150 155 160

Lys Ile Arg Arg Thr Lys Thr Arg Lys Thr Lys Lys Thr Ser Ala Pro
 165 170 175

Pro Glu Pro Asn Ala Asp Val Ala Gly Ala Asp Lys Glu Ala Leu Met
 180 185 190

Val Glu Ser Ser Gly Ala Glu Ala Glu Leu Gly Arg Pro Cys Asp Tyr
 195 200 205

Tyr Gly Asp Asp Cys Asn Lys Asn Leu Met Ser Ile Asn Gly Asp Asn
 210 215 220

Gly Val Leu Thr Phe Asp Asp Ile Ile Asp Leu Leu Leu Asp Glu
 225 230 235 240

Ser Asp Pro Gly His Leu Tyr Thr Asn Thr Thr Cys Gly Gly Gly
 245 250 255

Glu Leu His Asn Ile Arg Asp Ser Glu Gly Ala Arg Gly Phe Ser Asp
 260 265 270

Thr Trp Asn Gln Gly Asn Leu Asp Cys Leu Leu Gln Ser Cys Pro Ser
 275 280 285

Val Glu Ser Phe Leu Asn Tyr Asp His Gln Val Asn Asp Ala Ser Thr
 290 295 300

Asp Glu Phe Ile Asp Trp Asp Cys Val Trp Gln Glu Gly Ser Asp Asn
 305 310 315 320

Asn Leu Trp His Glu Lys Glu Asn Pro Asp Ser Met Val Ser Trp Leu
 325 330 335

Leu Asp Gly Asp Asp Glu Ala Thr Ile Gly Asn Ser Asn Cys Glu Asn
 340 345 350

Phe Gly Glu Pro Leu Asp His Asp Asp Glu Ser Ala Leu Val Ala Trp
 355 360 365

Leu Leu Ser
 370

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 <211> 1033
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G221

<400> 87
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 ggtgtttgga attctctcgc caaatctgca ggtctaaaac gaaccgggaa aagttgccgg 300
 ctccgggtggc tgaactatct ccgcggcggc gtacgacggg gaaacatcac tccagaagag 360
 caacttatca tcatggaact tcatgctaag tggggaaaca ggtggtcgaa aatcgccaaa 420
 catcttccag gaagaacgga caacgagatc aaaaatttct gtaggacaag aattcaaaaa 480
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 atggagactt attctccctac accgcacatca tatcaacata ccaatatgga attcaactat 660
 ggtaactatt cggccgcggc agtgcggca accgtggatt atccagtacc gatgaccgtt 720
 gatgatcaaa cccgtgaaaaa ctattggggc atggatgata tttggtcatc aatgcattta 780
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<210> 88
 <211> 226
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G221

<400> 88
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Glu Ala Glu Val Arg Lys Gly Pro Trp Thr Met Glu Glu Asp Leu Ile
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Leu Ile Asn Tyr Ile Ala Asn His Gly Asp Gly Val Trp Asn Ser Leu
 35 40 45

Ala Lys Ser Ala Gly Leu Lys Arg Thr Gly Lys Ser Cys Arg Leu Arg
 50 55 60

Trp Leu Asn Tyr Leu Arg Pro Asp Val Arg Arg Gly Asn Ile Thr Pro
 65 70 75 80

Glu Glu Gln Leu Ile Ile Met Glu Leu His Ala Lys Trp Gly Asn Arg
 85 90 95

Trp Ser Lys Ile Ala Lys His Leu Pro Gly Arg Thr Asp Asn Glu Ile
 100 105 110

Lys Asn Phe Cys Arg Thr Arg Ile Gln Lys Tyr Ile Lys Gln Ser Asp
 115 120 125

Val Thr Thr Thr Ser Ser Val Gly Ser His His Ser Ser Glu Ile Asn
 130 135 140

Asp Gln Ala Ala Ser Thr Ser Ser His Asn Val Phe Cys Thr Gln Asp
 145 150 155 160

Gln Ala Met Glu Thr Tyr Ser Pro Thr Pro Thr Ser Tyr Gln His Thr
 165 170 175

Asn Met Glu Phe Asn Tyr Gly Asn Tyr Ser Ala Ala Val Thr Ala
 180 185 190

Thr Val Asp Tyr Pro Val Pro Met Thr Val Asp Asp Gln Thr Gly Glu
 195 200 205

Asn Tyr Trp Gly Met Asp Asp Ile Trp Ser Ser Met His Leu Leu Asn
 210 215 220

Gly Asn
 225

<210> 89
 <211> 1952
 <212> DNA
 <213> *Arabidopsis thaliana*

<220>
 <223> G186

<220>
 <223> "n" bases at various positions throughout the
 sequence may be A, T, C, G, other or unknown

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 ctcactcttg attcatcttc tcttgatctt tttaaacccta atcgtatttc tcataagaat 180
 caccgacgtt tctcaaatacc ttggcgatg tctagaattt acgaagaaga tgatcagaag 240
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 gatcgtgaag atgaagattt ttcatctggc gttgctggag ataatgaccg tgaagttccc 360
 ggcgaagtgg atttcttctc cgacaagaaa tctagggttt gtcgtgaaga cgacgaagga 420
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 acaatggata atcaaaagct tagagaattt cttacacaag ttagcaacag ttacacttca 660
 cttcagatgc atcttggttc actaatgcag caacagcaac aacagaacaa taaggtata 720
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 aaccatccgt tgccgccagc cgcggtagcc atggcttcta ccaccacggc ggcggctaac 1260
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 cagcaatga cgaacttacc tccggaaatg ctacccatg taataggcca ggcattgtat 1560
 aaccaatcca agtttcggg gtcgcgttc tctggggct ctccctcgac ggcagcg 1620
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 ttccaactgg gttataggaa acagagaggta tatttcattt attcacattt gttctgtttc 1860
 gtacaaaat cccagtaaat atacaaaagc aaactataact caagttcata ttcgtaaaca 1920
 ctataatag tncgttnctt antaaaaaaa aa 1952

<210> 90

<211> 553

<212> PRT

<213> *Arabidopsis thaliana*

<220>

<223> G186

<400> 90

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Leu Leu Asn Pro Asn Arg Ile Ser His Lys Asn His Arg Arg Phe Ser
 20 25 30

Asn Pro Leu Ala Met Ser Arg Ile Asp Glu Glu Asp Asp Gln Lys Thr
 35 40 45

Arg Ile Ser Thr Asn Gly Ser Glu Phe Arg Phe Pro Val Ser Leu Ser
 50 55 60

Gly Ile Arg Asp Arg Glu Asp Glu Asp Phe Ser Ser Gly Val Ala Gly
 65 70 75 80

Asp Asn Asp Arg Glu Val Pro Gly Glu Val Asp Phe Phe Ser Asp Lys
 85 90 95

Lys Ser Arg Val Cys Arg Glu Asp Asp Glu Gly Phe Arg Val Lys Lys
 100 105 110

Glu Glu Gln Asp Asp Arg Thr Asp Val Asn Thr Gly Leu Asn Leu Arg
 115 120 125

Thr Thr Gly Asn Thr Lys Ser Asp Glu Ser Met Ile Asp Asp Gly Glu
 130 135 140

Ser Ser Glu Met Glu Asp Lys Arg Ala Lys Asn Glu Leu Val Lys Leu
 145 150 155 160

Gln Asp Glu Leu Lys Lys Met Thr Met Asp Asn Gln Lys Leu Arg Glu
 165 170 175

Leu Leu Thr Gln Val Ser Asn Ser Tyr Thr Ser Leu Gln Met His Leu
 180 185 190

Val Ser Leu Met Gln Gln Gln Gln Asn Asn Lys Val Ile Glu
 195 200 205

Ala Ala Glu Lys Pro Glu Glu Thr Ile Val Pro Arg Gln Phe Ile Asp
 210 215 220

Leu Gly Pro Thr Arg Ala Val Gly Glu Ala Glu Asp Val Ser Asn Ser
 225 230 235 240

Ser Ser Glu Asp Arg Thr Arg Ser Gly Gly Ser Ser Ala Ala Glu Arg
 245 250 255

Arg Ser Asn Gly Lys Arg Leu Gly Arg Glu Glu Ser Pro Glu Thr Glu
 260 265 270

Ser Asn Lys Ile Gln Lys Val Asn Ser Thr Thr Pro Thr Thr Phe Asp
 275 280 285

Gln Thr Ala Glu Ala Thr Met Arg Lys Ala Arg Val Ser Val Arg Ala
 290 295 300

Arg Ser Glu Ala Pro Met Ile Ser Asp Gly Cys Gln Trp Arg Lys Tyr
 305 310 315 320

Gly Gln Lys Met Ala Lys Gly Asn Pro Cys Pro Arg Ala Tyr Tyr Arg
 325 330 335

Cys Thr Met Ala Thr Gly Cys Pro Val Arg Lys Gln Val Gln Arg Cys
 340 345 350

Ala Glu Asp Arg Ser Ile Leu Ile Thr Thr Tyr Glu Gly Asn His Asn
 355 360 365

His Pro Leu Pro Pro Ala Ala Val Ala Met Ala Ser Thr Thr Thr Ala
 370 375 380

Ala Ala Asn Met Leu Leu Ser Gly Ser Met Ser Ser His Asp Gly Met
 385 390 395 400

Met Asn Pro Thr Asn Leu Leu Ala Arg Ala Val Leu Pro Cys Ser Thr
 405 410 415

Ser Met Ala Thr Ile Ser Ala Ser Ala Pro Phe Pro Thr Val Thr Leu
 420 425 430

Asp Leu Thr His Ser Pro Pro Pro Asn Gly Ser Asn Pro Ser Ser
 435 440 445

Ser Ala Ala Thr Asn Asn His Asn Ser Leu Met Gln Arg Pro Gln
 450 455 460

Gln Gln Gln Gln Gln Met Thr Asn Leu Pro Pro Gly Met Leu Pro His
 465 470 475 480

Val Ile Gly Gln Ala Leu Tyr Asn Gln Ser Lys Phe Ser Gly Leu Gln
 485 490 495

Phe Ser Gly Gly Ser Pro Ser Thr Ala Ala Phe Ser Gln Ser His Ala
 500 505 510

Val Ala Asp Thr Ile Thr Ala Leu Thr Ala Asp Pro Asn Phe Thr Ala
 515 520 525

Ala Leu Ala Ala Val Ile Ser Ser Met Ile Asn Gly Thr Asn His His
 530 535 540

Asp Gly Glu Gly Asn Asn Lys Asn Gln
 545 550

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 <211> 1554
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G562

<400> 91
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 tcccatggga tcactgcctc aaggtcaaaa ggatccacct ttaacaactc cggggacgct 480
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 aggagacaat gattcttaact ctacaagcaa attccatcaa ctgctcgata cgaaggctcg 1260

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 tatagttagag agagagagag agagagaggt gtgatgatta ttgatctata aattttcgga 1440
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<210> 92
 <211> 382
 <212> PRT
 <213> *Arabidopsis thaliana*

<220>
 <223> G562

<400> 92
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Pro Ser Ser Pro Pro Val Asp Gln Thr Asn Val His Val Tyr Pro Asp
 20 25 30

Trp Ala Ala Met Gln Ala Tyr Tyr Gly Pro Arg Val Ala Met Pro Pro
 35 40 45

Tyr Tyr Asn Ser Ala Met Ala Ala Ser Gly His Pro Pro Pro Pro Tyr
 50 55 60

Met Trp Asn Pro Gln His Met Met Ser Pro Ser Gly Ala Pro Tyr Ala
 65 70 75 80

Ala Val Tyr Pro His Gly Gly Val Tyr Ala His Pro Gly Ile Pro
 85 90 95

Met Gly Ser Leu Pro Gln Gly Gln Lys Asp Pro Pro Leu Thr Thr Pro
 100 105 110

Gly Thr Leu Leu Ser Ile Asp Thr Pro Thr Lys Ser Thr Gly Asn Thr
 115 120 125

Asp Asn Gly Leu Met Lys Lys Leu Lys Glu Phe Asp Gly Leu Ala Met
 130 135 140

Ser Leu Gly Asn Gly Asn Pro Glu Asn Gly Ala Asp Glu His Lys Arg
 145 150 155 160

Ser Arg Asn Ser Ser Glu Thr Asp Gly Ser Thr Asp Gly Ser Asp Gly
 165 170 175

Asn Thr Thr Gly Ala Asp Glu Pro Lys Leu Lys Arg Ser Arg Glu Gly
 180 185 190

Thr Pro Thr Lys Asp Gly Lys Gln Leu Val Gln Ala Ser Ser Phe His
 195 200 205

Ser Val Ser Pro Ser Ser Gly Asp Thr Gly Val Lys Leu Ile Gln Gly
 210 215 220

Ser Gly Ala Ile Leu Ser Pro Gly Val Ser Ala Asn Ser Asn Pro Phe
 225 230 235 240

Met Ser Gln Ser Leu Ala Met Val Pro Pro Glu Thr Trp Leu Gln Asn
 245 250 255

Glu Arg Glu Leu Lys Arg Glu Arg Arg Lys Gln Ser Asn Arg Glu Ser
 260 265 270

Ala Arg Arg Ser Arg Leu Arg Lys Gln Ala Glu Thr Glu Glu Leu Ala
 275 280 285

Arg Lys Val Glu Ala Leu Thr Ala Glu Asn Met Ala Leu Arg Ser Glu
 290 295 300

Leu Asn Gln Leu Asn Glu Lys Ser Asp Lys Leu Arg Gly Ala Asn Ala
 305 310 315 320

Thr Leu Leu Asp Lys Leu Lys Cys Ser Glu Pro Glu Lys Arg Val Pro
 325 330 335

Ala Asn Met Leu Ser Arg Val Lys Asn Ser Gly Ala Gly Asp Lys Asn
 340 345 350

Lys Asn Gln Gly Asp Asn Asp Ser Asn Ser Thr Ser Lys Phe His Gln
 355 360 365

Leu Leu Asp Thr Lys Pro Arg Ala Lys Ala Val Ala Ala Gly
 370 375 380

<210> 93

<211> 918

<212> DNA

<213> *Arabidopsis thaliana*

<220>

<223> G255

<400> 93

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<210> 94
 <211> 269
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G255

<400> 94
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 Gly Glu Gly Cys Trp Arg Ser Leu Pro Arg Ala Ala Gly Leu Leu Arg
 35 40 45
 Cys Gly Lys Ser Cys Arg Leu Arg Trp Ile Asn Tyr Leu Arg Pro Asp
 50 55 60
 Leu Lys Arg Gly Asn Phe Thr His Asp Glu Asp Glu Leu Ile Ile Lys
 65 70 75 80
 Leu His Ser Leu Leu Gly Asn Lys Trp Ser Leu Ile Ala Ala Arg Leu
 85 90 95
 Pro Gly Arg Thr Asp Asn Glu Ile Lys Asn Tyr Trp Asn Thr His Ile
 100 105 110
 Lys Arg Lys Leu Leu Ser Lys Gly Ile Asp Pro Ala Thr His Arg Gly
 115 120 125
 Ile Asn Glu Ala Lys Ile Ser Asp Leu Lys Lys Thr Lys Asp Gln Ile
 130 135 140
 Val Lys Asp Val Ser Phe Val Thr Lys Phe Glu Glu Thr Asp Lys Ser
 145 150 155 160
 Gly Asp Gln Lys Gln Asn Lys Tyr Ile Arg Asn Gly Leu Val Cys Lys
 165 170 175
 Glu Glu Arg Val Val Val Glu Glu Lys Ile Gly Pro Asp Leu Asn Leu
 180 185 190
 Glu Leu Arg Ile Ser Pro Pro Trp Gln Asn Gln Arg Glu Ile Ser Thr
 195 200 205
 Cys Thr Ala Ser Arg Phe Tyr Met Glu Asn Asp Met Glu Cys Ser Ser
 210 215 220
 Glu Thr Val Lys Cys Gln Thr Glu Asn Ser Ser Ser Ile Ser Tyr Ser
 225 230 235 240
 Ser Ile Asp Ile Ser Ser Ser Asn Val Gly Tyr Asp Phe Leu Gly Leu
 245 250 255

Lys Thr Arg Ile Leu Asp Phe Arg Ser Leu Glu Met Lys
 260 265

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 <211> 759
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G3

<400> 95
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 aggaggaagt ggggcaagtg ggtggctgag attcgtgagc ctaacaaacg ctcacggctt 180
 tggcttggct cttacacaac cgatatacgcc gccgctagag cctacgacgt ggccgtcttc 240
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 catagatact ggaaaatata ggtatgtata cattataaaa ttatctttagt tatcaaagaa 600
 tttttagat tctgattagc tttttggttt tggttttgat aagaactctg attagttgtc 660
 cggagacaaa accqqctaag agcaatccat gagaagctag cgagtgtttt ttagttcaag 720
 ttgtaatata aatgcataatt aattcttttag taattttgt 759

<210> 96
 <211> 153
 <212> PRT
 <213> Arabidopsis thaliana

<220>
 <223> G3

<400> 96
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 20 25 30

Gly Ile Arg Arg Arg Lys Trp Gly Lys Trp Val Ala Glu Ile Arg Glu
 35 40 45

Pro Asn Lys Arg Ser Arg Leu Trp Leu Gly Ser Tyr Thr Thr Asp Ile
 50 55 60

Ala Ala Ala Arg Ala Tyr Asp Val Ala Val Phe Tyr Leu Arg Gly Pro
 65 70 75 80

Ser Ala Arg Leu Asn Phe Pro Asp Leu Leu Leu Gln Glu Glu Asp His
 85 90 95

Leu Ser Ala Ala Thr Thr Ala Asp Met Pro Ala Ala Leu Ile Arg Glu
 100 105 110

Lys Ala Ala Glu Val Gly Ala Arg Val Asp Ala Leu Leu Ala Ser Ala
 115 120 125

Ala Pro Ser Met Ala His Ser Thr Pro Pro Val Ile Lys Pro Asp Leu
 130 135 140

Asn Gln Ile Pro Glu Ser Gly Asp Ile
 145 150

<210> 97
 <211> 965
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G713

<400> 97
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 aagaagaaga tgaagaagag caataaccaa aagaggtta acgaggaaca gatcaagtca 180
 cttgagctta tatttgagtc tgagacgagg cttgagccga ggaagaaggt tcaggttagct 240
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 35 40 45

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 50 55 60

Gly Leu Gln Pro Arg Gln Met Thr Ile Trp Phe Gln Asn Lys Arg Ala
 65 70 75 80

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 85 90 95

Asn Tyr Asn Asn Leu Ala Ser Gln Phe Glu Ile Met Lys Lys Glu Lys
 100 105 110

Gln Ser Leu Val Ser Glu Leu Gln Arg Leu Asn Glu Glu Met Gln Arg
 115 120 125

Pro Lys Glu Glu Lys His His Glu Cys Cys Gly Asp Gln Gly Leu Ala
 130 135 140

Leu Ser Ser Ser Thr Glu Ser His Asn Gly Lys Ser Glu Pro Glu Gly
 145 150 155 160

Arg Leu Asp Gln Gly Ser Val Leu Cys Asn Asp Gly Asp Tyr Asn Asn
 165 170 175

Asn Ile Lys Thr Glu Tyr Phe Arg Val Gln Gly Glu Thr Asp His Glu
 180 185 190

Leu Met Asn Ile Val Glu Lys Ala Asp Asp Ser Cys Leu Thr Ser Ser
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 Gly Phe Ser His Asn Gln Met Met Thr Tyr Thr Leu Cys Lys Val
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 Met Phe Asn Gly Gly Met Arg Glu Lys Ser Ser Ser Ser Pro Ser Ser
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 Gln Leu Val Asn Asn Ser Glu Gly Ser Ser Leu His Arg Glu Asp Pro
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 195 200 205

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Gln Ile Glu Asp Ala Ile Pro Ile Glu Glu Trp Glu Thr Trp Leu Asn
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Asp Ile Asp Asp Ala Lys Glu Lys Ser Ile Met Phe Met His Asp Asn
245 250 255

Arg Ser Asp Tyr Arg Pro Pro Asn Ser Leu Thr Gly Val Phe Ser Asp
260 265 270

Asp Val Ser Ser Asp Asp Asn Asp Ser Asp Leu Leu Thr Pro Lys Thr
 275 280 285

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His Arg Ile Asp Gln Ile Lys Asp Leu Gln Glu Ser Pro Thr Ser Thr
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 Arg Asn Cys Ser Gly Ile Ala Ala Arg Ala Cys Gly Leu Val Ser Leu
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 Glu Pro Met Lys Val Ala Glu Ile Leu Lys Asp Arg Pro Ser Trp Phe
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 325 330 335
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 Asp His Gln Ser Ala Ser Arg Thr Arg Asp Leu Ala Ser Ser Leu Asp
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 Gly Ser Thr Lys Thr Asp Ser Glu Thr Asn Ser Arg Leu Val Leu Thr
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 660 665 670
 Val Ala Leu Ala Ile Thr Pro Arg Pro Gly Ser Met Gln Leu Pro Thr
 675 680 685
 Ser Pro Glu Ala Leu Thr Leu Val Arg Trp Ile Thr Arg Ser Tyr Ser
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 Glu Val Glu Lys Arg Ala Ile Glu Arg Lys Asn Met Asp Leu Glu Met
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 Arg Leu Ala Asp Met Glu Ala Lys Tyr Tyr Leu Leu Gln Gln Glu Leu
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 Lys Arg Ala Ser Gly Tyr Asn Lys Thr Asn Phe Leu Ser Tyr Ser Asp
 165 170 175
 Ser Ser Thr Pro Asp Ile Ser Glu Asp Ser Gln Leu Ser Pro Leu Thr
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 Phe Ser Lys Gln Leu Phe Asn Ala Gln Asp Glu Leu Cys Arg Pro Ile
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 Ser Pro Gln Ser Ile Gly Pro Leu Thr Ser Arg Thr Val Asp Pro Ser
 210 215 220
 Thr Leu Ser Pro Lys Ser Leu Ser Ser Pro Asp Ser Ser Asn Ser Asn
 225 230 235 240

Ser Ser Asp Met Thr Gln His Pro Ala Val Val Leu Cys Asp Leu Gln
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 Cys Gln Ser Glu Leu Gly Gln Pro Trp Met Asn Ser Thr Tyr Leu Ser
 260 265 270
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 100 105 110

Asn Thr Glu Val Ser Glu Thr Met Ser Asn Leu Gln Ile Thr Ser Thr
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 130 135 140

Val Thr Phe Pro Val Arg Pro Gly Arg Gly Thr Leu Gly Lys Lys Val
 145 150 155 160

Met Val Arg Ala Asn His Phe Leu Val Gln Val Ala Asp Arg Asp Leu
 165 170 175

Tyr His Tyr Asp Val Ser Ile Asn Pro Glu Val Ile Ser Lys Thr Val
 180 185 190

Asn Arg Asn Val Met Lys Leu Leu Val Lys Asn Tyr Lys Asp Ser His
 195 200 205

Leu Gly Gly Lys Ser Pro Ala Tyr Asp Gly Arg Lys Ser Leu Tyr Thr
 210 215 220

Ala Gly Pro Leu Pro Phe Asp Ser Lys Glu Phe Val Val Asn Leu Ala
 225 230 235 240

Glu Lys Arg Ala Asp Gly Ser Ser Gly Lys Asp Arg Pro Phe Lys Val
 245 250 255

Ala Val Lys Asn Val Thr Ser Thr Asp Leu Tyr Gln Leu Gln Gln Phe
 260 265 270

Leu Asp Arg Lys Gln Arg Glu Ala Pro Tyr Asp Thr Ile Gln Val Leu
 275 280 285

Asp Val Val Leu Arg Asp Lys Pro Ser Asn Asp Tyr Val Ser Val Gly
 290 295 300

Arg Ser Phe Phe His Thr Ser Leu Gly Lys Asp Ala Arg Asp Gly Arg
 305 310 315 320

Gly Glu Leu Gly Asp Gly Ile Glu Tyr Trp Arg Gly Tyr Phe Gln Ser
 325 330 335

Leu Arg Leu Thr Gln Met Gly Leu Ser Leu Asn Ile Asp Val Ser Ala
 340 345 350

Arg Ser Phe Tyr Glu Pro Ile Val Val Thr Asp Phe Ile Ser Lys Phe
 355 360 365

Leu Asn Ile Arg Asp Leu Asn Arg Pro Leu Arg Asp Ser Asp Arg Leu
 370 375 380

 Lys Val Lys Lys Val Leu Arg Thr Leu Lys Val Lys Leu Leu His Trp
 385 390 395 400

 Asn Gly Thr Lys Ser Ala Lys Ile Ser Gly Ile Ser Ser Leu Pro Ile
 405 410 415

 Arg Glu Leu Arg Phe Thr Leu Glu Asp Lys Ser Glu Lys Thr Val Val
 420 425 430

 Gln Tyr Phe Ala Glu Lys Tyr Asn Tyr Arg Val Lys Tyr Gln Ala Leu
 435 440 445

 Pro Ala Ile Gln Thr Gly Ser Asp Thr Arg Pro Val Tyr Leu Pro Met
 450 455 460

 Glu Leu Cys Gln Ile Asp Glu Gly Gln Arg Tyr Thr Lys Arg Leu Asn
 465 470 475 480

 Glu Lys Gln Val Thr Ala Leu Leu Lys Ala Thr Cys Gln Arg Pro Pro
 485 490 495

 Asp Arg Glu Asn Ser Ile Lys Asn Leu Val Val Lys Asn Asn Tyr Asn
 500 505 510

 Asp Asp Leu Ser Lys Glu Phe Gly Met Ser Val Thr Thr Gln Leu Ala
 515 520 525

 Ser Ile Glu Ala Arg Val Leu Pro Pro Pro Met Leu Lys Tyr His Asp
 530 535 540

 Ser Gly Lys Glu Lys Met Val Asn Pro Arg Leu Gly Gln Trp Asn Met
 545 550 555 560

 Ile Asp Lys Lys Met Val Asn Gly Ala Lys Val Thr Ser Trp Thr Cys
 565 570 575

 Glu Phe Lys Pro Gln Pro Ala Ile Pro Phe Ile Ser Cys Pro Pro Glu
 580 585 590

 His Ile Glu Glu Ala Leu Leu Asp Ile His Lys Arg Ala Pro Gly Leu
 595 600 605

 Gln Leu Leu Ile Val Ile Leu Pro Asp Val Thr Gly Ser Tyr Gly Lys
 610 615 620

 Ile Lys Arg Ile Cys Glu Thr Glu Leu Gly Ile Val Ser Gln Cys Cys
 625 630 635 640

 Gln Pro Arg Gln Val Asn Lys Leu Asn Lys Gln Tyr Met Glu Asn Val
 645 650 655

 Ala Leu Lys Ile Asn Val Lys Thr Gly Gly Arg Asn Thr Val Leu Asn
 660 665 670

Asp Ala Ile Arg Arg Asn Ile Pro Leu Ile Thr Asp Arg Pro Thr Ile
 675 680 685
 Ile Met Gly Ala Asp Val Thr His Pro Gln Pro Gly Glu Asp Ser Ser
 690 695 700
 Pro Ser Ile Ala Ala Val Val Ala Ser Met Asp Trp Pro Glu Ile Asn
 705 710 715 720
 Lys Tyr Arg Gly Leu Val Ser Ala Gln Ala His Arg Glu Glu Ile Ile
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 Gln Asp Leu Tyr Lys Leu Val Gln Asp Pro Gln Arg Gly Leu Val His
 740 745 750
 Ser Gly Leu Ile Arg Glu His Phe Ile Ala Phe Arg Arg Ala Thr Gly
 755 760 765
 Gln Ile Pro Gln Arg Ile Ile Phe Tyr Arg Asp Gly Val Ser Glu Gly
 770 775 780
 Gln Phe Ser Gln Val Leu Leu His Glu Met Thr Ala Ile Arg Lys Ala
 785 790 795 800
 Cys Asn Ser Leu Gln Glu Asn Tyr Val Pro Arg Val Thr Phe Val Ile
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 Val Gln Lys Arg His His Thr Arg Leu Phe Pro Glu Gln His Gly Asn
 820 825 830
 Arg Asp Met Thr Asp Lys Ser Gly Asn Ile Gln Pro Gly Thr Val Val
 835 840 845
 Asp Thr Lys Ile Cys His Pro Asn Glu Phe Asp Phe Tyr Leu Asn Ser
 850 855 860
 His Ala Gly Ile Gln Gly Thr Ser Arg Pro Ala His Tyr His Val Leu
 865 870 875 880
 Leu Asp Glu Asn Gly Phe Thr Ala Asp Gln Leu Gln Met Leu Thr Asn
 885 890 895
 Asn Leu Cys Tyr Thr Tyr Ala Arg Cys Thr Lys Ser Val Ser Ile Val
 900 905 910
 Pro Pro Ala Tyr Tyr Ala His Leu Ala Ala Phe Arg Ala Arg Tyr Tyr
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 aatgctttt gatctcagcc gttgactaca gagAGCCTT cggcgaAGC ttctgattca 180
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 cctcatgtt cttcgcgt gcaatcAGCT tgcttcgagt ttggatttgc tcagccaATG 360
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 35 40 45
 Gly Val Val Asp Lys Gln Thr Ser Thr Thr Leu Phe Thr Phe Ser Pro
 50 55 60
 Gly Gly Glu Lys Ser Ser Arg Asp Val Pro Lys Pro His Val Ala Phe
 65 70 75 80

Ala Met Gln Ser Ala Cys Phe Glu Phe Gly Phe Ala Gln Pro Met Met
 85 90 95

Tyr Thr Lys His Pro His Val Glu Gln Tyr Tyr Gly Val Val Ser Ala
 100 105 110

Tyr Gly Ser Gln Arg Ser Ser Gly Arg Val Met Ile Pro Leu Lys Met
 115 120 125

Glu Thr Glu Glu Asp Gly Thr Ile Tyr Val Asn Ser Lys Gln Tyr His
 130 135 140

Gly Ile Ile Arg Arg Arg Gln Ser Arg Ala Lys Ala Glu Lys Leu Ser
 145 150 155 160

Arg Cys Arg Lys Pro Tyr Met His His Ser Arg His Leu His Ala Met
 165 170 175

Arg Arg Pro Arg Gly Ser Gly Gly Arg Phe Leu Asn Thr Lys Thr Ala
 180 185 190

Asp Ala Ala Lys Gln Ser Lys Pro Ser Asn Ser Gln Ser Ser Glu Val
 195 200 205

Phe His Pro Glu Asn Glu Thr Ile Asn Ser Ser Arg Glu Ala Asn Glu
 210 215 220

Ser Asn Leu Ser Asp Ser Ala Val Thr Ser Met Asp Tyr Phe Leu Ser
 225 230 235 240

Ser Ser Ala Tyr Ser Pro Gly Gly Met Val Met Pro Ile Lys Trp Asn
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Ala Ala Ala Met Asp Ile Gly Cys Cys Lys Leu Asn Ile
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<211> 1423

<212> DNA

<213> *Arabidopsis thaliana*

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<223> G1650

<400> 109

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 atatctcgag gcaaagaggg aacatatttc ttggcggtgt tgaagctgtt ccgtcgaaact 600
 cgaccctgtt gtcttcagcc actgaatcaa taccagcgac tcacggcacc gagagtcgag 660
 caacagtac tggcgagta tctcgactt ttgcagttcc tggcttggtt ccgaggggaa 720

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 ctccatgct ggatgatgtt atagagtagc tgaaatctct acagagccaa atacaggatg 1080
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 35 40 45
 Asp Pro Pro Leu Ile Leu Arg Gly Ser Gly Ser Gly Asp Gly Glu Gly
 50 55 60
 Asn Gly Pro Leu Pro Gln Pro Pro Pro Pro Leu Tyr His Gln Gln Ser
 65 70 75 80
 Leu Phe Ile Gln Glu Asp Glu Met Ala Ser Trp Leu His Gln Pro Asn
 85 90 95
 Arg Gln Asp Tyr Leu Tyr Ser Gln Leu Leu Tyr Ser Gly Val Ala Ser
 100 105 110
 Thr His Pro Gln Ser Leu Ala Ser Leu Glu Pro Pro Pro Pro Arg
 115 120 125
 Ala Gln Tyr Ile Leu Ala Ala Asp Arg Pro Thr Gly His Ile Leu Ala
 130 135 140
 Glu Arg Arg Ala Glu Asn Phe Met Asn Ile Ser Arg Gln Arg Gly Asn
 145 150 155 160
 Ile Phe Leu Gly Gly Val Glu Ala Val Pro Ser Asn Ser Thr Leu Leu
 165 170 175
 Ser Ser Ala Thr Glu Ser Ile Pro Ala Thr His Gly Thr Glu Ser Arg
 180 185 190

Ala Thr Val Thr Gly Gly Val Ser Arg Thr Phe Ala Val Pro Gly Leu
 195 200 205
 Gly Pro Arg Gly Lys Ala Val Ala Ile Glu Thr Ala Gly Thr Gln Ser
 210 215 220
 Trp Gly Leu Cys Lys Ala Glu Thr Glu Pro Val Gln Arg Gln Pro Ala
 225 230 235 240
 Thr Glu Thr Asp Ile Thr Asp Glu Arg Lys Arg Lys Thr Arg Glu Glu
 245 250 255
 Thr Asn Val Glu Asn Gln Gly Thr Glu Glu Ala Arg Asp Ser Thr Ser
 260 265 270
 Ser Lys Arg Ser Arg Ala Ala Ile Met His Lys Leu Ser Glu Arg Arg
 275 280 285
 Arg Arg Gln Lys Ile Asn Glu Met Met Lys Ala Leu Gln Glu Leu Leu
 290 295 300
 Pro Arg Cys Thr Lys Thr Asp Arg Ser Ser Met Leu Asp Asp Val Ile
 305 310 315 320
 Glu Tyr Val Lys Ser Leu Gln Ser Gln Ile Gln Asp Val Leu Asn Gly
 325 330 335
 Thr Cys Tyr Asp Ser Thr Asp Asp Val Cys Gly Glu Tyr Thr Thr Thr
 340 345 350
 Val His Ala Pro His Gly His Gly Tyr Glu Ser Ala Ser Cys Ile His
 355 360 365
 Thr Phe Pro
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 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G241

<400> 111
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 cctaagcaag ctggtatgta aaaattataa tatcaaattt ctttaattttg atcaaatttc 180
 ttacattaat tattggtaat tattatttac aggtctttt 240
 acttaggtgg atgaactatt taaagcctga tattaaacgt ggcaatttca ccaaagaaga 300
 ggaagatgct atcatcagct tacaccaaattt acttggcaat aggtatttta cttcaacata 360
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 <212> PRT
 <213> Arabidopsis thaliana

<220>
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 Gly His Ser Asn Trp Arg Ala Leu Pro Lys Gln Ala Gly Leu Leu Arg
 35 40 45
 Cys Gly Lys Ser Cys Arg Leu Arg Trp Met Asn Tyr Leu Lys Pro Asp
 50 55 60
 Ile Lys Arg Gly Asn Phe Thr Lys Glu Glu Asp Ala Ile Ile Ser
 65 70 80
 Leu His Gln Ile Leu Gly Asn Arg Trp Ser Ala Ile Ala Ala Lys Leu
 85 90 95
 Pro Gly Arg Thr Asp Asn Glu Ile Lys Asn Val Trp His Thr His Leu
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 Lys Lys Arg Leu Glu Asp Leu Ser Thr Ser
 115 120

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 <211> 375
 <212> DNA
 <213> Arabidopsis thaliana

<220>
 <223> G348

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 ggtggaccaa ctggccctaa ggtcttttct tctaccctta attactatata tcataacttt 180
 gtttgatctt aagataattc atcaagtgtt cttaagttgt ttattttgat ttgtgggtggg 240
 atttgcagtc acttgcaat gcatgtggaa ttagacacag aaaacagaga cgatcagagt 300
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<211> 1332
<212> DNA
<213> *Arabidopsis thaliana*

<220>
<223> G521

<400> 115
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<212> DNA

<213> Arabidopsis thaliana

<220>

<223> G1274

<400> 116

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<211> 2352

<212> DNA

<213> Arabidopsis thaliana

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<223> G182

<400> 117

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